Available online on 15.02.2026 at <http://jddtonline.info>

Journal of Drug Delivery and Therapeutics

Open Access to Pharmaceutical and Medical Research

Copyright © 2026 The Author(s): This is an open-access article distributed under the terms of the CC BY-NC 4.0 which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited



Open Access Full Text Article

Review Article

Tailored Liposomal Nanocarriers for Precision Breast Cancer Therapy

Ajinkya Shrirang Holkar , Vinod Jagannathrao Mokale *

University Department of Pharmaceutical Sciences, MGM University, Chh. Sambhajinagar, MS, India.

Article Info:

Abstract



Article History:

Received 17 Nov 2025
Reviewed 09 Jan 2026
Accepted 31 Jan 2026
Published 15 Feb 2026

Cite this article as:

Holkar AS, Mokale VJ, Tailored Liposomal Nanocarriers for Precision Breast Cancer Therapy, Journal of Drug Delivery and Therapeutics. 2026; 16(2):71-91 DOI: <http://dx.doi.org/10.22270/jddt.v16i2.7544>

For Correspondence:

Mokale Vinod Jagannathrao, University Department of Pharmaceutical Sciences, MGM University, Chh. Sambhajinagar, MS, India.

Breast cancer remains one of the most frequently diagnosed malignancies worldwide and continues to represent a major public health concern despite substantial advances in diagnostic and therapeutic strategies. Conventional chemotherapy, although effective in many cases, is often associated with non-specific drug distribution, severe systemic toxicity, limited bioavailability, and the development of multidrug resistance. These limitations significantly compromise therapeutic outcomes and patient quality of life. Liposome-based drug delivery systems have developed as a highly promising nanotechnological approach to address these challenges by improving drug solubility, stability, circulation time, and tumor selectivity. Through rational design and surface functionalization, liposomes can be engineered to achieve passive and active targeting, controlled drug release, and reduced off-target effects. This review provides an expanded and original discussion of liposomal nanocarriers developed for targeted breast cancer therapy, covering formulation strategies, classification, targeting mechanisms, stimuli-responsive systems, clinical applications, current challenges, and future perspectives.

Keywords: Liposomal nanocarriers; Breast cancer therapy; Targeted drug delivery; Nanomedicine; Stimuli-responsive systems; Precision oncology

Introduction:

Cancer continues to pose a significant global public health challenge and remains one of the foremost causes of death worldwide. Recent epidemiological data from the GLOBOCAN 2021 database indicate that breast cancer has overtaken lung cancer as the most commonly diagnosed malignancy among women. In that year alone, an estimated 2.3 million new breast cancer cases were reported globally, accompanied by approximately 680,000 disease-related deaths across 185 countries¹. While the incidence of breast cancer varies markedly with ethnicity, socioeconomic conditions, geographical location, and healthcare accessibility, long-term epidemiological trends demonstrate a consistent rise in both incidence and mortality over the last 25 years⁵.

Breast cancer arises from the malignant transformation of epithelial cells lining the ducts or lobules of the mammary gland. The disease is characterized by substantial biological and clinical heterogeneity, encompassing a wide spectrum of histopathological subtypes and molecular alterations, which complicates disease classification and therapeutic management. On the basis of histological origin, breast cancers are broadly divided into ductal and lobular carcinomas. Ductal lesions may occur in either non-invasive or invasive forms (Fig. 1). Non-invasive tumors,

collectively referred to as carcinoma in situ, remain confined to the ductal or lobular structures without breaching the surrounding tissue. Among these, ductal carcinoma in situ (DCIS) accounts for nearly 90% of cases, whereas lobular carcinoma in situ (LCIS) is comparatively rare⁶. These non-invasive lesions generally exhibit favorable clinical outcomes and are biologically distinct from invasive disease.

In contrast, invasive breast cancer is defined by the capacity of malignant cells to penetrate the basement membrane and invade adjacent stromal and adipose tissues. Invasive ductal carcinoma represents approximately 80% of all invasive breast cancer cases and comprises multiple histological variants, including medullary, mucinous, tubular, and papillary carcinomas. Additional invasive subtypes include invasive lobular carcinoma, inflammatory breast cancer, and Paget's disease of the nipple and breast⁷. Less common malignancies, such as phyllodes tumors derived from stromal components and breast carcinomas exhibiting neuroendocrine differentiation, are also recognized within the invasive breast cancer spectrum⁸.

Progress in molecular biology and genomics has substantially refined breast cancer classification. Investigations into tumor etiology, gene expression patterns, and clinical behavior have revealed distinct molecular subtypes within both ductal and lobular

carcinomas⁹. These molecular categories are primarily defined by the expression status of estrogen receptors (ER), progesterone receptors (PR), and human epidermal growth factor receptor 2 (HER2), which serve as key biomarkers for prognostic assessment and therapeutic decision-making^{4,10}. Clinically relevant subtypes include Luminal A (ER+ and/or PR+, HER2-), Luminal B (ER+ and/or PR+, HER2+), and basal-like tumors lacking ER, PR, and HER2 expression¹¹. Large-scale transcriptomic analyses have further classified breast cancer into five intrinsic subtypes: Luminal A, Luminal B, HER2-enriched, basal-like and claudin-low^{12,13}.

Genetic predisposition is a critical contributor to breast cancer susceptibility. Inherited mutations in tumor suppressor genes, particularly BRCA1 and BRCA2, confer a markedly increased lifetime risk of developing breast cancer, estimated at approximately 70% and 60%, respectively^{6,14}. Among the molecular subtypes, basal-like breast cancer—commonly referred to as triple-negative breast cancer (TNBC)—accounts for approximately 15–20% of all cases. TNBC is associated with aggressive clinical behavior, high histological grade, absence of established molecular targets, and poor patient prognosis^{10,15,16}.

Advances in the understanding of breast cancer biology have facilitated the development of sophisticated therapeutic strategies and targeted drug delivery approaches. Contemporary treatment regimens are highly individualized and are determined by tumor size, histopathological grade, molecular subtype, disease stage, proliferative activity, and lymph node involvement. Comprehensive guidelines for the diagnosis and management of breast cancer have been extensively reviewed by Moo *et al.*¹⁷. Standard care typically employs a multimodal strategy, integrating surgery, chemotherapy, endocrine therapy, radiotherapy, and immunotherapy. Notably, higher rates of disease recurrence are observed in basal-like and Luminal B tumors compared with Luminal A breast cancers¹⁸.

Cytotoxic chemotherapy remains a cornerstone of breast cancer treatment and may be administered in either the neoadjuvant or adjuvant setting. Frequently used chemotherapeutic agents include anthracyclines such as doxorubicin and epirubicin, taxanes including paclitaxel and docetaxel, platinum-based drugs such as cisplatin and carboplatin, and antimetabolites like

gemcitabine and fluorouracil¹⁹. In hormone receptor-positive breast cancer, chemotherapy is often combined with endocrine therapies, including tamoxifen, fulvestrant, and aromatase inhibitors such as letrozole, to enhance therapeutic efficacy^{20–22}. Immunotherapeutic interventions—encompassing monoclonal antibodies, cytokine therapies, cancer vaccines, and adoptive cell transfer—are predominantly utilized in HER2-positive breast cancer²³.

Despite its widespread use, chemotherapy is associated with several inherent limitations. Poor tumor selectivity frequently leads to systemic toxicity and a broad spectrum of adverse effects^{24,25}. Moreover, chemotherapy is occasionally prescribed in clinical contexts where less intensive treatment options may suffice, resulting in avoidable physical, psychological, and financial burdens for patients^{26,27}. A further major obstacle is the emergence of multidrug resistance (MDR), which significantly contributes to therapeutic failure and disease relapse. MDR is commonly driven by the overexpression of ATP-binding cassette transporters, including P-glycoprotein, which actively expel anticancer agents from tumor cells and reduce intracellular drug concentrations^{28–31}.

Metastatic spread to distant organs remains the leading cause of breast cancer-associated mortality. Metastatic breast cancer is generally regarded as incurable and is characterized by poor survival outcomes and limited treatment options^{37,38}. In response to these challenges, nanotechnology-based drug delivery platforms—such as liposomes, polymeric micelles, dendrimers, and lipid nanocapsules—have gained considerable attention as innovative tools for cancer therapy⁴⁰. These nanoscale systems allow for controlled and targeted drug delivery through surface modification with ligands including peptides, antibodies, and proteins, thereby improving pharmacokinetics, therapeutic efficacy, and safety profiles^{41–45}.

First-generation nanomedicines, such as PEGylated liposomal doxorubicin (Doxil®/Caelyx®) and albumin-bound paclitaxel (Abraxane®), primarily rely on the enhanced permeability and retention (EPR) effect and represent significant milestones in the clinical application of nanotechnology for cancer treatment^{46,47}. Among the various nanocarrier systems, liposomal drug delivery platforms have demonstrated particular promise due to their biocompatibility, structural versatility, and proven success in clinical settings.

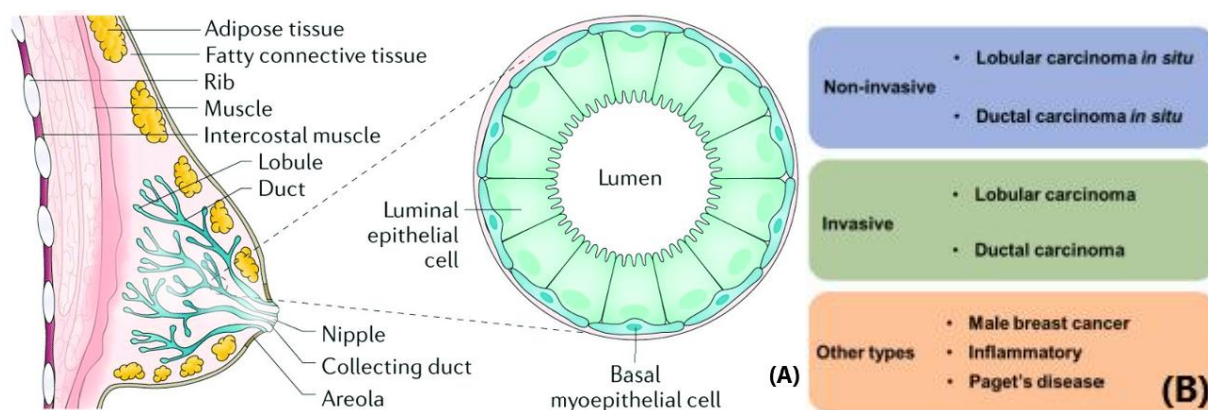


Figure 1: (A) Diagram showing ductal & lobular epithelium of breast, Ref ⁶
(B) Breast cancer types

Liposomes are vesicular nanostructures that were first described in 1965 and are characterized by their spherical architecture composed of one or more concentric lipid bilayers enclosing an aqueous interior ⁴⁸⁻⁵⁰. These vesicles are formed through the spontaneous self-assembly of amphiphilic lipids, most commonly phospholipids, when dispersed in an aqueous environment. Owing to their amphiphilic nature, phospholipids arrange themselves into bilayer structures that closely resemble biological cell membranes ^{51,52}. This biomimetic characteristic endows liposomes with admirable biocompatibility and biodegradability, making them highly attractive candidates for pharmaceutical and biomedical applications.

One of the most notable advantages of liposomes is their remarkable versatility in drug encapsulation. Hydrophilic compounds can be entrapped within the aqueous core, while lipophilic molecules are included into the hydrophobic region of the lipid bilayer. Amphiphilic agents may localize at the lipid-water interface, allowing liposomes to accommodate a broad spectrum of therapeutic molecules ^{53,54}. Furthermore, the physicochemical properties of liposomes can be finely tuned by modifying their lipid composition. Parameters such as bilayer fluidity, surface charge, hydration behaviour, and preparation method can be precisely controlled, enabling the design of liposomal systems tailored to specific drug delivery requirements ^{29,55}.

Phospholipids of both natural and synthetic origin are the primary building blocks used in liposome formulation ⁵⁶. However, liposomes composed exclusively of phospholipids often suffer from limited physical stability, short shelf-life, and increased membrane permeability, which may result in premature drug leakage. To overcome these limitations, sterols—most notably cholesterol—are frequently incorporated into the lipid bilayer. Cholesterol plays a critical role in modulating membrane rigidity, reducing bilayer permeability, and enhancing overall liposomal stability during storage and circulation ^{58,59}.

In addition to improving drug solubility and delivery efficiency, liposomes act as protective carriers that shield encapsulated drugs from degradation caused by enzymatic activity, chemical reactions, and environmental stressors such as variations in pH, temperature, and ionic strength⁶². The incorporation of auxiliary components can further enhance liposomal performance. Antioxidants such as vitamin E or its derivatives, counting d- α -tocopheryl polyethylene glycol 1000 succinate (TPGS), as well as polymers like chitosan and polyethylene glycol (PEG), are commonly integrated into liposomal membranes to improve stability, prolong circulation time, and optimize biodistribution profiles ⁶³.

Overall, liposomes offer numerous advantages as drug delivery vehicles, including spontaneous self-assembly, high drug-loading capacity, and enhanced solubility of poorly water-soluble drugs, improved drug stability, excellent biocompatibility, minimal toxicity, biodegradability, and low immunogenicity ⁶⁴⁻⁶⁸. Advances in liposomal engineering have further expanded their functionality, enabling the development of stimuli-responsive systems that discharge their payload in response to specific internal or external triggers such as pH changes, temperature variations, redox conditions, enzymatic activity, light, ultrasound, or magnetic fields. Additionally, surface functionalization strategies, including the development of immunoliposomes, have facilitated active, site-specific targeting, leading to controlled drug release, and co-delivery of multiple therapeutics, improved bio distribution, and reduced non-specific uptake by healthy tissues ⁶⁹⁻⁷³.

At present, four liposome-based formulations have received clinical approval for use in breast cancer therapy: Doxil®/Caelyx®, Myocet®, Lipodox®, and Lipusu®. Among these, Doxil®/Caelyx® is a PEGylatednanoliposomal formulation containing doxorubicin hydrochloride (DOX HCl) and is widely used in the treatment of several malignancies, including metastatic breast cancer ⁸⁵. Myocet® is a non-PEGylated liposomal formulation of doxorubicin that has been approved in the European Union since 2000 for use in combination regimens for metastatic breast cancer ⁶⁴. Lipodox®, another PEGylated liposomal formulation of

DOX HCl, was initially introduced in the United States as an alternative during shortages of Doxil® and has since been regarded as its generic equivalent⁸⁷. Lipusu® is a non-PEGylated liposomal formulation encapsulating paclitaxel (PTX) and has been approved in China for the treatment of HER2-positive metastatic breast cancer^{81,82}. However, comprehensive details regarding the lipid composition of Lipusu® have not been publicly disclosed. In addition to these approved products, DaunoXome®, a thermosensitive liposomal formulation of daunorubicin, has been evaluated for metastatic breast cancer but has not yet obtained clinical approval⁹⁰⁻⁹².

Notably, none of the currently approved liposomal formulations for breast cancer therapy employ active targeting strategies. Instead, their tumor accumulation relies predominantly on passive targeting mechanisms mediated by the enhanced permeability and retention (EPR) effect^{93,94}. Passive targeting refers to the

preferential accumulation of macromolecules and nanoscale carriers within tumor interstitial spaces as a result of the abnormal vascular architecture of tumor tissue [95]. Rapid tumor growth often exceeds oxygen accessibility, leading to hypoxic conditions that stimulate the release of pro-angiogenic factors such as vascular endothelial enlargement factor (VEGF). This process results in irregular angiogenesis, leaky vasculature, and deficient lymphatic drainage. As a consequence, nanoparticles—particularly those with diameters below approximately 400 nm—can accumulate within tumor and inflamed tissues. Nevertheless, the EPR effect is highly variable and is generally less pronounced in early-stage tumors and poorly vascularized tissues. Its effectiveness is mainly observed in solid tumors exceeding approximately 4.6 mm in diameter, with vascular pore size strongly influenced by tumor type, location, and pathological status^{93,94}.

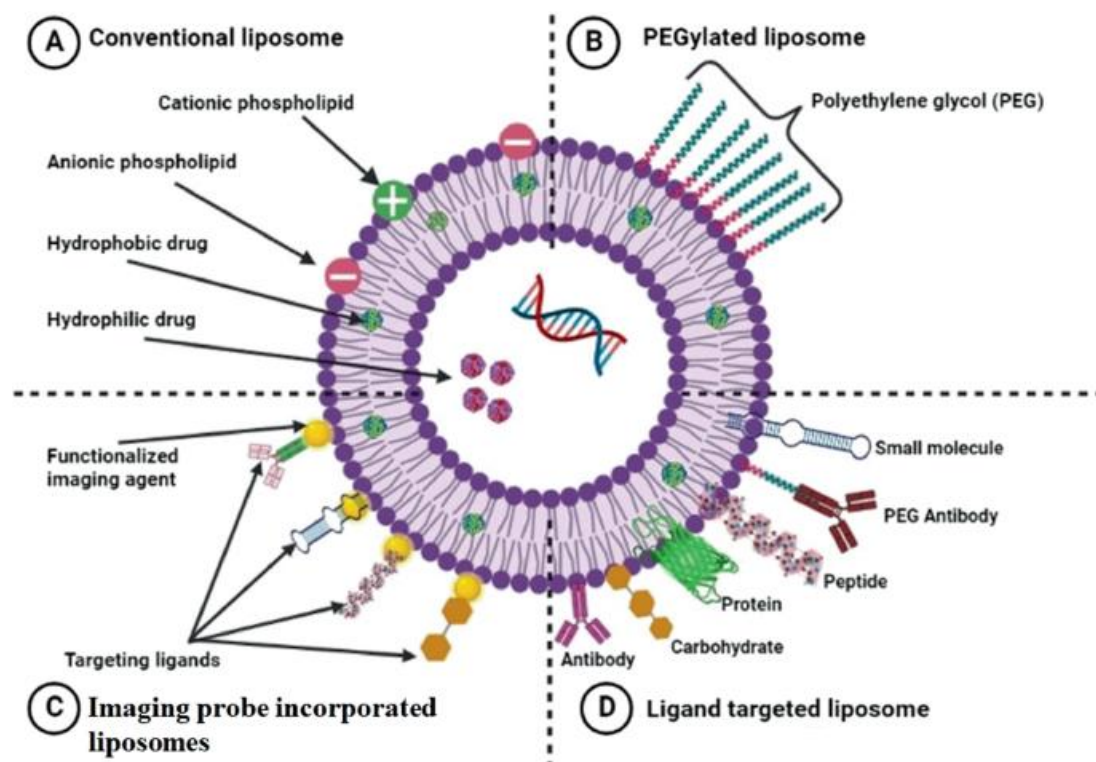


Figure 2: Liposome-based drug delivery strategies^{96,97}.

Following intravenous administration, non-PEGylated liposomal formulations such as Myocet® or Lipusu® circulate within the vascular compartment and are ultimately removed from systemic circulation by renal excretion or uptake by the mononuclear phagocyte system (MPS), also referred to as the reticuloendothelial system (RES)^{96,97}. Nanoscale carriers with diameters of approximately 8 nm or smaller undergo limited metabolic processing and are primarily eliminated through the kidneys, whereas larger liposomes are preferentially cleared by the MPS via opsonization-mediated pathways. During opsonization, plasma proteins, known as opsonins, adsorb onto the surface of

liposomal nanoparticles, marking them for recognition and subsequent phagocytosis by immune cells⁹⁸.

Surface modification of nanocarriers with inert polymers such as polyethylene glycol (PEG) can significantly attenuate opsonization. PEGylation creates a steric barrier around the nanoparticle surface, resulting in repulsive interactions with circulating blood components and conferring so-called “stealth” properties^{99,100}. This stealth effect reduces MPS-mediated clearance, thereby prolonging systemic circulation and improving pharmacokinetic performance. As an illustrative example, the PEGylated liposomal formulation Doxil®/Caelyx® demonstrates a

clearance half-life approximately 100-fold longer than that of free doxorubicin ¹⁰¹.

2. Targeted Nanoliposome-Based Approaches in Breast Cancer Therapy

Actively targeted liposomal drug delivery systems have gained considerable attention as an effective strategy to improve precision and treatment outcomes in cancer therapy. By incorporating specific targeting ligands on their surface, these systems enable preferential interaction with cancer cells, thereby enhancing therapeutic selectivity. This approach offers several important advantages, including: (i) enhanced cellular uptake of anticancer agents by tumor cells while minimizing drug exposure to healthy tissues, which in turn reduces systemic toxicity and lowers the likelihood of developing multidrug resistance (MDR); (ii) the potential to traverse physiological barriers such as the blood-brain barrier (BBB); and (iii) the ability to selectively recognize, image, and treat metastatic, recurrent, and breast cancer-linked cell populations ¹⁰².

Substantial interest in actively targeted nanomedicine has been demonstrated through both preclinical

investigations and clinical evaluations, particularly for the treatment of solid tumors. Regardless of its strong theoretical appeal, however, the practical implementation of active targeting remains challenging. Successful targeting requires not only the identification of appropriate and biologically relevant receptors but also the careful modification of liposomes with targeting ligands that exhibit high binding affinity without compromising the stealth properties necessary for prolonged circulation.

To achieve effective ligand attachment, liposomal surfaces are commonly functionalized through chemical modification using reactive groups that enable the conjugation of diverse targeting moieties. Several well-established conjugation strategies are employed for this purpose, including imine and amide bond formation, disulfide linkages, thiol-maleimide click chemistry reactions, and hydrazone-based crosslinking methods ^{103,104}. These functionalization approaches allow precise control over ligand density and orientation, which are critical parameters for optimizing targeting efficiency and therapeutic performance.

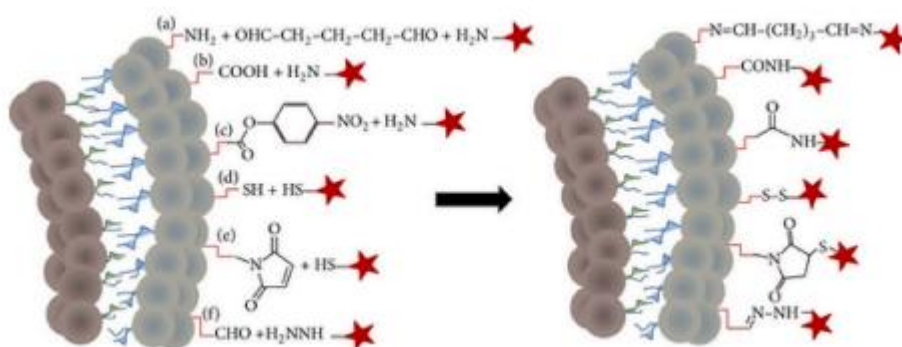


Figure 3: Overview of six principal chemical approaches (a–f) used for liposomal surface functionalization. Star symbols denote attached targeting ligands. Adapted from Ref. ¹⁰⁵.

Among these strategies, the thiol-maleimide click chemistry reaction is particularly popular, extensively utilized for conjugation between nanoparticles and various targeting agents ¹⁰⁶. Substitute methods for liposomal surface functionalization include adsorption or intercalation via electrostatic or hydrophobic interactions ^{105,107}. A variety of targeting ligands have been explored for actively targeted liposomal systems, including small molecules, monoclonal antibodies (mAbs), peptides, and aptamers. These ligands are designed to selectively bind to molecular targets expressed on breast cancer cells or within the tumor microenvironment (TME), thereby enhancing site-specific drug delivery. Initial drug-targeting strategies predominantly relied on full-length monoclonal antibodies because of their high binding affinity and specificity. However, limitations such as restricted tissue penetration, potential immunogenicity, complex manufacturing processes, and high production costs have shifted attention toward smaller antibody-derived constructs. Antibody fragments, including antigen-

binding fragments (Fab) and single-chain variable fragments (scFv), have emerged as more favorable alternatives due to their reduced size, improved permeability, and lower immunogenic potential ^{94,108}.

Peptides represent another widely used class of targeting ligands, owing to their ease of synthesis, relatively low cost, and ability to achieve selective binding while minimizing nonspecific interactions and opsonization. Despite these advantages, peptide-based ligands are inherently vulnerable to enzymatic degradation, which can limit their stability and in vivo performance ^{109,110}. Small-molecule ligands, such as sorafenib, offer benefits including high membrane permeability and cost-effectiveness; however, their limited targeting specificity may reduce selectivity toward cancer cells. In contrast, aptamers—short single-stranded DNA, RNA, or peptide sequences—exhibit exceptional binding affinity and specificity toward a wide range of targets, including small molecules, proteins, viruses, and whole cells. Compared with antibodies, aptamers provide advantages such as

smaller size, enhanced chemical stability, ease of large-scale synthesis, and straightforward chemical modification. Nevertheless, their clinical application is challenged by rapid systemic clearance and susceptibility to degradation in biological environments^{111–113}.

2.1. Cell Surface Receptor-Mediated Targeting Strategies

2.1.1. CXCR4-Directed Liposomal Systems for Cancer Treatment

C-X-C chemokine receptor type 4 (CXCR4) is a transmembrane G protein-coupled receptor that is widely expressed across numerous physiological systems. It plays a critical role in regulating cell migration, embryonic development, hematopoietic cell trafficking, and neuronal processes such as neurite and axonal outgrowth¹⁷². CXCR4 is present on the surface of diverse cell types, including hematopoietic and endothelial cells, neurons, stem cells, and a wide range of malignant cells. Deregulated or elevated CXCR4 expression has been strongly coupled with haematological malignancies and is recognized as an adverse prognostic marker in several solid tumors, including breast cancer^{173,174}.

Accumulating evidence has identified the CXCR4/CXCL12 signaling axis as a key driver of breast cancer progression and metastatic dissemination. CXCL12, also known as stromal cell-derived factor-1 (SDF-1), functions as a potent chemokine that directs CXCR4-expressing tumor cells toward organs with high CXCL12 expression, such as the lungs, bones, and lymph nodes. This chemotactic gradient facilitates tumor cell homing and colonization at distant metastatic sites^{175,176}. Within the tumor microenvironment (TME), CXCR4 signaling further contributes to immune cell recruitment, stromal reorganization, and enhanced tumor cell motility, thereby supporting tumor growth and invasion.

Exploiting this biology, Lu *et al.* developed a CXCR4-targeted liposomal delivery system to improve the therapeutic efficacy of the CXCR4 antagonist AMD2000¹¹⁶. AMD2000, which has been clinically approved since 2008 for the treatment of non-Hodgkin's lymphoma and multiple myeloma, was incorporated into the liposomal formulation in a dual manner: encapsulated within the aqueous core and simultaneously conjugated to the liposomal surface. This design enabled the nanocarrier to serve both as a targeting moiety and as a therapeutic agent. Consequently, CXCR4 signalling was inhibited at the cell surface through ligand-receptor interaction and intracellular following liposomal internalization. This dual-action strategy effectively modulated immune and stromal components of the TME, resulting in its functional reprogramming and structural remodeling.

In a complementary approach, Zhang *et al.* reported a peptide-directed liposomal platform that combined chemotherapy with photothermal therapy to enhance breast cancer treatment outcomes¹¹⁷. This system employed a novel targeting peptide, p12 (QGSRRRNTVDDWISRRRALC), conjugated to PEGylated

liposomes co-encapsulating doxorubicin (DOX) and indocyanine green (ICG), a clinically approved photothermal agent. The p12 peptide promoted selective tumor accumulation of the liposomes, thereby limiting off-target distribution and reducing DOX-associated toxicities, including cardiotoxicity and metastatic spread. In addition, ICG enabled localized photothermal activation, triggering controlled DOX release at tumor sites upon exposure to temperatures exceeding 41 °C.

Despite the therapeutic promise of targeting the CXCL12/CXCR4 axis—highlighted by the clinical use of AMD2000—important challenges remain. The regulatory complexity of this signaling pathway and the long-term consequences of sustained CXCR4 inhibition are not yet fully understood. Prolonged CXCR4 blockade has been associated with adverse hematological effects, including leucocytosis, thrombocytopenia, and splenomegaly. These toxicities are attributed to the broad physiological expression of CXCR4 in multiple organs, such as the heart, liver, spleen, and kidneys¹⁷⁷.

Recent advances have further expanded CXCR4-targeted nanotherapeutic strategies to include gene-silencing approaches. Guo *et al.* developed pH-responsive liposomes functionalized with CXCR4-targeting ligands and loaded with small interfering RNA (siRNA) against lipocalin-2 (Lcn2), a protein frequently overexpressed in epithelial cancers and closely linked to epithelial-mesenchymal transition (EMT)¹¹⁴. This combined strategy of receptor targeting and Lcn2 gene suppression significantly reduced metastatic breast cancer cell migration, particularly in triple-negative breast cancer (TNBC) models. Supporting these findings, Liu *et al.* demonstrated that liposomes decorated with varying surface densities of the CXCR4-binding peptide DV1 exhibited differential cellular uptake and effectively inhibited TNBC cell migration. This effect was mediated through down regulation of motility-related proteins triggered by CXCR4-dependent signalling pathways at the cell surface¹¹⁵.

2.1.2. Targeting Cell Surface-Associated Nucleosomes

During programmed cell death and necrotic processes, intracellular nuclear components may be released into the extracellular environment, triggering immune responses that lead to the generation of antinuclear antibodies (ANAs) directed against these nuclear antigens. ANAs are widely used as diagnostic and prognostic markers in systemic immune-mediated diseases, with specific antibody subsets closely associated with distinct clinical conditions. For instance, antibodies against double-stranded DNA are a hallmark of systemic lupus erythematosus¹⁷⁸. A related group of nuclear targets, collectively known as extractable nuclear antigens (ENAs), can be isolated from cell nuclei under saline conditions and include ribonucleoproteins and non-histone proteins such as Smith (Sm), ribonucleoprotein, and scleroderma-70 (Scl-70). In addition to their established role in autoimmune disease diagnostics, these nuclear antigens have also been explored as potential biomarkers in oncology. Notably,

their application in breast cancer has shown promise for improving early disease detection and clinical assessment^{179,180}.

Building on this concept, Torchilin and colleagues developed a monoclonal antinuclear antibody, mAb 2C5, that specifically recognizes tumor-associated cell surface nucleosomes across a range of cancer types. This antibody was employed to functionalize Doxil® liposomes, resulting in targeted nanocarriers capable of selectively binding to tumor cells. In vitro studies demonstrated a 3- to 8-fold increase in cellular uptake and internalization compared with non-targeted liposomes, with enhanced cytotoxicity observed even in doxorubicin-resistant cell lines^{118,119}. Subsequent in vivo evaluation using ¹¹¹In-labeled mAb 2C5-liposomes and whole-body γ -scintigraphic imaging confirmed preferential tumor accumulation and superior antitumor efficacy in subcutaneous 4T1 murine tumor models¹²⁰.

Expanding this strategy, Narayanaswamy and Torchilin developed dual-drug liposomal systems combining paclitaxel (PTX) and salinomycin to target both bulk breast cancer cells and cancer stem cells (CSCs), with the goal of mitigating tumor growth and metastasis¹²¹. Despite promising preclinical results, as of 2022 no clinical trials involving mAb 2C5 have been registered on clinicaltrials.gov, prompting exploration of alternative nanocarriers, including polymeric micelles and dendrimers, for potential clinical translation.

2.1.3. Eph Receptor Tyrosine Kinases as Therapeutic Targets

Eph receptors, a family of tyrosine kinase receptors, play pivotal roles in various cellular processes such as cell-cell interactions, proliferation, differentiation, signalling, migration, and tissue morphogenesis, as well as in pathological processes¹⁸¹. Among the fourteen members of the Eph receptor tyrosine kinase family, EphA2 has emerged as one of the most extensively studied receptors in oncology. Elevated expression of EphA2 has been reported across a broad range of malignancies, including cancers of the brain, bladder, breast, lung, skin, ovary, and prostate¹⁸². In breast cancer specifically, EphA2 plays a central role in multiple aspects of tumor biology, including uncontrolled cell proliferation, angiogenesis, therapeutic resistance, tumor progression, cellular migration, and metastatic dissemination¹⁸³. Importantly, aggressive breast cancer phenotypes that lack estrogen receptor alpha (ER α) expression consistently display increased EphA2 levels, further highlighting its association with poor prognosis and disease aggressiveness¹⁸³. These characteristics position EphA2 as a highly attractive molecular target for the development of targeted therapeutic strategies.

Recognizing the therapeutic potential of EphA2, several research groups have focused on designing EphA2-directed drug delivery systems. A prominent example is MM-200, an EphA2-targeted nanoliposomal formulation encapsulating the chemotherapeutic agent docetaxel. This system has been developed for the treatment of

multiple tumor types, including triple-negative breast cancer (TNBC), where effective targeted therapies are limited¹²². Phase I clinical trials (NCT03076372) have been conducted to evaluate the safety and tolerability of this approach in human subjects. However, as of 2022, detailed outcomes beyond safety assessments have not yet been publicly reported¹⁸⁴.

Building on this platform, the same research group further investigated a combinatorial treatment strategy that integrates chemotherapy with immunotherapy. In this approach, docetaxel delivered via the EphA2-targeted nanoliposomal system was combined with immune checkpoint inhibitors directed against programmed cell death protein 1 (PD-1), programmed death ligand 1 (PD-L1), and cytotoxic T-lymphocyte-associated protein 4 (CTLA-4). This combination was designed to overcome tumor immune evasion, therapeutic resistance, and disease recurrence—particularly in TNBC, which is often characterized by low intratumoral T-cell infiltration and poor response to immunotherapy alone¹⁸⁵.

In a TNBC tumor model, treatment with EphA2-targeted liposomes co-delivering docetaxel and an anti-programmed cell death receptor-1 (PD-1) antibody produced a therapeutic response in approximately 60% of cases, with sustained resistance to tumor rechallenge and a pronounced immunomodulatory effect. In a related approach, doxorubicin (DOX)-loaded stealth liposomes were surface-functionalized with the homing peptide YSAYPDSVPMMSK and evaluated under both in vitro and in vivo conditions¹²⁴. Notably, modification with YSAYPDSVPMMSK significantly enhanced the antitumor activity of DOX by promoting apoptosis in cancer cells, suppressing tumor progression and CD31 expression, and limiting angiogenic and metastatic potential. Collectively, these findings underscore the therapeutic promise of EphA2-targeted strategies in breast cancer and support their potential translational relevance based on encouraging preclinical and emerging clinical evidence.

2.1.4. Folate Receptor-Mediated Drug Delivery in Breast Cancer

Reduced folates are essential cofactors in amino acid metabolism and nucleic acid synthesis, playing a pivotal role in normal cell survival. Folate receptors (FRs) are glycoprotein receptors for vitamin B9, with four identified isoforms (α , β , γ , and δ) that exhibit tissue-specific expression patterns¹⁸⁶. Among these, folate receptor- α (FR α) is frequently overexpressed in tumors due to the elevated requirement for folate during DNA repair and replication associated with carcinogenesis. Consequently, FR α has emerged as a biomarker and therapeutic target across multiple malignancies, including breast, ovarian, brain, lung, and colorectal cancers^{187,188}.

Folate-decorated, long-circulating, and pH-responsive liposomal platforms have been widely explored as therapeutic systems for metastatic, multidrug-resistant (MDR), and triple-negative breast cancer (TNBC). As an illustrative example, Gazzano *et al.* engineered

liposomes encapsulating doxorubicin (DOX) chemically linked to nitric oxide (NO)-donating groups to overcome P-glycoprotein-mediated drug efflux in MDR breast cancer models¹³². Functionalization of the liposomal surface with folate enabled receptor-mediated internalization, promoting intracellular trafficking toward both nuclear and mitochondrial compartments. Within these organelles, DOX triggered DNA damage, cell cycle arrest, and activation of mitochondria-dependent apoptotic pathways.

In animal models, the folate-targeted liposomal formulation markedly suppressed tumor growth in breast cancers expressing P-gp and folate receptor α (FR α), whereas treatment with free DOX or the clinically approved formulation Caelyx® failed to produce comparable antitumor effects. Importantly, both primary tumor cells and cells isolated from treated tumors retained sensitivity to successive treatment cycles, suggesting sustained therapeutic efficacy and reduced propensity for resistance development.

In another approach, Deng et al. designed folate-targeted liposomes co-encapsulating DOX and functionalized with PEG chains cleavable by matrix metalloproteinase-2 (MMP-2)¹³³. This strategy leveraged chemotherapy-induced immunogenic cell death to convert tumors into an in situ “vaccine” while simultaneously targeting 4T1 breast cancer cells and immunosuppressive M2 tumor-associated macrophages (M2-TAMs) via the folate receptor. Coupled with cytosine-phosphate-guanine (CpG) therapy to enhance T cell activation, this combinatorial approach not only reduced primary tumor growth but also suppressed lung metastases and metastatic nodal expansion.

Despite these advances, challenges remain for folate receptor-targeted nanocarriers. Circulating folate from diet or supplements can compete with receptor-mediated uptake, limiting therapeutic efficiency. Furthermore, the high expression of FR α in normal kidney tissues can lead to off-target accumulation, necessitating careful consideration in clinical translation.

2.2. Transmembrane Receptors as Targets in Breast Cancer Therapy

2.2.1. Biotin receptor

Biotin is a water-soluble essential vitamin that plays a critical role in numerous metabolic pathways. Cellular uptake of biotin occurs through two primary mechanisms: passive diffusion at supraphysiological concentrations and active, carrier-mediated transport at lower concentrations. The latter process is facilitated by biotin-specific receptors and sodium-dependent multivitamin transporters (SMVTs), which are widely expressed in absorptive tissues and are responsible for the transport of several vitamins and essential cofactors across cell membranes^{189,190}. Increasing evidence suggests that altered expression of biotin receptors and SMVTs is associated with various malignancies, including breast cancer, thereby highlighting these transport systems as attractive targets for diagnostic imaging and targeted therapeutic delivery^{191,192}.

To investigate how ligand architecture influences the performance of targeted liposomal nanocarriers in breast cancer, Lu et al. engineered liposomes functionalized with double-branched biotin ligands at different surface densities¹⁵⁷. Their findings revealed that liposomes modified with double-branched biotin exhibited markedly enhanced interaction with SMVT-expressing breast cancer cells, as demonstrated by increased cellular uptake in vitro and improved tumor accumulation in vivo. In addition, paclitaxel (PTX)-loaded liposomes incorporating double-branched biotin showed superior antitumor efficacy compared with non-targeted or single-ligand systems, indicating that increased ligand density on the liposomal surface can significantly enhance targeting efficiency. Building on these results, the same group further examined the effects of higher-order ligand branching by incorporating tri-branched and tetra-branched biotin moieties onto the liposomal surface, thereby providing deeper insight into the relationship between ligand multiplicity and targeting performance¹⁵⁸.

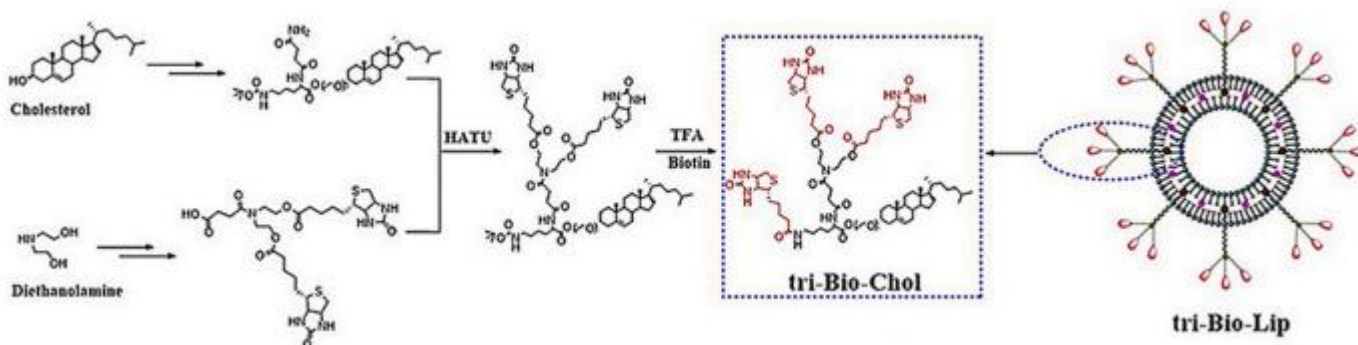


Figure 4: Example of biotin receptor-targeted liposomes: Tri-branched biotin-functionalized liposomes loaded with paclitaxel (PTX) were used to demonstrate that both ligand density and spatial orientation significantly influence cellular uptake in breast cancer cells. Adopted from Ref.¹⁵⁸.

Cellular uptake and cytotoxicity studies demonstrated that paclitaxel (PTX)-loaded tri-branched biotin-functionalized liposomes achieved the highest levels of internalization and anti-proliferative activity in breast cancer cells. In vivo imaging using 4T1 tumor-bearing mice confirmed these observations, highlighting that both the density and spatial configuration of biotin residues modulate the binding affinity between targeted liposomes and sodium-dependent multivitamin transporters (SMVT) on tumor cells.

Building on this concept, Huang et al.¹⁵⁹ developed a dual-targeting liposomal system functionalized with both biotin and glucose on a double-branched surface. This strategy significantly enhanced cellular uptake and tumor accumulation in vitro and in vivo compared with liposomes modified with a single targeting ligand. These results emphasize the importance of ligand density and multivalent presentation in optimizing receptor-mediated uptake and underscore the potential of biotin-based targeting approaches to improve the therapeutic performance of liposomal drug delivery systems in breast cancer.

2.2.2. Cluster of differentiation ⁴⁴

CD44 is a widely expressed transmembrane glycoprotein involved in cell adhesion, proliferation, and migration, with aberrant expression strongly associated with tumor progression, angiogenesis, and metastasis in breast cancer, particularly triple-negative breast cancer

(TNBC)^{193,194}. CD44 primarily binds hyaluronic acid (HA), a major extracellular matrix component, and this interaction activates signaling pathways that promote tumorigenesis, making the CD44–HA axis an attractive therapeutic target^{195,196}.

Exploiting this interaction, HA-functionalized liposomal drug delivery systems have been extensively developed for targeted breast cancer therapy. Lv et al. engineered thermosensitive liposomes encapsulating the MMP inhibitor marimastat and an HA-conjugated paclitaxel (PTX) prodrug, enabling heat-triggered drug release that enhanced tumor penetration and significantly inhibited tumor growth and angiogenesis in vivo¹⁶⁰. Similarly, Han et al. designed HA-conjugated gemcitabine-loaded liposomes to simultaneously target breast cancer cells and cancer stem cells, improving intracellular drug uptake while reducing systemic toxicity and therapeutic resistance¹⁶¹. In another innovative approach, Jiang et al. developed a liposomal “nanodepot” system for the co-delivery of TRAIL and doxorubicin, achieving enhanced anticancer efficacy through the simultaneous activation of complementary therapeutic mechanisms¹⁶².

Overall, HA-decorated liposomes represent a versatile and effective strategy for CD44-mediated targeting in breast cancer, offering improved tumor selectivity, enhanced drug accumulation, and superior therapeutic outcomes.

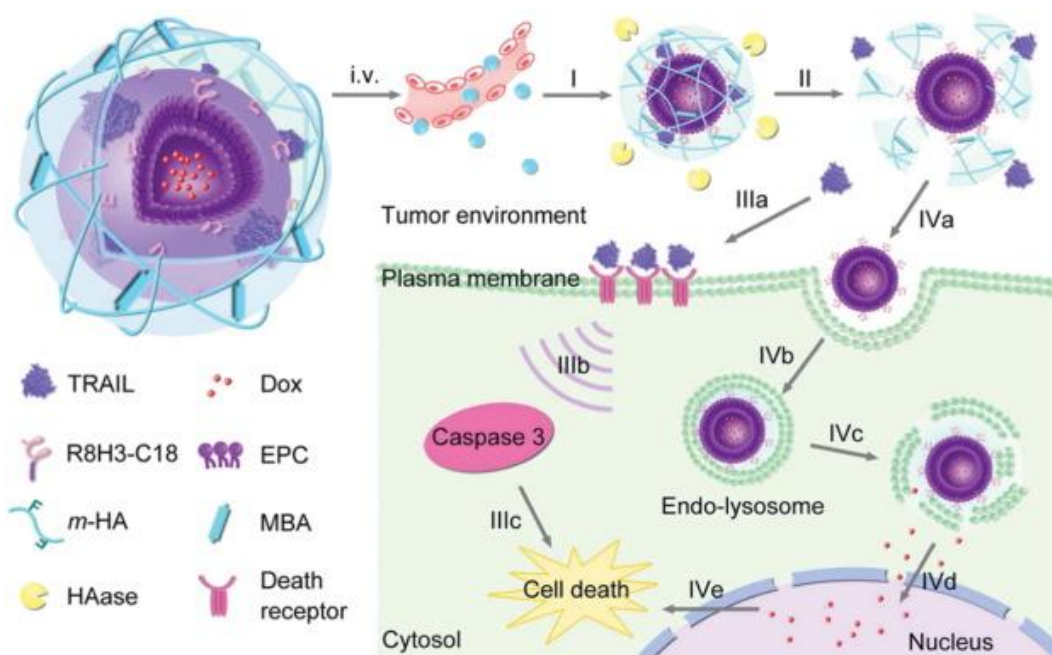


Figure 5: Illustration of the TRAIL/DOX-Gelipo design, showing the HA cross-linked outer shell encapsulating DOX and TRAIL, and the proposed multistage delivery of TRAIL to the cell surface and DOX to the nucleus as an example of CD44 liposomal targeting of breast cancer Ref¹⁶².

The strategy employed by Jiang et al. involved loading doxorubicin (DOX) into the aqueous core of the liposomes, while tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) was confined to the outer shell of the crosslinked hyaluronic acid (HA) shell¹⁶². This arrangement aimed to target cancer cells

overexpressing CD44, as the HA shell would degrade in the tumor microenvironment (TME) due to hyaluronidases, releasing TRAIL and facilitating liposome internalization.

Yang *et al.* reported that chitosan can effectively target CD44, a receptor highly expressed on breast cancer stem cells (CSCs) and within tumor tissues¹⁶⁴. In their study, chitosan-functionalized liposomes loaded with gambogic acid and radiolabeled with ⁸⁹Zr were engineered to selectively bind CD44 on TNBC CSCs. These nanocarriers demonstrated efficient tumor accumulation and exhibited pronounced antitumor activity in *in vivo* models.

In a related investigation, Ding *et al.* designed chitosan oligosaccharide-based liposomes encapsulating the photosensitizer HPPH together with the hypoxia-activated prodrug TH302 for CD44-directed therapy in TNBC¹⁶³. This multifunctional nanoplatform enabled simultaneous CD44 targeting, diagnostic imaging, photodynamic therapy, and hypoxia-responsive chemotherapy. By leveraging the hypoxic conditions generated following photodynamic treatment, TH302 was selectively activated, resulting in synergistic anticancer effects in both *in vitro* and *in vivo* models.

Furthermore, Guo *et al.* developed a liposomal formulation conjugated with anti-interleukin-6 receptor (IL6R) antibodies to modulate the tumor microenvironment (TME) of CD44-positive breast cancer cells in TNBC and luminal breast cancer mouse models¹⁶⁵. Inhibition of IL6R signaling attenuated stemness and angiogenesis in tumor-bearing mice and significantly reduced the metastatic potential of breast cancer stem cells to the lungs. Collectively, these studies highlight the therapeutic promise of CD44-targeted liposomal drug delivery systems for breast cancer, demonstrating their potential to effectively target cancer stem cells and remodel the tumor microenvironment, thereby improving overall treatment efficacy.

2.2.3. HER1 and HER2 Receptors in Targeted Breast Cancer Nanotherapy

The human epidermal growth factor receptor (HER) family, comprising HER1 (EGFR) to HER4, is a group of receptor tyrosine kinases that regulate critical cellular processes including proliferation, differentiation, survival, and tissue repair. HER1 and HER2 are frequently overexpressed in breast cancer, contributing to uncontrolled cell growth, resistance to apoptosis, enhanced metastasis, and increased angiogenesis^{197,198}. HER1 binds specific extracellular ligands from the epidermal growth factor family, whereas HER2 lacks a known endogenous ligand but forms heterodimers with other HER family members to activate downstream signaling pathways.

Overexpression of HER1 is observed in multiple breast cancer subtypes, particularly triple-negative breast cancer (TNBC), and correlates with poor prognosis. Therapeutic strategies targeting HER1, collectively referred to as anti-EGFR therapy, primarily involve monoclonal antibodies (mAbs) that block ligand binding and inhibit receptor signaling. Notable HER1-targeting mAbs include panitumumab, cetuximab (CET), and zalutumumab, which interfere with EGFR-mediated

pathways, thereby reducing tumor proliferation, angiogenesis, and metastatic potential¹⁹⁹.

HER1-targeted liposomal drug delivery systems have been developed to improve specificity and therapeutic efficacy. For example, CET-functionalized thermo-sensitive liposomes co-encapsulating doxorubicin (DOX) and citric acid-coated iron oxide nanoparticles enabled pH-triggered chemotherapy release combined with near-infrared (NIR) photothermal therapy. *In vitro* studies demonstrated enhanced uptake of CET-coated liposomes in HER1-positive breast cancer cells, leading to reduced viability, while *in vivo* studies in tumor-bearing mice showed increased local tumor heating and improved antitumor outcomes.

To reduce limitations associated with full-length antibodies, smaller fragments such as Fab and single-chain variable fragments (scFv) have been employed. Su *et al.* developed a pre-targeting system using anti-PEG Fab' and anti-HER1 scFv fragments, termed a PEG-engager, which enhanced the internalization and retention of PEGylated DOX liposomes in TNBC xenograft models while minimizing off-target effects.

Aptamer-based targeting strategies have also shown promise. Kim *et al.* engineered theranostic liposomes containing CdSe/ZnS quantum dots for imaging and siRNA for TNBC therapy, functionalized with an anti-HER1 aptamer. This system facilitated efficient cytoplasmic delivery of siRNA and demonstrated higher tumor accumulation compared with non-targeted liposomes *in vivo*, highlighting the potential of aptamer-functionalized nanocarriers for receptor-specific delivery.

HER2, often overexpressed in approximately 20–30% of breast cancers, has similarly been exploited for targeted nanotherapy. Liposomes functionalized with HER2-specific antibodies, affibodies, or peptides have been shown to enhance selective tumor uptake, improve drug delivery efficiency, and reduce systemic toxicity. Dual-targeting approaches, simultaneously addressing HER2 and other tumor-associated receptors, have been investigated to overcome receptor heterogeneity and multidrug resistance, further underscoring the versatility of HER1 and HER2 as cell surface targets in breast cancer nanoliposome therapy.

2.2.4. Luteinizing hormone-releasing hormone receptor

The luteinizing hormone-releasing hormone (LHRH/GnRHR) receptor regulates reproductive hormone secretion and is minimally expressed in normal visceral tissues but highly overexpressed in several hormone-related cancers, including breast cancer, making it an attractive target for drug delivery^{200,201}. He *et al.* developed gonadotropin-functionalized liposomes to improve the delivery of mitoxantrone, achieving enhanced uptake in LHRH receptor-overexpressing MCF-7 cells compared with receptor-negative cells. The system was further adapted for theranostic use by incorporating magnetic iron oxide nanoparticles for MRI^{165,166}. *In vivo*, the targeted liposomes produced significant tumor growth inhibition

with reduced systemic toxicity, primarily through receptor-mediated uptake and passive accumulation via the enhanced permeability and retention (EPR) effect, although imaging contrast diminished at later time points.

2.3. Intracellular Receptors as Therapeutic Targets in Breast Cancer Nanoliposomes

2.3.1. Estrogen Receptors as Intracellular Targets in Breast Cancer Nanotherapy

A substantial proportion of breast cancers overexpress estrogen receptors (ERs) and depend on estrogen signaling for tumor growth. ERs belong to the nuclear hormone receptor superfamily and exist as nuclear, extra-nuclear, and G protein-coupled receptors, with estrone (E1), estradiol (E2), and estriol (E3) serving as endogenous ligands. Among the two main subtypes, ER α is predominantly expressed in mammary tissue and drives the proliferation of ER-positive breast cancer, whereas ER β is more abundant in the prostate, making ER α the primary therapeutic target in breast cancer²⁰²⁻²⁰⁴. Although endocrine therapies are effective in ER-positive disease, relapse and resistance remain significant clinical challenges^{205,206}.

To enhance therapeutic efficacy, several liposomal systems targeting estrogen receptors have been developed using E1 or E2 as targeting ligands. E2-modified cationic liposomes have been employed to deliver antisense oligonucleotides against ER α/β mRNA, thereby sensitizing cancer cells to chemotherapy. In parallel, E1-targeted liposomes have been engineered for intracellular drug delivery using stimuli-responsive platforms, including pH- and ultrasound-triggered systems²⁰⁷. For example, pH-responsive doxorubicin-loaded liposomes enabled enhanced nuclear delivery and increased cytotoxicity in ER-positive breast cancer cells. Similarly, Han *et al.* reported E1-conjugated, PEGylated paclitaxel-loaded liposomes that exhibited significantly lower IC50 values in ER-positive MCF-7 cells and superior tumor growth inhibition *in vivo* compared with non-targeted formulations, with rapid tumor accumulation observed within hours after administration¹⁶⁸⁻¹⁷⁰.

2.4. Enzymes

2.4.1. Matrix Metalloproteinases as Therapeutic Targets in Breast Cancer Nanoliposomes

Matrix metalloproteinases (MMPs) constitute a family of enzymes crucial for ECM remodeling, wound healing, and angiogenesis, operating with zinc as a cofactor. Deregulated MMP activity has been implicated in various diseases, including cardiovascular disorders, inflammation, and cancer²⁰⁸. Different MMP types exhibit specificity in cleaving ECM components, categorizing them into groups like collagenases, membrane-types, and gelatinases. Among these, gelatinases, such as MMP-2 and MMP-9, have been particularly implicated in tumor progression, influencing growth, migration, invasion, and metastasis^{209,210}.

Elevated MMP levels in tumors correlate with adverse outcomes, including recurrence, invasion, and metastasis. However, MMPs also serve as valuable biomarkers and therapeutic targets, especially in controlled drug delivery systems. MMP-targeted liposomes have been developed to release drugs specifically in tumor tissues. For instance, liposomes functionalized with chlorotoxin, a peptide from scorpion venom known to bind MMP-2, exhibited enhanced uptake in metastatic breast cancer cells and demonstrated improved cytotoxicity and antimetastatic effects in mouse models^{211,212}.

In another approach, high MMP concentrations within tumors were exploited as an internal targeting mechanism. A novel liposomal co-delivery system developed by Ramadass *et al.* combined epigallocatechingallate (EGCG), an MMP inhibitor, with the chemotherapeutic agent paclitaxel (PTX). This synergistic system outperformed individual drug-loaded liposomes in inhibiting MMP-2 and MMP-9, suppressing invasion, enhancing cytotoxicity, and promoting apoptosis. These findings underscore the potential of MMP-targeted liposomal delivery systems in cancer therapy.

2.4.2. Tumor-Targeted Liposomal Therapy via Secretory Phospholipase A₂ Activation

Phospholipases represent a broad family of enzymes comprising more than 30 isoforms that catalyze the hydrolysis of phospholipids and are classified according to their catalytic mechanisms, structural characteristics, evolutionary relationships, and cellular localization²¹³. Among these, secretory phospholipase A₂ (sPLA₂) has gained considerable attention due to its involvement in inflammatory disorders, atherosclerosis, and tumor progression in several malignancies, including prostate, pancreatic, and breast cancers²¹⁴⁻²¹⁶. A distinguishing feature of sPLA₂ is its strong preference for hydrolyzing phospholipids within lipid bilayers, particularly those containing phosphatidylserine, rather than free lipids. This enzymatic specificity makes sPLA₂ an attractive endogenous trigger for the development of enzyme-responsive liposomal drug delivery systems capable of localized and controlled drug release within tumor tissues.

Oxaliplatin is a platinum-based chemotherapeutic agent that inhibits DNA synthesis and is widely used in colorectal cancer treatment. Notably, phase II clinical studies have demonstrated its activity in metastatic and triple-negative breast cancer patients who had previously received anthracycline- and/or taxane-based therapies²¹⁷⁻²²¹. However, the clinical utility of oxaliplatin is limited by dose-related toxicities, including myelosuppression, peripheral neuropathy, and gastrointestinal adverse effects. To improve its therapeutic index, PEGylated oxaliplatin-loaded liposomes have been extensively explored. One of the most advanced examples is LiPlaCis, and sPLA₂-triggered liposomal cisplatin formulation. A phase I clinical trial evaluated LiPlaCis in patients with advanced or refractory solid tumors, including metastatic breast cancer (NCT01861496), but safety

concerns resulted in temporary suspension and subsequent reformulation in 2009²²². Continued preclinical investigations and phase I/II clinical studies have since examined LiPlaCis across a range of advanced solid tumors, such as head and neck, colorectal, gastric, skin, and breast cancers^{171,223–225}.

In a related study, Ostrem et al. developed a sPLA2-responsive liposomal platform by carefully adjusting membrane fluidity and cholesterol content to enable enzyme-specific release of encapsulated oxaliplatin¹⁷¹. While in vitro experiments confirmed selective drug release in the presence of sPLA2, intravenous administration in sPLA2-secreting MT-3 tumor-bearing mice led to severe systemic toxicity, necessitating early termination of the study. The authors attributed this outcome to premature liposome activation in circulation, a consequence of elevated serum sPLA2 levels observed in certain outbred mouse strains—a phenomenon that may also occur in cancer patients. These findings underscore a critical translational challenge associated with enzyme-triggered liposomal systems. More recently, an mRNA-based predictive biomarker for LiPlaCis responsiveness has shown promising results in heavily pretreated metastatic breast cancer patients, supporting its progression toward randomized phase II clinical evaluation²²⁶.

3. Challenges and Prospects of Targeted Nanoliposomal Drug Delivery

The targeted Nanoliposomal drug delivery landscape presents numerous challenges and avenues for future exploration. Phospholipases, a diverse group of enzymes with over 30 isoforms, play a pivotal role in cleaving phospholipids, offering potential targets for intervention. Among these, secretory phospholipase A2 (sPLA2) stands out due to its association with inflammatory conditions, atherosclerosis, and various cancers, including prostate, pancreatic, and breast cancers. Its preference for cleave negatively charged phospholipid head groups suggests it as an ideal candidate for targeted drug delivery, mainly in liposomal formulations.

Efforts have been made to capitalize on sPLA2's activity for controlled and localized drug release through sPLA2-responsive liposomes. For instance, the development of PEGylatedoxaliplatin-loaded liposomes aims to mitigate the toxicity commonly associated with this chemotherapy agent, especially in colorectal cancer treatment. However, challenges persist, as demonstrated by the discontinuation of clinical trials for LiPlaCis, a cisplatin-encapsulating liposomal formulation, due to safety concerns.

Recent studies have highlighted the need for precise control over drug release triggered by sPLA2. Optimization of liposomal delivery systems, such as adjusting fluidity and cholesterol levels, shows promise in vitro but faces hurdles when translated to in vivo models. Systemic toxicity observed in animal models underscores the complexity of enzyme-triggered drug release in clinical settings. Notably, variations in sPLA2 expression levels among individuals, both in animal

models and cancer patients, present significant hurdles for achieving targeted drug release without adverse effects. Looking ahead, future research should address these challenges by refining nanoliposomal formulations to achieve greater specificity and efficacy. Additionally, advances in predictive models, such as mRNA-based drug response predictors, hold potential for tailoring treatment strategies to individual patients, offering a personalized approach to targeted nanoliposomal drug delivery. However, rigorous preclinical testing and clinical validation are imperative to ensure the safety and efficacy of these innovative approaches before widespread clinical adoption.

4. Summary

The consumption of functionalized liposome represents a promising approach to enhance breast cancer treatment through targeted drug delivery. By incorporating specific ligands or responsive triggers, such as those targeting over expressed receptors or enzymes in breast cancer cells, these modified liposomal nanocarriers offer precision in drug delivery while minimizing systemic toxicity. This approach enables the encapsulation of a variety of therapeutics, from traditional chemotherapeutic agents to innovative biologics and gene therapies, providing a versatile toolset for fighting breast cancer. Continued research in this area is expected to give in advanced liposomal formulations, eventually leading to improved treatment outcomes and enhanced quality of life for breast cancer patients.

Acknowledgement: I sincerely thank my guide, Dr. Vinod Mokale (University Department of Pharmaceutical Sciences, Chh. Sambhajinagar, MH), for his valuable guidance and support throughout the entire publication process

Ethical Approval: Not applicable.

Consent for Publication: All authors have read and approved the final manuscript and consent to its publication.

Human and Animal Ethical Rights: Not applicable.

Conflict of Interest: The authors declare no conflict of interest, and no funding was required to conduct these review data.

Availability of Data and Materials: The datasets used during the current review are available from the corresponding author on reasonable request.

Funding: No funding received

Author Contribution: Ajinkya Holkar-Writing of original review paper, Dr. Vinod Mokale-review and editing, conceptualisation and supervision

References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F, Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, *CA Cancer J Clin*, 2021; 71:209-249 <https://doi.org/10.3322/caac.21660> PMID:33538338
2. Howlader N, Altekruse SF, Li CI, Chen VW, Clarke CA, Ries LAG, Cronin KA, US incidence of breast cancer subtypes defined by joint

- hormone-receptor and HER2 status, *J Natl Cancer Inst*, 2014; 106 <https://doi.org/10.1093/jnci/dju055> PMID:24777111
PMCID:PMC4580552
3. Akinyemiju TF, Pisu M, Waterbor JW, Altekruse SF, Socioeconomic status and incidence of breast cancer by hormone receptor subtype, *SpringerPlus*, 2015; 4:1-8 <https://doi.org/10.1186/s40064-015-1282-2> PMID:26405628
PMCID:PMC4573746
 4. Momenimovahed Z, Salehiniya H, Epidemiological characteristics of and risk factors for breast cancer in the world, *Breast Cancer*, 2019; 11:151-164 <https://doi.org/10.2147/BCTT.S176070>
PMCID:PMC6462164
 5. Azamjahn N, Soltan-Zadeh Y, Zayeri F, Global trend of breast cancer mortality rate: a 25-year study, *Asian Pac J Cancer Prev*, 2019; 20:2015-2020 <https://doi.org/10.31557/APJCP.2019.20.7.2015>
PMid:31350959 PMCID:PMC6745227
 6. Akram M, Iqbal M, Daniyal M, Khan AU, Awareness and current knowledge of breast cancer, *Biol Res*, 2017; 50:33 <https://doi.org/10.1186/s40659-017-0140-9> PMID:28969709
PMCID:PMC5625777
 7. Núñez C, Capelo JL, Igrejas G, Alfonso A, Botana LM, Lodeiro C, An overview of the effective combination therapies for the treatment of breast cancer, *Biomaterials*, 2016; 97:34-50 <https://doi.org/10.1016/j.biomaterials.2016.04.027>
PMid:27162073
 8. Trevisi E, La Salvia A, Daniele L, Brizzi MP, De Rosa G, Scagliotti GV, Di Maio M, Neuroendocrine breast carcinoma: a rare but challenging entity, *Med Oncol*, 2020; 37:70 <https://doi.org/10.1007/s12032-020-01396-4> PMID:32712767
PMCID:PMC7382662
 9. Curtis C, Shah SP, Chin SF, Turashvili G, Rueda OM, Dunning MJ, Speed D, Lynch AG, Samarajiwa S, Yuan Y, Graf S, Ha G, Haffari G, Bashashati A, Russell R, McKinney S, Aparicio S, Brenton JD, Ellis L, Huntsman D, Pinder S, Murphy L, Bardwell H, Ding Z, Jones L, Liu B, Papatheodorou I, Sammut SJ, Wishart G, Chia S, Gelmon K, Speers C, Watson P, Blamey R, Green A, MacMillan D, Rakha E, Gillett C, Grigoriadis A, De Rinaldis E, Tutt A, Parisien M, Troup S, Chan D, Fielding C, Maia AT, McGuire S, Osborne M, Sayalero SM, Spiteri I, Hadfield J, Bell L, Chow K, Gale N, Kovalik M, Ng Y, Prentice L, Tavaré S, Markowitz F, Langerød A, Provenzano E, Purushotham A, Børresen-Dale AL, Caldas C, The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups, *Nature*, 2012; 486:346-352 <https://doi.org/10.1038/nature10983> PMID:22522925
PMCID:PMC3440846
 10. Leong ASY, Zhuang Z, The changing role of pathology in breast cancer diagnosis and treatment, *Pathobiology*, 2011; 78:99-114 <https://doi.org/10.1159/000292644> PMID:21677473
PMCID:PMC3128144
 11. Goldhirsch A, Wood WC, Coates AS, Gelber RD, Thürlimann B, Senn HJ, Strategies for subtypes-dealing with the diversity of breast cancer: highlights of the St Gallen international expert consensus on the primary therapy of early breast cancer 2011, *Ann Oncol*, 2011; 22:1736-1747 <https://doi.org/10.1093/annonc/mdr304>
PMid:21709140 PMCID:PMC3144634
 12. Prat A, Parker JS, Karginova O, Fan C, Livasy C, Herschkowitz JI, He X, Perou CM, Phenotypic and molecular characterization of the claudin-low intrinsic subtype of breast cancer, *Breast Cancer Res*, 2010; 12:1-18 <https://doi.org/10.1186/bcr2635> PMID:20813035
PMCID:PMC3096954
 13. Prat A, Pineda E, Adamo B, Galvan P, Fernández A, Gaba L, Díez M, Viladot M, Arance A, Muñoz M, Clinical implications of the intrinsic molecular subtypes of breast cancer, *Breast*, 2015; 24:S26-S35 <https://doi.org/10.1016/j.breast.2015.07.008> PMID:26253814
 14. Narod SA, BRCA mutations in the management of breast cancer: the state of the art, *Nat Rev Clin Oncol*, 2010; 7:702-707 <https://doi.org/10.1038/nrclinonc.2010.166> PMID:20956982
 15. Onitilo AA, Engel JM, Greenlee RT, Mukesh BN, Breast cancer subtypes based on ER/PR and Her2 expression: comparison of clinicopathologic features and survival, *Clin Med Res*, 2009; 7:4-13 <https://doi.org/10.3121/cmr.2009.825> PMID:19574486
PMCID:PMC2705275
 16. Malorni L, Shetty PB, De Angelis C, Hilsenbeck S, Rimawi MF, Elledge R, Osborne CK, De Placido S, Arpino G, Clinical and biologic features of triple-negative breast cancers in a large cohort of patients with long-term follow-up, *Breast Cancer Res Treat*, 2012; 136:795-804 <https://doi.org/10.1007/s10549-012-2315-y>
PMid:23124476 PMCID:PMC3513514
 17. Moo TA, Sanford R, Dang C, Morrow M, Overview of breast cancer therapy, *Pet Clin*, 2018; 13:339-354 <https://doi.org/10.1016/j.cpet.2018.02.006> PMID:30100074
PMCID:PMC6092031
 18. Lowery AJ, Kell MR, Glynn RW, Kerin MJ, Sweeney KJ, Locoregional recurrence after breast cancer surgery: a systematic review by receptor phenotype, *Breast Cancer Res Treat*, 2012; 133:831-841 <https://doi.org/10.1007/s10549-011-1891-6> PMID:22147079
 19. Denduluri N, Chavez-MacGregor M, Telli ML, Eisen A, Graff SL, Hassett MJ, Holloway JN, Hurria A, King TA, Lyman GH, Partridge AH, Somerfield MR, Trudeau ME, Wolff AC, Giordano SH, Selection of optimal adjuvant chemotherapy and targeted therapy for early breast cancer: ASCO clinical practice guideline focused update, *J Clin Oncol*, 2018; 36:2433-2443 <https://doi.org/10.1200/JCO.2018.78.8604> PMID:29787356
 20. Goldstein SR, Siddhanti S, Ciaccia AV, Plouffe L, A pharmacological review of selective oestrogen receptor modulators, *Hum Reprod Update*, 2000; 6:212-224 <https://doi.org/10.1093/humupd/6.3.212> PMID:10874566
 21. Smith IE, Dowsett M, Aromatase inhibitors in breast cancer, *N Engl J Med*, 2003; 348:2431-2442 <https://doi.org/10.1056/NEJMra023246> PMID:12802030
 22. Ali S, Mondal N, Choudhry H, Rasool M, Pushparaj PN, Khan MA, Mahfooz M, Sami GA, Jarullah J, Ali A, Jamal MS, Current management strategies in breast cancer by targeting key altered molecular players, *Front Oncol*, 2016; 6 <https://doi.org/10.3389/fonc.2016.00045>
 23. Alavi M, Hamidi M, Passive and active targeting in cancer therapy by liposomes and lipid nanoparticles, *Drug Metab Pers Ther*, 2019; 34 <https://doi.org/10.1515/dmpt-2018-0032>
PMid:30707682
 24. Mokhatri-Hesari P, Montazeri A, Health-related quality of life in breast cancer patients: review of reviews from 2008 to 2018, *Health Qual Life Outcomes*, 2020; 18:1-25 <https://doi.org/10.1186/s12955-020-01591-x> PMID:33046106
PMCID:PMC7552560
 25. Teixeira S, Carvalho MA, Castanheira EMS, Functionalized liposome and albumin-based systems as carriers for poorly water-soluble anticancer drugs: an updated review, *Biomedicines*, 2022; 10:486 <https://doi.org/10.3390/biomedicines10020486>
PMid:35203695 PMCID:PMC8962385
 26. Schleicher SM, Bach PB, Matsoukas K, Korenstein D, Medication overuse in oncology: current trends and future implications for patients and society, *Lancet Oncol*, 2018; 19:e200-e208 [https://doi.org/10.1016/S1470-2045\(18\)30099-8](https://doi.org/10.1016/S1470-2045(18)30099-8)
PMid:29611528
 27. Wadhvani N, Jatoi I, Overuse of neo-adjuvant chemotherapy for primary breast cancer, *Indian J Surg Oncol*, 2020; 11:12-14 <https://doi.org/10.1007/s13193-019-01002-8> PMID:32205961
PMCID:PMC7064655
 28. Li YJ, Lei YH, Yao N, Wang CR, Hu N, Ye WC, Zhang DM, Chen ZS, Autophagy and multidrug resistance in cancer, *Chin J Cancer*, 2017; 36:52 <https://doi.org/10.1186/s40880-017-0219-2>
PMid:28646911 PMCID:PMC5482965
 29. Singh SK, Singh S, Willard J, Singh R, Drug delivery approaches for breast cancer, *Int J Nanomed*, 2017; 12:6205-6218 <https://doi.org/10.2147/IJN.S140325> PMID:28883730
PMCID:PMC5576701
 30. Assaraf YG, Brozovic A, Gonçalves AC, Jurkovicova D, Line A, Machuqueiro M, Saponara S, Sarmento-Ribeiro AB, Xavier CPR, Vasconcelos MH, The multi-factorial nature of clinical multidrug

- resistance in cancer, *Drug Resist Updates*, 2019; 46:100645
<https://doi.org/10.1016/j.drup.2019.100645> PMID:31585396
31. Giacomini KM, Huang SM, Tweedie DJ, Benet LZ, Brouwer KLR, Chu X, Dahlin A, Evers R, Fischer V, Hillgren KL, Hoffmaster KA, Ishikawa T, Keppler D, Kim RB, Lee CA, Niemi M, Polli JW, Sugiyama Y, Swaan PW, Ware JA, Wright SH, Yee SW, Zamek-Gliszczynski MJ, Zhang L, Membrane transporters in drug development, *Nat Rev Drug Discov*, 2010; 9:215-236
<https://doi.org/10.1038/nrd3028> PMID:20190787
 PMCid:PMC3326076
 32. Tacar O, Sriamornsak P, Dass CR, Doxorubicin: an update on anticancer molecular action, toxicity and novel drug delivery systems, *J Pharm Pharmacol*, 2013; 65:157-170
<https://doi.org/10.1111/j.2042-7158.2012.01567.x>
 PMID:23278683
 33. He H, Liu C, Wu Y, Zhang X, Fan J, Cao Y, A multiscale physiologically-based pharmacokinetic model for doxorubicin to explore its mechanisms of cytotoxicity and cardiotoxicity in human physiological contexts, *Pharm Res*, 2018; 35:1-10
<https://doi.org/10.1007/s11095-018-2456-8> PMID:29987398
 PMCid:PMC6533104
 34. Zhang N, Shu G, Qiao E, Xu X, Shen L, Lu C, Chen W, Fang S, Yang Y, Song J, Zhao Z, Tu J, Xu M, Chen M, Du Y, Ji J, DNA-functionalized liposomes in vivo fusion for NIR-II/MRI guided pretargeted ferroptosis therapy of metastatic breast cancer, *ACS Appl Mater Interfaces*, 2022; 14:20603-20615
<https://doi.org/10.1021/acsmi.2c01105> PMID:35476429
 35. Gottesman MM, Fojo T, Bates SE, Multidrug resistance in cancer: role of ATP-dependent transporters, *Nat Rev Cancer*, 2002; 2:48-58
<https://doi.org/10.1038/nrc706> PMID:11902585
 36. Mao Z, Shen K, Zhu L, Xu M, Yu F, Xue D, Li H, Xue C, Comparisons of cardiotoxicity and efficacy of anthracycline-based therapies in breast cancer: a network meta-analysis of randomized clinical trials, *Oncol Res Treat*, 2019; 42:405-413
<https://doi.org/10.1159/000500204> PMID:31104059
 37. Caswell-Jin JL, Plevritis SK, Tian L, Cadham CJ, Xu C, Stout NK, Sledge GW, Mandelblatt JS, Kurian AW, Change in Survival in Metastatic Breast Cancer with Treatment Advances: Meta-Analysis and Systematic Review, *JNCI Cancer Spectr*, 2018; 2
<https://doi.org/10.1093/jncics/pky062> PMID:30627694
 PMCid:PMC6305243
 38. Cardoso F, Spence D, Mertz S, Corneliussen-James D, Sabelko K, Gralow J, Cardoso MJ, Peccatori F, Paonessa D, Benares A, Sakurai N, Beishon M, Barker SJ, Mayer M, Global analysis of advanced/metastatic breast cancer: decade report (2005-2015), *Breast*, 2018; 39:131-138
<https://doi.org/10.1016/j.breast.2018.03.002> PMID:29679849
 39. Li A, Schleicher SM, Andre F, Mitri ZI, Genomic alteration in metastatic breast cancer and its treatment, *Am Soc Clin Oncol Educ Book*, 2020; 1-14
https://doi.org/10.1200/EDBK_280463
 PMID:32213086
 40. Liu R, Zhou J, Yang S, Zhang Z, Efficacy and safety of pegylated liposomal doxorubicin-based chemotherapy of AIDS-related Kaposi's sarcoma, *Am J Ther*, 2018; 25:e719-e721
<https://doi.org/10.1097/MJT.0000000000000736>
 PMID:29509553
 41. Di Paolo A, Liposomal anticancer therapy: pharmacokinetic and clinical aspects, *J Chemother*, 2004; 90-93
<https://doi.org/10.1179/joc.2004.16.Supplement-1.90>
 PMID:15688620
 42. Miele E, Spinelli GP, Miele E, Tomao F, Tomao S, Albumin-bound formulation of paclitaxel (Abraxane® ABI-007) in the treatment of breast cancer, *Int J Nanomed*, 2009; 4:99-105
<https://doi.org/10.2147/IJN.S3061> PMID:19516888
 PMCid:PMC2720743
 43. Eloy JO, Claro de Souza M, Petrilli R, Barcellos JPA, Lee RJ, Marchetti JM, Liposomes as carriers of hydrophilic small molecule drugs: strategies to enhance encapsulation and delivery, *Colloids Surf B Biointerfaces*, 2014; 123:345-363
<https://doi.org/10.1016/j.colsurfb.2014.09.029> PMID:25280609
 44. Ansari L, Shiehzadeh F, Taherzadeh Z, Nikoofal-Sahlabadi S, Momtazi-Borojeni AA, Sahebkar A, Eslami S, The most prevalent side effects of pegylated liposomal doxorubicin monotherapy in women with metastatic breast cancer: a systematic review of clinical trials, *Cancer Gene Ther*, 2017; 24:189-193
<https://doi.org/10.1038/cgt.2017.9> PMID:28409561
 45. Fang X, Cao J, Shen A, Advances in anti-breast cancer drugs and the application of nano-drug delivery systems in breast cancer therapy, *J Drug Deliv Sci Technol*, 2020; 57:101662
<https://doi.org/10.1016/j.jddst.2020.101662>
 46. Shi J, Kantoff PW, Wooster R, Farokhzad OC, Cancer nanomedicine: progress, challenges and opportunities, *Nat Rev Cancer*, 2017; 17:20-37
<https://doi.org/10.1038/nrc.2016.108> PMID:27834398
 PMCid:PMC5575742
 47. Hare JJ, Lammers T, Ashford MB, Puri S, Storm G, Barry ST, Challenges and strategies in anti-cancer nanomedicine development: an industry perspective, *Adv Drug Deliv Rev*, 2017; 108:25-38
<https://doi.org/10.1016/j.addr.2016.04.025>
 PMID:27137110
 48. Bangham AD, A correlation between surface charge and coagulant action of phospholipids, *Nature*, 1961; 192:1197-1198
<https://doi.org/10.1038/1921197a0> PMID:13864660
 49. Singh G, Darwin R, Panda KC, Afzal SA, Katiyar S, Dhakar RC, Mani S, Gene expression and hormonal signaling in osteoporosis: from molecular mechanisms to clinical breakthroughs, *Journal of Biomaterials Science, Polymer Edition*, 2024;1-36
<https://doi.org/10.1080/09205063.2024.2445376>
<https://doi.org/10.1080/09205063.2024.2445376>
 PMID:39729311
 50. Elkhoury K, Koçak P, Kang A, Arab-Tehrany E, Ellis Ward J, Shin SR, Engineering smart targeting nanovesicles and their combination with hydrogels for controlled drug delivery, *Pharmaceutics*, 2020; 12:849
<https://doi.org/10.3390/pharmaceutics12090849>
 PMID:32906833 PMCid:PMC7559099
 51. Velot E, Elkhoury K, Kahn C, Kempf H, Linder M, Arab-Tehrany E, Bianchi A, Efficient TGF-β1 delivery to articular chondrocytes in vitro using agro-based liposomes, *Int J Mol Sci*, 2022; 23:2864
<https://doi.org/10.3390/ijms23052864> PMID:35270005
 PMCid:PMC8911360
 52. Passeri E, Bun P, Elkhoury K, Linder M, Malaplate C, Yen FT, Arab-Tehrany E, Transfer phenomena of nanoliposomes by live imaging of primary cultures of cortical neurons, *Pharmaceutics*, 2022; 14:2172
<https://doi.org/10.3390/pharmaceutics14102172>
 PMID:36297607 PMCid:PMC9608836
 53. Elkhoury K, Sanchez-Gonzalez L, Lavrador P, Almeida R, Gaspar V, Kahn C, Cleymand F, Arab-Tehrany E, Mano JF, Gelatin methacryloyl (GelMA) nanocomposite hydrogels embedding bioactive Naringin liposomes, *Polymers*, 2020; 12:2944
<https://doi.org/10.3390/polym12122944> PMID:33317207
 PMCid:PMC7764353
 54. Arab-Tehrany E, Elkhoury K, Francius G, Jierry L, Mano JF, Kahn C, Linder M, Curcumin loaded nanoliposomes localization by nanoscale characterization, *Int J Mol Sci*, 2020; 21:7276
<https://doi.org/10.3390/ijms21197276> PMID:33019782
 PMCid:PMC7584047
 55. Webster DM, Sundaram P, Byrne ME, Injectable nanomaterials for drug delivery: carriers, targeting moieties, and therapeutics, *Eur J Pharm Biopharm*, 2013; 84:1-20
<https://doi.org/10.1016/j.ejpb.2012.12.009> PMID:23313176
 56. Shende P, Ture N, Gaud RS, Trotta F, Lipid- and polymer-based plexes as therapeutic carriers for bioactive molecules, *Int J Pharm*, 2019; 558:250-260
<https://doi.org/10.1016/j.ijpharm.2018.12.085> PMID:30641179
 57. Vemuri S, Rhodes CT, Preparation and characterization of liposomes as therapeutic delivery systems: a review, *Pharm Acta Helv*, 1995; 70:95-111
[https://doi.org/10.1016/0031-6865\(95\)00010-7](https://doi.org/10.1016/0031-6865(95)00010-7) PMID:7651973
 58. Maurya SD, Prajapati S, Gupta A, Saxena G, Dhakar RC, Formulation Development and Evaluation of Ethosome of Stavudine, *Indian J.Pharm. Educ. Res.* 2010;44(1)

59. Gregoriadis G, Davis C, Stability of liposomes in vivo and in vitro is promoted by their cholesterol content and the presence of blood cells, *Biochem Biophys Res Commun*, 1979; 89:1287-1293 [https://doi.org/10.1016/0006-291X\(79\)92148-X](https://doi.org/10.1016/0006-291X(79)92148-X) PMID:496958
60. Drummond DC, Noble CO, Hayes ME, Park JW, Kirpotin DB, Pharmacokinetics and in vivo drug release rates in liposomal nanocarrier development, *J Pharm Sci*, 2008; 97:4696-4740 <https://doi.org/10.1002/jps.21358> PMID:18351638
61. Laouini A, Jaafar-Maalej C, Limayem-Blouza I, Sfar S, Charcosset C, Fessi H, Preparation, characterization and applications of liposomes: state of the art, *J Colloid Sci Biotechnol*, 2012; 1:147-168 <https://doi.org/10.1166/jcsb.2012.1020>
62. Chaudhry Q, Watkins R, Castle L, Nanotechnologies in food: what, why and how?, *RSC Nanosci Nanotechnol*, 2017; 1-19 <https://doi.org/10.1039/9781782626879-00001> PMID:PMC5357853
63. Khorasani S, Danaei M, Mozafari MR, Nanoliposome technology for the food and nutraceutical industries, *Trends Food Sci Technol*, 2018; 79:106-115 <https://doi.org/10.1016/j.tifs.2018.07.009>
64. Bulbake U, Doppalapudi S, Kommineni N, Khan W, Liposomal formulations in clinical use: an updated review, *Pharmaceutics*, 2017; 9:12 <https://doi.org/10.3390/pharmaceutics9020012> PMID:28346375 PMID:PMC5489929
65. Saraf S, Jain A, Tiwari A, Verma A, Panda PK, Jain SK, Advances in liposomal drug delivery to cancer: an overview, *J Drug Deliv Sci Technol*, 2020; 56:101549 <https://doi.org/10.1016/j.jddst.2020.101549>
66. Large DE, Abdelmessih RG, Fink EA, Auguste DT, Liposome composition in drug delivery design, synthesis, characterization, and clinical application, *Adv Drug Deliv Rev*, 2021; 176:113851 <https://doi.org/10.1016/j.addr.2021.113851> PMID:34224787
67. Elkhoury K, Chen M, Koçak P, Enciso-Martínez E, Bassous NJ, Lee MC, Byambaa B, Rezaei Z, Li Y, Ubina Lopez ME, Gurian M, Sobahi N, Hussain MA, Sanchez-Gonzalez L, Leijten J, Hassan S, Arab-Tehrany E, Ward JE, Shin SR, Hybrid extracellular vesicles-liposome incorporated advanced bioink to deliver microRNA, *Biofabrication*, 2022; 14:045008 <https://doi.org/10.1088/1758-5090/ac8621> PMID:35917808 PMID:PMC9594995
68. Elkhoury K, Russell CS, Sanchez-Gonzalez L, Mostafavi A, Williams TJ, Kahn C, Peppas NA, Arab-Tehrany E, Tamayol A, Soft-nanoparticle functionalization of natural hydrogels for tissue engineering applications, *Adv Healthc Mater*, 2019; 8:1900506 <https://doi.org/10.1002/adhm.201900506> PMID:31402589 PMID:PMC6752977
69. Antoniou AI, Giofrè S, Seneci P, Passarella D, Pellegrino S, Stimulus-responsive liposomes for biomedical applications, *Drug Discov Today*, 2021; 26:1794-1824 <https://doi.org/10.1016/j.drudis.2021.05.010> PMID:34058372
70. Srujana S, Anjamma M, Alimuddin, Singh B, Dhakar RC, Natarajan S, Hechhu R. A Comprehensive Study on the Synthesis and Characterization of TiO₂ Nanoparticles Using Aloe vera Plant Extract and Their Photocatalytic Activity against MB Dye. *Adsorption Science & Technology*. 2022;2022 <https://doi.org/10.1155/2022/7244006>
71. Belfiore L, Saunders DN, Ranson M, Thurecht KJ, Storm G, Vine KL, Towards clinical translation of ligand-functionalized liposomes in targeted cancer therapy: challenges and opportunities, *J Contr Release*, 2018; 277:1-13 <https://doi.org/10.1016/j.jconrel.2018.02.040> PMID:29501721
72. Sonju JJ, Dahal A, Singh SS, Jois SD, Peptide-functionalized liposomes as therapeutic and diagnostic tools for cancer treatment, *J Contr Release*, 2021; 329:624-644 <https://doi.org/10.1016/j.jconrel.2020.09.055> PMID:33010333 PMID:PMC8082750
73. Parajapati S, Maurya S, Das M, Tilak VK, Verma KK, Dhakar RC. Potential Application of Dendrimers in Drug Delivery: A Concise Review and Update. *Journal of Drug Delivery and Therapeutics*. 2016;6(2):71-88 <https://doi.org/10.22270/jddt.v6i2.1195>
74. Gabizon A, Peretz T, Sulkes A, Amselem S, Ben-Yosef R, Ben-Baruch N, Catane R, Biran S, Barenholz Y, Systemic administration of doxorubicin-containing liposomes in cancer patients: a phase I study, *Eur J Cancer Clin Oncol*, 1989; 25:1795-1803 [https://doi.org/10.1016/0277-5379\(89\)90350-7](https://doi.org/10.1016/0277-5379(89)90350-7) PMID:2632261
75. Gabizon A, Catane R, Uziely B, Kaufman B, Safra T, Cohen R, Martin F, Huang A, Barenholz Y, Prolonged circulation time and enhanced accumulation in malignant exudates of doxorubicin encapsulated in polyethylene-glycol coated liposomes, *Cancer Res*, 1994; 54:987-992
76. Keller AM, Mennel RG, Georgoulas VA, Nabholz JM, Erazo A, Lluch A, Vogel CL, Kaufmann M, von Minckwitz G, Henderson C, Mellars L, Alland L, Tendler C, Randomized phase III trial of pegylated liposomal doxorubicin versus vinorelbine or mitomycin C plus vinblastine in women with taxane-refractory advanced breast cancer, *J Clin Oncol*, 2004; 22:3893-3901 <https://doi.org/10.1200/JCO.2004.08.157> PMID:15459210
77. Batist G, Ramakrishnan G, Rao CS, Chandrasekharan A, Gutheil J, Guthrie T, Shah P, Khojasteh A, Nair MK, Hoelzer K, Tkaczuk K, Park YC, Lee LW, Reduced cardiotoxicity and preserved antitumor efficacy of liposome-encapsulated doxorubicin and cyclophosphamide compared with conventional doxorubicin and cyclophosphamide in a randomized, multicenter trial of metastatic breast cancer, *J Clin Oncol*, 2001; 19:1444-1454 <https://doi.org/10.1200/JCO.2001.19.5.1444> PMID:11230490
78. Batist G, Barton J, Chaikin P, Swenson C, Welles L, Myocet (liposome-encapsulated doxorubicin citrate): a new approach in breast cancer therapy, *Expert Opin Pharmacother*, 2002; 3:1739-1751 <https://doi.org/10.1517/14656566.3.12.1739> PMID:12472371
79. Chan S, Davidson N, Juozaityte E, Erdkamp F, Pluzanska A, Azarnia N, Lee LW, Phase III trial of liposomal doxorubicin and cyclophosphamide compared with epirubicin and cyclophosphamide as first-line therapy for metastatic breast cancer, *Ann Oncol*, 2004; 15:1527-1532 <https://doi.org/10.1093/annonc/mdh393> PMID:15367414
80. Burade V, Bhowmick S, Maiti K, Zalawadia R, Ruan H, Thennati R, Lipodox® (generic doxorubicin hydrochloride liposome injection): in vivo efficacy and bioequivalence versus Caelyx® (doxorubicin hydrochloride liposome injection) in human mammary carcinoma (MX-1) xenograft and syngeneic fibrosarcoma (WEHI 164) mouse models, *BMC Cancer*, 2017; 17:405 <https://doi.org/10.1186/s12885-017-3377-3> PMID:28587612 PMID:PMC5461687
81. Xu X, Wang L, Xu H, Huang X, Qian Y, Xiang J, Clinical comparison between paclitaxel liposome (Lipusu®) and paclitaxel for treatment of patients with metastatic gastric cancer, *Asian Pac J Cancer Prev*, 2013; 14:2591-2594 <https://doi.org/10.7314/APJCP.2013.14.4.2591> PMID:23725180
82. Wang H, Cheng G, Du Y, Ye L, Chen W, Zhang L, Wang T, Tian J, Fu F, Hypersensitivity reaction studies of a polyethoxylated castor oil-free, liposome-based alternative paclitaxel formulation, *Mol Med Rep*, 2013; 7:947-952 <https://doi.org/10.3892/mmr.2013.1264> PMID:23291923 PMID:PMC3597461
83. Chou H, Lin H, Liu JM, A tale of the two PEGylated liposomal doxorubicins, *OncoTargets Ther*, 2015; 8:1719-1720 <https://doi.org/10.2147/OTT.S79089> PMID:26203262 PMID:PMC4508070
84. FDA approves generic version of Doxil; expected to help resolve shortage, *Oncol Times*, 2013; 35:25 <https://doi.org/10.1097/01.COT.0000428636.40337.70>
85. Barenholz Y, Doxil® - the first FDA-approved nano-drug: lessons learned, *J Contr Release*, 2012; 160:117-134 <https://doi.org/10.1016/j.jconrel.2012.03.020> PMID:22484195
86. Eloy JO, Petrilli R, Trevizan LNF, Chorilli M, Immunoliposomes: a review on functionalization strategies and targets for drug delivery, *Colloids Surf B Biointerfaces*, 2017; 159:454-467 <https://doi.org/10.1016/j.colsurfb.2017.07.085> PMID:28837895
87. Gaspar RS, Florindo HF, Silva LC, Videira MA, Corvo ML, Martins BF, Silva-Lima B, Regulatory Aspects of Oncologicals:

- Nanosystems Main Challenges, Springer, Cham, 2014; pp. 425-452 https://doi.org/10.1007/978-3-319-08084-0_15
88. Zhu L, Chen L, Progress in research on paclitaxel and tumor immunotherapy, *Cell Mol Biol Lett*, 2019; 24:1-11 <https://doi.org/10.1186/s11658-019-0164-y> PMID:31223315 PMCID:PMC6567594
89. Bernabeu E, Cagel M, Lagomarsino E, Moretton M, Chiappetta DA, Paclitaxel: what has been done and the challenges remain ahead, *Int J Pharm*, 2017; 526:474-495 <https://doi.org/10.1016/j.ijpharm.2017.05.016> PMID:28501439
90. Gill PS, Wernz J, Scadden DT, Cohen P, Mukwaya GM, von Roenn JH, Jacobs M, Kempin S, Silverberg I, Gonzales G, Rarick MU, Myers AM, Shepherd F, Sawka C, Pike MC, Ross ME, Randomized phase III trial of liposomal daunorubicin versus doxorubicin, bleomycin, and vincristine in AIDS-related Kaposi's sarcoma, *J Clin Oncol*, 1996; 14:2353-2364 <https://doi.org/10.1200/JCO.1996.14.8.2353> PMID:8708728
91. Bellott R, Auvrignon A, Leblanc T, Péral Y, Gandemer V, Bertrand Y, Méchinaud F, Bellenger P, Vernois J, Leverger G, Baruchel A, Robert J, Pharmacokinetics of liposomal daunorubicin (DaunoXome) during a phase I-II study in children with relapsed acute lymphoblastic leukaemia, *Cancer Chemother Pharmacol*, 2000; 47:15-21 <https://doi.org/10.1007/s002800000206> PMID:11221955
92. O'Byrne KJ, Thomas AL, Sharma RA, DeCatris M, Shields F, Beare S, Steward WP, A phase I dose-escalating study of DaunoXome, liposomal daunorubicin, in metastatic breast cancer, *Br J Cancer*, 2002; 87:15-20 <https://doi.org/10.1038/sj.bjc.6600344> PMID:12085249 PMCID:PMC2364277
93. Danhier F, Feron O, Pr at V, To exploit the tumor microenvironment: passive and active tumor targeting of nanocarriers for anti-cancer drug delivery, *J Contr Release*, 2010; 148:135-146 <https://doi.org/10.1016/j.jconrel.2010.08.027> PMID:20797419
94. Sawant RR, Torchilin VP, Challenges in development of targeted liposomal therapeutics, *AAPS J*, 2012; 14:303-315 <https://doi.org/10.1208/s12248-012-9330-0> PMID:22415612 PMCID:PMC3326155
95. Reineke J, Terminology matters: there is no targeting, but retention, *J Contr Release*, 2018; 273:180-183 <https://doi.org/10.1016/j.jconrel.2018.01.016> PMID:29360476
96. Scherphof GL, Dijkstra J, Spanjer HH, Derksen JT, Roerdink FH, Uptake and intracellular processing of targeted and nontargeted liposomes by rat Kupffer cells in vivo and in vitro, *Ann NY Acad Sci*, 1985; 446:368-384 <https://doi.org/10.1111/j.1749-6632.1985.tb18414.x> PMID:2409883
97. Longmire M, Choyke PL, Kobayashi H, Clearance properties of nano-sized particles and molecules as imaging agents: considerations and caveats, *Nanomed*, 2008; 3:703-717 <https://doi.org/10.2217/17435889.3.5.703> PMID:18817471 PMCID:PMC3407669
98. Moghimi SM, Farhangrazi ZS, Nanomedicine and the complement paradigm, *Nanomed Nanotechnol Biol Med*, 2013; 9:458-460 <https://doi.org/10.1016/j.nano.2013.02.011> PMID:23499667
99. Torchilin VP, Targeted pharmaceutical nanocarriers for cancer therapy and imaging, *AAPS J*, 2007; 9:e128-e147 <https://doi.org/10.1208/aapsj0902015> PMID:17614355 PMCID:PMC2751402
100. Suk JS, Xu Q, Kim N, Hanes J, Ensign LM, PEGylation as a strategy for improving nanoparticle-based drug and gene delivery, *Adv Drug Deliv Rev*, 2016; 99:28-51 <https://doi.org/10.1016/j.addr.2015.09.012> PMID:26456916 PMCID:PMC4798869
101. Charrois GJR, Allen TM, Multiple injections of pegylated liposomal doxorubicin: pharmacokinetics and therapeutic activity, *J Pharmacol Exp Ther*, 2003; 306:1058-1067 <https://doi.org/10.1124/jpet.103.053413> PMID:12808004
102. Gu W, Meng F, Haag R, Zhong Z, Actively targeted nanomedicines for precision cancer therapy: concept, construction, challenges and clinical translation, *J Contr Release*, 2021; 329:676-695 <https://doi.org/10.1016/j.jconrel.2020.10.003> PMID:33022328
103. Almeida B, Nag OK, Rogers KE, Delehanty JB, Recent progress in bioconjugation strategies for liposome-mediated drug delivery, *Mol Basel Switz*, 2020; 25:5672 <https://doi.org/10.3390/molecules25235672> PMID:33271886 PMCID:PMC7730700
104. de Lima PHC, Butera AP, Cabe a LF, Ribeiro-Viana RM, Liposome surface modification by phospholipid chemical reactions, *Chem Phys Lipids*, 2021; 237:105084 <https://doi.org/10.1016/j.chemphyslip.2021.105084> PMID:33891960
105. Marqu s-Gallego P, de Kroon AIPM, Ligation strategies for targeting liposomal nanocarriers, *BioMed Res Int*, 2014; 2014:e129458 <https://doi.org/10.1155/2014/129458> PMID:25126543 PMCID:PMC4122157
106. Taiariol L, Chaix C, Farre C, Moreau E, Click and bioorthogonal chemistry: the future of active targeting of nanoparticles for nanomedicines?, *Chem Rev*, 2022; 122:340-384 <https://doi.org/10.1021/acs.chemrev.1c00484> PMID:34705429
107. Riaz M, Riaz M, Zhang X, Lin C, Wong K, Chen X, Zhang G, Lu A, Yang Z, Surface functionalization and targeting strategies of liposomes in solid tumor therapy: a review, *Int J Mol Sci*, 2018; 19:195 <https://doi.org/10.3390/ijms19010195> PMID:29315231 PMCID:PMC5796144
108. Crivianu-Gaita V, Thompson M, Aptamers, antibody scFv, and antibody Fab' fragments: an overview and comparison of three of the most versatile biosensor biorecognition elements, *Biosens Bioelectron*, 2016; 85:32-45 <https://doi.org/10.1016/j.bios.2016.04.091> PMID:27155114
109. Forssen E, Willis M, Ligand-targeted liposomes, *Adv Drug Deliv Rev*, 1998; 29:249-271 [https://doi.org/10.1016/S0169-409X\(97\)00083-5](https://doi.org/10.1016/S0169-409X(97)00083-5) PMID:10837594
110. Ruoslahti E, Peptides as targeting elements and tissue penetration devices for nanoparticles, *Adv Mater*, 2012; 24:3747-3756 <https://doi.org/10.1002/adma.201200454> PMID:22550056 PMCID:PMC3947925
111. Xu Y, Phillips JA, Yan J, Li Q, Fan ZH, Tan W, Aptamer-based microfluidic device for enrichment, sorting, and detection of multiple cancer cells, *Anal Chem*, 2009; 81:7436-7442 <https://doi.org/10.1021/ac9012072> PMID:19715365 PMCID:PMC3164879
112. Dickey DD, Giangrande PH, Oligonucleotide aptamers: a next-generation technology for the capture and detection of circulating tumor cells, *Methods*, 2016; 97:94-103 <https://doi.org/10.1016/j.ymeth.2015.11.020> PMID:26631715 PMCID:PMC4792782
113. Zhou Z, Liu M, Jiang J, The potential of aptamers for cancer research, *Anal Biochem*, 2018; 549:91-95 <https://doi.org/10.1016/j.ab.2018.03.008> PMID:29548926
114. Guo P, You J-O, Yang J, Jia D, Moses MA, Auguste DT, Inhibiting metastatic breast cancer cell migration via the synergy of targeted, pH-triggered siRNA delivery and chemokine axis blockade, *Mol Pharm*, 2014; 11:755-765 <https://doi.org/10.1021/mp4004699> PMID:24467226 PMCID:PMC3993942
115. Liu D, Guo P, McCarthy C, Wang B, Tao Y, Auguste DT, Peptide density targets and impedes triple negative breast cancer metastasis, *Nat Commun*, 2018; 9:1-11 <https://doi.org/10.1038/s41467-018-05035-5> PMID:29973594 PMCID:PMC6031661
116. Lu G, Qiu Y, Su X, Targeting CXCL12-CXCR4 signaling enhances immune checkpoint blockade therapy against triple negative breast cancer, *Eur J Pharmaceut Sci*, 2021; 157:105606 <https://doi.org/10.1016/j.ejps.2020.105606> PMID:33131745
117. Zhang K, Fang X, You Q, Lin Y, Ma L, Xu S, Ge Y, Xu H, Yang Y, Wang C, Novel peptide-directed liposomes for targeted combination therapy of breast tumors, *Mater Adv*, 2020; 1:3483-3495 <https://doi.org/10.1039/D0MA00536C>

118. Lukyanov AN, Elbayoumi TA, Chakilam AR, Torchilin VP, Tumor-targeted liposomes: doxorubicin-loaded long-circulating liposomes modified with anticancer antibody, *J Contr Release*, 2004; 100:135-144 <https://doi.org/10.1016/j.jconrel.2004.08.007> PMID:15491817
119. Elbayoumi T, Torchilin V, Enhanced cytotoxicity of monoclonal anticancer antibody 2C5-modified doxorubicin-loaded PEGylated liposomes against various tumor cell lines, *Eur J Pharmaceut Sci*, 2007; 32:159-168 <https://doi.org/10.1016/j.ejps.2007.05.113> PMID:17707615 PMCID:PMC2151083
120. Elbayoumi T, Torchilin VP, Tumor-targeted nanomedicines: enhanced antitumor efficacy in vivo of doxorubicin-loaded, long-circulating liposomes modified with cancer-specific monoclonal antibody, *Clin Cancer Res*, 2009; 15:1973-1980 <https://doi.org/10.1158/1078-0432.CCR-08-2392> PMID:19276264 PMCID:PMC2762655
121. Narayanaswamy R, Torchilin VP, Targeted delivery of combination therapeutics using monoclonal antibody 2C5-modified immunoliposomes for cancer therapy, *Pharm Res*, 2021; 38:429-450 <https://doi.org/10.1007/s11095-021-02986-1> PMID:33655395
122. Kamoun WS, Kirpotin DB, Huang ZR, Tipparaju SK, Noble CO, Hayes ME, Luus L, Koshkaryev A, Kim J, Olivier K, Kornaga T, Oyama S, Askoxylakis V, Pien C, Kuesters G, Dumont N, Lugovskoy AA, Schihl SA, Wilton JH, Geddie ML, Suchy J, Grabow S, Kohli N, Reynolds CP, Blaydes R, Zhou Y, Sawyer AJ, Marks JD, Drummond DC, Antitumor activity and tolerability of an EphA2-targeted nanotherapeutic in multiple mouse models, *Nat Biomed Eng*, 2019; 3:264-280 <https://doi.org/10.1038/s41551-019-0385-4> PMID:30952988
123. Kamoun WS, Dugast A-S, Suchy JJ, Grabow S, Fulton RB, Sampson JF, Luus L, Santiago M, Koshkaryev A, Sun G, Askoxylakis V, Tam E, Huang ZR, Drummond DC, Sawyer AJ, Synergy between EphA2-ILs-DTXp, a novel EphA2-targeted nanoliposomal taxane, and PD-1 inhibitors in preclinical tumor models, *Mol Cancer Therapeut*, 2020; 19:270-281 <https://doi.org/10.1158/1535-7163.MCT-19-0414> PMID:31597714
124. Guo Z, He B, Yuan L, Dai W, Zhang H, Wang X, Wang J, Zhang X, Zhang Q, Dual targeting for metastatic breast cancer and tumor neovasculature by EphA2-mediated nanocarriers, *Int J Pharm*, 2015; 493:380-389 <https://doi.org/10.1016/j.ijpharm.2015.05.051> PMID:26004003
125. Barbosa MV, Monteiro LOF, Carneiro G, Malagutti AR, Vilela JMC, Andrade MS, Oliveira MC, Carvalho-Junior AD, Leite EA, Experimental design of a liposomal lipid system: a potential strategy for paclitaxel-based breast cancer treatment, *Colloids Surf B Biointerfaces*, 2015; 136:553-561 <https://doi.org/10.1016/j.colsurfb.2015.09.055> PMID:26454545
126. Monteiro LOF, Fernandes RS, Oda CMR, Lopes SC, Townsend DM, Cardoso VN, Oliveira MC, Leite EA, Rubello D, Barros ALB, Paclitaxel-loaded folate-coated long circulating and pH-sensitive liposomes as a potential drug delivery system: a biodistribution study, *Biomed Pharmacother*, 2018; 97:489-495 <https://doi.org/10.1016/j.biopha.2017.10.135> PMID:29091899 PMCID:PMC6361139
127. de Oliveira Silva J, Fernandes RS, Ramos Oda CM, Ferreira TH, Machado Botelho AF, Martins Melo M, de Miranda MC, Assis Gomes D, Dantas Cassali G, Townsend DM, Rubello D, Oliveira MC, Barros ALB, Folate-coated, long-circulating and pH-sensitive liposomes enhance doxorubicin antitumor effect in a breast cancer animal model, *Biomed Pharmacother*, 2019; 118:109323 <https://doi.org/10.1016/j.biopha.2019.109323> PMID:31400669 PMCID:PMC7104811
128. Chen Y, Cheng Y, Zhao P, Zhang S, Li M, He C, Zhang X, Yang T, Yan R, Ye P, Ma X, Xiang G, Co-delivery of doxorubicin and imatinib by pH sensitive cleavable PEGylated nanoliposomes with folate-mediated targeting to overcome multidrug resistance, *Int J Pharm*, 2018; 542:266-279 <https://doi.org/10.1016/j.ijpharm.2018.03.024> PMID:29551747
129. Soe ZC, Thapa RK, Ou W, Gautam M, Nguyen HT, Jin SG, Ku SK, Oh KT, Choi HG, Yong CS, Kim JO, Folate receptor-mediated celestrol and irinotecan combination delivery using liposomes for effective chemotherapy, *Colloids Surf B Biointerfaces*, 2018; 170:718-728 <https://doi.org/10.1016/j.colsurfb.2018.07.013> PMID:30005409
130. Du Nguyen V, Min HK, Kim CS, Han J, Park JO, Choi E, Folate receptor-targeted liposomal nanocomplex for effective synergistic photothermal-chemotherapy of breast cancer in vivo, *Colloids Surf B Biointerfaces*, 2019; 173:539-548 <https://doi.org/10.1016/j.colsurfb.2018.10.013> PMID:30343218
131. Sneider A, Jadia R, Piel B, Van Dyke D, Tsiros C, Rai P, Engineering remotely triggered liposomes to target triple negative breast cancer, *Oncomedicine*, 2017; 2:1-13 <https://doi.org/10.7150/oncm.17406> PMID:28174679 PMCID:PMC5292187
132. Gazzano E, Rolando B, Chegave K, Salaroglio C, Kopecka J, Pedrini I, Saponara S, Sorge M, Buondonno I, Stella B, Marengo A, Valoti M, Brancaccio M, Fruttero R, Gasco A, Arpicco S, Riganti C, Folate-targeted liposomal nitrooxy-doxorubicin: an effective tool against P-glycoprotein-positive and folate receptor-positive tumors, *J Contr Release*, 2018; 270:37-52 <https://doi.org/10.1016/j.jconrel.2017.11.042> PMID:29191785
133. Deng C, Zhang Q, Jia M, Zhao J, Sun X, Gong T, Zhang Z, Tumors and their microenvironment dual-targeting chemotherapy with local immune adjuvant therapy for effective antitumor immunity against breast cancer, *Adv Sci*, 2019; 6:1801868 <https://doi.org/10.1002/adv.201801868> PMID:30937266 PMCID:PMC6425447
134. Guo P, Yang J, Jia D, Moses MA, Auguste DT, ICAM-1-targeted, Lcn2 siRNA-encapsulating liposomes are potent anti-angiogenic agents for triple negative breast cancer, *Theranostics*, 2016; 6:1-15 <https://doi.org/10.7150/thno.12167> PMID:26722369 PMCID:PMC4679350
135. Orthmann A, Zeisig R, Süß R, Lorenz D, Lemm M, Fichtner I, Treatment of experimental brain metastasis with MTO-liposomes: impact of fluidity and LRP-targeting on the therapeutic result, *Pharm Res*, 2012; 29:1949-1959 <https://doi.org/10.1007/s11095-012-0723-7> PMID:22399388
136. Orthmann A, Peiker L, Fichtner I, Hoffmann A, Hilger R, Zeisig R, Improved treatment of MT-3 breast cancer and brain metastases in a mouse xenograft by LRP-targeted oxaliplatin liposomes, *J Biomed Nanotechnol*, 2016; 12:56-68 <https://doi.org/10.1166/jbn.2016.2143> PMID:27301172
137. Moura V, Lacerda M, Figueiredo P, Corvo M, Cruz M, Soares R, de Lima M, Simoes J, Moreira J, Targeted and intracellular triggered delivery of therapeutics to cancer cells and the tumor microenvironment: impact on the treatment of breast cancer, *Breast Cancer Res Treat*, 2012; 133:61-73 <https://doi.org/10.1007/s10549-011-1688-7> PMID:21805188
138. Fonseca NA, Rodrigues AS, Rodrigues-Santos P, Alves V, Gregorio AC, Valério-Fernandes Á, Gomes-da-Silva LC, Rosa MS, Moura V, Ramalho-Santos J, Simoes J, Moreira JN, Nucleolin overexpression in breast cancer cell sub-populations with different stem-like phenotype enables targeted intracellular delivery of synergistic drug combination, *Biomaterials*, 2015; 69:76-88 <https://doi.org/10.1016/j.biomaterials.2015.08.007> PMID:26283155
139. Xing H, Tang L, Yang X, Hwang K, Wang W, Yin Q, Wong N, Dobrucki L, Yasui N, Katzenellenbogen J, Helferich W, Cheng J, Lu Y, Selective delivery of an anticancer drug with aptamer-functionalized liposomes to breast cancer cells in vitro and in vivo, *J Mater Chem B*, 2013; 1:5288-5297 <https://doi.org/10.1039/c3tb20412j> PMID:24159374 PMCID:PMC3800741
140. Liao ZX, Chuang EY, Lin CC, Ho YC, Lin KJ, Cheng PY, Chen KJ, Wei HJ, Sung HW, An AS1411 aptamer-conjugated liposomal system containing a bubble-generating agent for tumor-specific chemotherapy that overcomes multidrug resistance, *J Contr Release*, 2015; 208:42-51 <https://doi.org/10.1016/j.jconrel.2015.01.032> PMID:25637705
141. Li X, Wu X, Yang H, Li L, Ye Z, Rao Y, A nuclear targeted Dox-aptamer loaded liposome delivery platform for the circumvention of drug resistance in breast cancer, *Biomed Pharmacother*, 2019;

- 117:109072 <https://doi.org/10.1016/j.biopha.2019.109072> PMID:31202169
142. Yu S, Bi X, Yang L, Wu S, Yu Y, Jiang B, Zhang A, Lan K, Duan S, Co-delivery of paclitaxel and PLK1-targeted siRNA using aptamer-functionalized cationic liposome for synergistic anti-breast cancer effects in vivo, *J Biomed Nanotechnol*, 2019; 15:1135-1148 <https://doi.org/10.1166/jbn.2019.2751> PMID:31072423
143. Abnous K, Danesh NM, Ramezani M, Alibolandi M, Bahreyni A, Lavaee P, Moosavian SA, Taghdisi SM, A smart ATP-responsive chemotherapy drug-free delivery system using a DNA nanostructure for synergistic treatment of breast cancer in vitro and in vivo, *J Drug Target*, 2020; 28:852-859 <https://doi.org/10.1080/1061186X.2020.1712407> PMID:31916879
144. Shi J, Sun M, Li X, Zhao Y, Ju R, Mu L, Yan Y, Li X, Zeng F, Lu W, A combination of targeted sunitinib liposomes and targeted vinorelbine liposomes for targeted invasive breast cancer, *J Biomed Nanotechnol*, 2015; 11:1568-1582 <https://doi.org/10.1166/jbn.2015.2075> PMID:26485927
145. Han S, Baek J, Kim M, Hwang S, Cho C, Surface modification of paclitaxel-loaded liposomes using d- α -tocopheryl polyethylene glycol 1000 succinate: enhanced cellular uptake and cytotoxicity in multidrug resistant breast cancer cells, *Chem Phys Lipids*, 2018; 213:39-47 <https://doi.org/10.1016/j.chemphyslip.2018.03.005> PMID:29550143
146. Li N, Fu T, Fei W, Han T, Gu X, Hou Y, Liu Y, Yang J, Vitamin E D- α -tocopheryl polyethylene glycol 1000 succinate-conjugated liposomal docetaxel reverses multidrug resistance in breast cancer cells, *J Pharm Pharmacol*, 2019; 71:1243-1254 <https://doi.org/10.1111/jphp.13126> PMID:31215039
147. Bharti R, Dey G, Banerjee I, Dey KK, Parida S, Kumar BNP, Das CK, Pal I, Mukherjee M, Misra M, Pradhan AK, Emdad L, Das SK, Fisher PB, Mandal M, Somatostatin receptor targeted liposomes with Diacerein inhibit IL-6 for breast cancer therapy, *Cancer Lett*, 2017; 388:292-302 <https://doi.org/10.1016/j.canlet.2016.12.021> PMID:28025102
148. Ju RJ, Cheng L, Peng XM, Wang T, Li CQ, Song XL, Liu S, Chao JP, Li XT, Octreotide-modified liposomes containing daunorubicin and dihydroartemisinin for treatment of invasive breast cancer, *Artif Cell Nanomed Biotechnol*, 2018; 46:616-628 <https://doi.org/10.1080/21691401.2018.1433187> PMID:29381101
149. Gote V, Pal D, Octreotide-targeted Lcn2 siRNA PEGylated liposomes as a treatment for metastatic breast cancer, *Bioengineering*, 2021; 8:44 <https://doi.org/10.3390/bioengineering8040044> PMID:33916786 PMID:PMC8067132
150. Mukherjee A, Prasad TK, Rao NM, Banerjee R, Haloperidol-associated stealth liposomes: a potent carrier for delivering genes to human breast cancer cells, *J Biol Chem*, 2005; 280:15619-15627 <https://doi.org/10.1074/jbc.M409723200> PMID:15695518
151. Zhang Y, Huang Y, Zhang P, Gao X, Gibbs RB, Li S, Incorporation of a selective sigma-2 receptor ligand enhances uptake of liposomes by multiple cancer cells, *Int J Nanomed*, 2012; 7:4473 <https://doi.org/10.2147/IJN.S31981> PMID:22927761 PMID:PMC3422102
152. Gandhi R, Khatri N, Baradia D, Vhora I, Misra A, Surface-modified Epirubicin HCl liposomes and its in vitro assessment in breast cancer cell-line: MCF-7, *Drug Deliv*, 2015; 23:1152-1162 <https://doi.org/10.3109/10717544.2014.999960> PMID:25586675
153. Fu J, Li W, Xin X, Chen D, Hu H, Transferrin-modified nanoliposome co-delivery strategies for enhancing the cancer therapy, *J Pharm Sci*, 2020; 109:2426-2436 <https://doi.org/10.1016/j.xphs.2019.11.013> PMID:31760084
154. Belfiore L, Saunders D, Ranson M, Vine K, N-Alkylisatin-loaded liposomes target the urokinase plasminogen activator system in breast cancer, *Pharmaceutics*, 2020; 12:641 <https://doi.org/10.3390/pharmaceutics12070641> PMID:32645963 PMID:PMC7408009
155. Cochran BJ, Croucher DR, Lobov S, Saunders DN, Ranson M, Dependence on endocytic receptor binding via a minimal binding motif underlies the differential prognostic profiles of SerpinE1 and SerpinB2 in cancer, *J Biol Chem*, 2011; 286:24467-24475 <https://doi.org/10.1074/jbc.M111.225706> PMID:21606492 PMID:PMC3129226
156. Stutchbury TK, Al-ejeh F, Stillfried GE, Croucher DR, Andrews J, Irving D, Links M, Ranson M, Preclinical evaluation of 213Bi-labeled plasminogen activator inhibitor type 2 in an orthotopic murine xenogenic model of human breast carcinoma, *Mol Cancer Therapeut*, 2007; 6:203-212 <https://doi.org/10.1158/1535-7163.MCT-06-0264> PMID:17237280
157. Lu R, Zhou L, Yue Q, Liu Q, Cai X, Xiao W, Hai L, Guo L, Wu Y, Liposomes modified with double-branched biotin: a novel and effective way to promote breast cancer targeting, *Bioorg Med Chem*, 2019; 27:3115-3127 <https://doi.org/10.1016/j.bmc.2019.05.039> PMID:31155297
158. Tang B, Peng Y, Yue Q, Pu Y, Li R, Zhao Y, Hai L, Guo L, Wu Y, Design, preparation and evaluation of different branched biotin modified liposomes for targeting breast cancer, *Eur J Med Chem*, 2020; 193:112204 <https://doi.org/10.1016/j.ejmech.2020.112204> PMID:32172035
159. Huang M, Pu Y, Peng Y, Fu Q, Guo L, Wu Y, Zheng Y, Biotin and glucose dual-targeting, ligand-modified liposomes promote breast tumor-specific drug delivery, *Bioorg Med Chem Lett*, 2020; 30:127151 <https://doi.org/10.1016/j.bmcl.2020.127151> PMID:32317211
160. Lv Y, Xu C, Zhao X, Lin C, Yang X, Xin X, Zhang L, Qin C, Han X, Yang L, He W, Yin L, Nanoplatform assembled from a CD44-targeted prodrug and smart liposomes for dual targeting of tumor microenvironment and cancer cells, *ACS Nano*, 2018; 12:1519-1536 <https://doi.org/10.1021/acsnano.7b08051> PMID:29350904
161. Han NK, Shin DH, Kim JS, Weon KY, Jang CY, Kim JS, Hyaluronan-conjugated liposomes encapsulating gemcitabine for breast cancer stem cells, *Int J Nanomed*, 2016; 11:1413-1426 <https://doi.org/10.2147/IJN.S95850> PMID:27103799 PMID:PMC4827594
162. Jiang T, Mo R, Bellotti A, Zhou J, Gu Z, Gel-liposome-mediated co-delivery of anticancer membrane-associated proteins and small-molecule drugs for enhanced therapeutic efficacy, *Adv Funct Mater*, 2014; 24:2295-2304 <https://doi.org/10.1002/adfm.201303222>
163. Ding Y, Yang R, Yu W, Hu C, Zhang Z, Liu D, An Y, Wang X, He C, Liu P, Tang Q, Chen D, Chitosan oligosaccharide decorated liposomes combined with TH302 for photodynamic therapy in triple negative breast cancer, *J Nanobiotechnol*, 2021; 19:1-17 <https://doi.org/10.1186/s12951-021-00891-8> PMID:34011362 PMID:PMC8136194
164. Yang R, Lu M, Ming L, Chen Y, Cheng K, Zhou J, Jiang S, Lin Z, Chen D, 89Zr-labeled multifunctional liposomes conjugate chitosan for PET-trackable triple-negative breast cancer stem cell targeted therapy, *Int J Nanomed*, 2020; 15:9061-9078 <https://doi.org/10.2147/IJN.S262786> PMID:33239874 PMID:PMC7680801
165. He Y, Zhang L, Song C, Luteinizing hormone-releasing hormone receptor-mediated delivery of mitoxantrone using LHRH analogs modified with PEGylated liposomes, *Int J Nanomed*, 2010; 5:697-705 <https://doi.org/10.2147/IJN.S12129> PMID:20957221 PMID:PMC2948949
166. He Y, Zhang L, Zhu D, Song C, Design of multifunctional magnetic iron oxide nanoparticles/mitoxantrone-loaded liposomes for both magnetic resonance imaging and targeted cancer therapy, *Int J Nanomed*, 2014; 9:4055-4066 <https://doi.org/10.2147/IJN.S61880> PMID:25187709 PMID:PMC4149452
167. Paliwal S, Paliwal R, Pal H, Saxena A, Sharma P, Gupta P, Agrawal G, Vyas S, Estrogen-anchored pH-sensitive liposomes as nanomodule designed for site-specific delivery of doxorubicin in breast cancer therapy, *Mol Pharm*, 2012; 9:176-186 <https://doi.org/10.1021/mp200439z> PMID:22091702

168. Salkho NM, Paul V, Kawak P, Vitor RF, Martins AM, Al Sayah M, Husseini GA, Ultrasonically controlled estrone-modified liposomes for estrogen-positive breast cancer therapy, *Artif Cell Nanomed Biotechnol*, 2018; 46:462-472 <https://doi.org/10.1080/21691401.2018.1459634> PMID:29644867
169. Han B, Yang Y, Chen J, Tang H, Sun Y, Zhang Z, Wang Z, Li Y, Li Y, Luan X, Li Q, Ren Z, Zhou X, Cong D, Liu Z, Meng Q, Sun F, Pei J, Preparation, characterization, and pharmacokinetic study of a novel long-acting targeted paclitaxel liposome with antitumor activity, *Int J Nanomed*, 2020; 15:553-571 <https://doi.org/10.2147/IJN.S228715> PMID:32158208 PMCID:PMC6986409
170. Qin C, He B, Dai W, Zhang H, Wang X, Wang J, Zhang X, Wang G, Yin L, Zhang Q, Inhibition of metastatic tumor growth and metastasis via targeting metastatic breast cancer by chlorotoxin-modified liposomes, *Mol Pharm*, 2014; 11:3233-3241 <https://doi.org/10.1021/mp400691z> PMID:24559485
171. strem RG, Parhamifar L, Pourhassan H, Clergeaud G, Nielsen OL, Kjær A, Hansen AE, Andresen TL, Secretory phospholipase A2 responsive liposomes exhibit a potent anti-neoplastic effect in vitro, but induce unforeseen severe toxicity in vivo, *J Contr Release*, 2017; 262:212-221 <https://doi.org/10.1016/j.jconrel.2017.07.031> PMID:28754610
172. Oshiro-Júnior J, Rodero C, Hanck-Silva G, Sato M, Alves R, Eloy J, Chorilli M, Stimuli-responsive drug delivery nanocarriers in the treatment of breast cancer, *Curr Med Chem*, 2020; 27:2494-2513 <https://doi.org/10.2174/0929867325666181009120610> PMID:30306849
173. Müller A, Homey B, Soto H, Ge N, Catron D, Buchanan ME, McClanahan T, Murphy E, Yuan W, Wagner SN, Barrera JL, Mohar A, Verastegui E, Zlotnik A, Involvement of chemokine receptors in breast cancer metastasis, *Nature*, 2001; 410:50-56 <https://doi.org/10.1038/35065016> PMID:11242036
174. Zlotnik A, Chemokines and cancer, *Int J Cancer*, 2006; 119:2026-2029 <https://doi.org/10.1002/ijc.22024> PMID:16671092
175. Bleul CC, Fuhlbrigge RC, Casasnovas JM, Aiuti A, Springer TA, A highly efficacious lymphocyte chemoattractant, stromal cell-derived factor 1 (SDF-1), *J Exp Med*, 1996; 184:1101-1109 <https://doi.org/10.1084/jem.184.3.1101> PMID:9064327 PMCID:PMC2192798
176. Mukherjee D, Zhao J, The role of chemokine receptor CXCR4 in breast cancer metastasis, *Am J Cancer Res*, 2013; 3:46-57
177. Wang Y, Xie Y, Oupický D, Potential of CXCR4/CXCL12 chemokine axis in cancer drug delivery, *Curr Pharmacol Rep*, 2016; 2:1-10 <https://doi.org/10.1007/s40495-015-0044-8> PMID:27088072 PMCID:PMC4827436
178. Robertson JM, James JA, Preclinical systemic lupus erythematosus, *Rheum Dis Clin*, 2014; 40:621-635 <https://doi.org/10.1016/j.rdc.2014.07.004> PMID:25437281 PMCID:PMC4301850
179. Vlasea A, Falagan S, Gutiérrez-Gutiérrez G, Moreno-Rubio J, Merino M, Zambrana F, Casado E, Sereno M, Antinuclear antibodies and cancer: a literature review, *Crit Rev Oncol Hematol*, 2018; 127:42-49 <https://doi.org/10.1016/j.critrevonc.2018.05.002> PMID:29891110
180. Nisihara R, Machoski M, Neppel A, Maestri C, Messias-Reason I, Skare T, Antinuclear antibodies in patients with breast cancer, *Clin Exp Immunol*, 2018; 193:178-182 <https://doi.org/10.1111/cei.13136> PMID:29645079 PMCID:PMC6046476
181. Wilson K, Shiuan E, Brantley-Sieders DM, Oncogenic functions and therapeutic targeting of EphA2 in cancer, *Oncogene*, 2021; 40:2483-2498 <https://doi.org/10.1038/s41388-021-01714-8> PMID:33686241 PMCID:PMC8035212
182. Xiao T, Xiao Y, Wang W, Tang YY, Xiao Z, Su M, Targeting EphA2 in cancer, *J Hematol Oncol*, 2020; 13:1-17 <https://doi.org/10.1186/s13045-020-00944-9> PMID:32811512 PMCID:PMC7433191
183. Zhao P, Jiang D, Huang Y, Chen C, EphA2: a promising therapeutic target in breast cancer, *J Genet Genomics*, 2021; 48:261-267 <https://doi.org/10.1016/j.jgg.2021.02.011> PMID:33962882
184. Ernstoff MS, Ma WW, Tsai FYC, Munster PN, Zhang T, Kamoun W, Pipas JM, Chen S, Santillana S, Askoxylakis V, A phase 1 study evaluating the safety, pharmacology and preliminary activity of MM-310 in patients with solid tumors, *J Clin Oncol*, 2018; 36:TPS2604 https://doi.org/10.1200/JCO.2018.36.15_suppl.TPS2604
185. Kamoun WS, Dugast AS, Suchy JJ, Grabow S, Fulton RB, Sampson JF, Luus L, Santiago M, Koshkaryev A, Sun G, Askoxylakis V, Tam E, Huang ZR, Drummond DC, Sawyer AJ, Synergy between EphA2-ILs-DTXp, a novel EphA2-targeted nanoliposomal taxane, and PD-1 inhibitors in preclinical tumor models, *Mol Cancer Therapeut*, 2020; 19:270-281 <https://doi.org/10.1158/1535-7163.MCT-19-0414> PMID:31597714
186. Kumar P, Huo P, Liu B, Formulation strategies for folate-targeted liposomes and their biomedical applications, *Pharmaceutics*, 2019; 11:381 <https://doi.org/10.3390/pharmaceutics11080381> PMID:31382369 PMCID:PMC6722551
187. Xu L, Bai Q, Zhang X, Yang H, Folate-mediated chemotherapy and diagnostics: an updated review and outlook, *J Contr Release*, 2017; 252:73-82 <https://doi.org/10.1016/j.jconrel.2017.02.023> PMID:28235591 PMCID:PMC5479736
188. Tagde P, Kulkarni GT, Mishra DK, Kesharwani P, Recent advances in folic acid engineered nanocarriers for treatment of breast cancer, *J Drug Deliv Sci Technol*, 2020; 56:101613 <https://doi.org/10.1016/j.jddst.2020.101613> 189. Zempleni J, Wijeratne S, Hassan Y, Biotin, *BioFactors*, 2009; 35:36-46 <https://doi.org/10.1002/biof.8> PMID:19319844 PMCID:PMC4757853
190. Vadlapudi A, Vadlapatla R, Mitra A, Sodium dependent multivitamin transporter (SMVT): a potential target for drug delivery, *Curr Drug Targets*, 2012; 13:994-1003 <https://doi.org/10.2174/138945012800675650> PMID:22420308 PMCID:PMC4406285
191. Vadlapudi AD, Vadlapatla RK, Pal D, Mitra AK, Biotin uptake by T47D breast cancer cells: functional and molecular evidence of sodium-dependent multivitamin transporter (SMVT), *Int J Pharm*, 2013; 441:535-543 <https://doi.org/10.1016/j.ijpharm.2012.10.047> PMID:23142496
192. Ren WX, Han J, Uhm S, Jang YJ, Kang C, Kim JH, Kim JS, Recent development of biotin conjugation in biological imaging, sensing, and target delivery, *Chem Commun*, 2015; 51:10403-10418 <https://doi.org/10.1039/C5CC03075G> PMID:26021457
193. Zheng Z, Shao N, Weng H, Li W, Zhang J, Zhang L, Yang L, Ye S, Correlation between epidermal growth factor receptor and tumor stem cell markers CD44/CD24 and their relationship with prognosis in breast invasive ductal carcinoma, *Med Oncol*, 2015; 32:1-11 <https://doi.org/10.1007/s12032-014-0275-2> PMID:25429827 PMCID:PMC4246130
194. Jin J, Krishnamachary B, Mironchik Y, Kobayashi H, Bhujwalla ZM, Phototheranostics of CD44-positive cell populations in triple negative breast cancer, *Sci Rep*, 2016; 6:1-12 <https://doi.org/10.1038/srep27871> PMID:27302409 PMCID:PMC4908597
195. Yang C, He Y, Zhang H, Liu Y, Wang W, Du Y, Gao F, Selective killing of breast cancer cells expressing activated CD44 using CD44 ligand-coated nanoparticles in vitro and in vivo, *Oncotarget*, 2015; 6:15283-15296 <https://doi.org/10.18632/oncotarget.3681> PMID:25909172 PMCID:PMC4558151
196. Gupta R, Lall R, Srivastava A, Sinha A, Hyaluronic acid: molecular mechanisms and therapeutic trajectory, *Front Vet Sci*, 2019; 6:192 <https://doi.org/10.3389/fvets.2019.00192> PMID:31294035 PMCID:PMC6603175
197. Lee HJ, Seo AN, Kim EJ, Jang MH, Kim YJ, Kim JH, Kim SW, Ryu HS, Park IA, Im SA, Gong G, Jung KH, Kim HJ, Park SY, Prognostic and predictive values of EGFR overexpression and EGFR copy number alteration in HER2-positive breast cancer, *Br J Cancer*, 2015; 112:103-111 <https://doi.org/10.1038/bjc.2014.556> PMID:25349977 PMCID:PMC4453607

198. Luiz MT, Dutra JAP, Tofani LB, de Araújo JTC, Di Filippo LD, Marchetti JM, Chorilli M, Targeted liposomes: a nonviral gene delivery system for cancer therapy, *Pharmaceutics*, 2022; 14:821 <https://doi.org/10.3390/pharmaceutics14040821> PMID:35456655 PMCID:PMC9030342
199. Bou-Assaly W, Mukherji S, Cetuximab (Erbix), *Am J Neuroradiol*, 2010; 31:626-627 <https://doi.org/10.3174/ajnr.A2054> PMID:20167650 PMCID:PMC7964212
200. Flanagan CA, Manilall A, Gonadotropin-releasing hormone (GnRH) receptor structure and GnRH binding, *Front Endocrinol*, 2017; 8:274 <https://doi.org/10.3389/fendo.2017.00274> PMID:29123501 PMCID:PMC5662886
201. Schally AV, Szepeshazi K, Nagy A, Comaru-Schally AM, Halmos G, New approaches to therapy of cancers of the stomach, colon and pancreas based on peptide analogs, *Cell Mol Life Sci*, 2004; 61:1042-1068 <https://doi.org/10.1007/s00018-004-3434-3> PMID:15112052 PMCID:PMC11138622
202. Wu Y, Zhang Z, Cenciarini ME, Proietti CJ, Amasino M, Hong T, Yang M, Liao Y, Chiang HC, Kaklamani VG, Jeselsohn R, Vadlamudi RK, Huang THM, Li R, De Angelis C, Fu X, Elizalde PV, Schiff R, Brown M, Xu K, Tamoxifen resistance in breast cancer is regulated by the EZH2-ER α -GREB1 transcriptional axis, *Cancer Res*, 2018; 78:671-684 <https://doi.org/10.1158/0008-5472.CAN-17-1327> PMID:29212856 PMCID:PMC5967248
203. Jallow F, O'Leary KA, Rugowski DE, Guerrero JF, Ponik SM, Schuler LA, Dynamic interactions between the extracellular matrix and estrogen activity in progression of ER+ breast cancer, *Oncogene*, 2019; 38:6913-6925 <https://doi.org/10.1038/s41388-019-0941-0> PMID:31406251 PMCID:PMC6814534
204. Liu Y, Ma H, Yao J, ER α , a key target for cancer therapy: a review, *OncoTargets Ther*, 2020; 13:2183-2191 <https://doi.org/10.2147/OTT.S236532> PMID:32210584 PMCID:PMC7073439
205. Omoto Y, Iwase H, Clinical significance of estrogen receptor β in breast and prostate cancer from biological aspects, *Cancer Sci*, 2015; 106:337-343 <https://doi.org/10.1111/cas.12613> PMID:25611678 PMCID:PMC4409875
206. Arnal JF, Lenfant F, Metivier R, Flouriot G, Henrion D, Adlanmerini M, Fontaine C, Gourdy P, Chambon P, Katzenellenbogen B, Katzenellenbogen J, Membrane and nuclear estrogen receptor alpha actions: from tissue specificity to medical implications, *Physiol Rev*, 2017; 97:1045-1087 <https://doi.org/10.1152/physrev.00024.2016> PMID:28539435
207. Spooner D, Litton A, Chlebowski RT, Caffier H, Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials, *Lancet*, 2005; 365:1687-1717 [https://doi.org/10.1016/S0140-6736\(05\)66544-0](https://doi.org/10.1016/S0140-6736(05)66544-0) PMID:15894097
208. Yan C, Boyd D, Regulation of matrix metalloproteinase gene expression, *J Cell Physiol*, 2007; 211:19-26 <https://doi.org/10.1002/jcp.20948> PMID:17167774
209. Cathcart J, Pulkoski-Gross A, Cao J, Targeting matrix metalloproteinases in cancer: bringing new life to old ideas, *Genes Dis*, 2015; 2:26-34 <https://doi.org/10.1016/j.gendis.2014.12.002> PMID:26097889 PMCID:PMC4474140
210. Alaseem A, Alhazzani K, Dondapati P, Alobid S, Bishayee A, Rathinavelu A, Matrix metalloproteinases: a challenging paradigm of cancer management, *Semin Cancer Biol*, 2019; 56:100-115 <https://doi.org/10.1016/j.semcancer.2017.11.008> PMID:29155240
211. Medina OP, Haikola M, Tahtinen M, Simpura I, Kaukinen S, Valtanen H, Zhu Y, Kuosmanen S, Cao W, Reunanen J, Nurminen T, Saris PEJ, Smith-Jones P, Bradbury M, Larson S, Kairemo K, Liposomal tumor targeting in drug delivery utilizing MMP-2- and MMP-9-binding ligands, *J Drug Deliv*, 2011; 2011:1-9 <https://doi.org/10.1155/2011/160515> PMID:21490745 PMCID:PMC3066593
212. Isaacson KJ, Jensen MM, Subrahmanyam NB, Ghandehari H, Matrix metalloproteinases as targets for controlled delivery in cancer: an analysis of upregulation and expression, *J Contr Release*, 2017; 259:62-72 <https://doi.org/10.1016/j.jconrel.2017.01.034> PMID:28153760 PMCID:PMC5537048
213. Quach ND, Arnold RD, Cummings BS, Secretory phospholipase A2 enzymes as pharmacological targets for treatment of disease, *Biochem Pharmacol*, 2014; 90:338-348 <https://doi.org/10.1016/j.bcp.2014.05.022> PMID:24907600 PMCID:PMC4104246
214. Yamashita S, Yamashita J, Ogawa M, Overexpression of group II phospholipase A2 in human breast cancer tissues is closely associated with their malignant potency, *Br J Cancer*, 1994; 69:1166-1170 <https://doi.org/10.1038/bjc.1994.229> PMID:8198986 PMCID:PMC1969450
215. Dennis E, Cao J, Hsu Y, Magriotti V, Kokotos G, Phospholipase A2 enzymes: physical structure, biological function, disease implication, chemical inhibition, and therapeutic intervention, *Chem Rev*, 2011; 111:6130-6185 <https://doi.org/10.1021/cr200085w> PMID:21910409 PMCID:PMC3196595
216. Jespersen SS, Stovgaard ES, Nielsen D, Christensen TD, Buhl ASK, Christensen IJ, Balslev E, Expression of secretory phospholipase A2 group IIa in breast cancer and correlation to prognosis in a cohort of advanced breast cancer patients, *Appl Immunohistochem Mol Morphol*, 2021; 29:E5-E9 <https://doi.org/10.1097/PAI.0000000000000854> PMID:32217848
217. André T, Boni C, Mounedji-Boudiaf L, Navarro M, Tabernero J, Hickish T, Topham C, Zaninelli M, Clingan P, Bridgewater J, Tabah-Fisch I, de Gramont A, Oxaliplatin, fluorouracil, and leucovorin as adjuvant treatment for colon cancer, *N Engl J Med*, 2009; 350:2343-2351 <https://doi.org/10.1056/NEJMoa032709> PMID:15175436
218. Zhang J, Wang L, Wang Z, Hu X, Wang B, Cao J, Lv F, Zhen C, Zhang S, Shao Z, A phase II trial of biweekly vinorelbine and oxaliplatin in second- or third-line metastatic triple-negative breast cancer, *Cancer Biol Ther*, 2015; 16:225-232 <https://doi.org/10.4161/15384047.2014.986973> PMID:25648299 PMCID:PMC4622533
219. Garufi C, Nistico C, Brienza S, Vaccaro A, D'Ottavio A, Zappalà AR, Aschelter AM, Terzoli E, Single-agent oxaliplatin in pretreated advanced breast cancer patients: a phase II study, *Ann Oncol*, 2001; 12:179-182 <https://doi.org/10.1023/A:1008386419047> PMID:11300320
220. Fei F, Chen C, Xue J, Di GH, Lu JS, Liu GY, Shao ZM, Wu J, Efficacy and safety of docetaxel combined with oxaliplatin as a neoadjuvant chemotherapy regimen for Chinese triple-negative local advanced breast cancer patients: a prospective, open, and unicentric phase II clinical trial, *Am J Clin Oncol Cancer Clin Trials*, 2013; 36:545-551 <https://doi.org/10.1097/COC.0b013e31825d5317> PMID:22868245
221. Zelek L, Cottu P, Tubiana-Hulin M, Vannetzel JM, Chollet Ph, Misset JL, Chouaki N, Marty M, Gamelin E, Culine S, Dieras V, Mackenzie S, Spielmann M, Phase II study of oxaliplatin and fluorouracil in taxane- and anthracycline-pretreated breast cancer patients, *J Clin Oncol*, 2002; 20:2551-2558 <https://doi.org/10.1200/JCO.2002.06.164> PMID:12011135
222. De Jonge MJA, Slingerland M, Loos WJ, Wiemer EAC, Burger H, Mathijssen RHJ, Kroep JR, Den Hollander MAG, Van Der Biessen D, Lam MH, Verweij J, Gelderblom H, Early cessation of the clinical development of LiPlaCis, a liposomal cisplatin formulation, *Eur J Cancer*, 2010; 46:3016-3021 <https://doi.org/10.1016/j.ejca.2010.07.015> PMID:20801016
223. Pourhassan H, Clergeaud G, Hansen A, Østrem R, Fliedner F, Melander F, Nielsen O, O'Sullivan C, Kjær A, Andresen TL, Revisiting the use of sPLA2-sensitive liposomes in cancer therapy, *J Contr Release*, 2017; 261:163-173 <https://doi.org/10.1016/j.jconrel.2017.06.024> PMID:28662900

224. Lassen U, Mau-Sørensen M, Buhl UH, Madsen MW, Balslev E, Pluim D, Schellens JHM, Knudsen S, Jensen PB, Phase I dose-escalating PoC study to evaluate the safety and tolerability of LiPlaCis (liposomal cisplatin formulation) in patients with advanced or refractory tumors, *Cancer Res*, 2016; 76:CT154-CT154 <https://doi.org/10.1158/1538-7445.AM2016-CT154>
225. Lassen UN, Knudsen S, Hertel PB, Kumler I, Nielsen D, Ejlertsen B, Mau-Sørensen MM, Brunner N, Buhl UH, Madsen MW, Buhl IK, Hansen A, Jensen T, Balslev E, Askaa J, Vestlev PM, Laenholm AV, Jensen PB, Use of microRNA to identify stage IV breast cancer patients to be targeted with phospholipase A2 disrupted cisplatin carrying liposomes: an ongoing phase I trial, *J Clin Oncol*, 2014; 32:TPS1139-TPS1139 https://doi.org/10.1200/jco.2014.32.15_suppl.tps1139
226. Jakobsen EH, Nielsen D, Danoe H, Linnet S, Hansen J, Lassen UN, Balslev E, Glavicic V, Bogovic J, Knudsen S, Ejlertsen B, Knoop ASK, Buhl UH, Madsen MW, Buhl IK, Hansen A, Jensen T, Rasmussen A, Jensen PB, Langkjer ST, Liposomal cisplatin response prediction in heavily pretreated breast cancer patients: a multigene biomarker in a prospective phase 2 study, *J Clin Oncol*, 2018; 36:e13077 https://doi.org/10.1200/JCO.2018.36.15_suppl.e13077