



## Phytochemical Profile, TLC Profiling and Antioxidant Potential of *Prunus Avium*

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### Abstract

This work looks at the Chromatographic content of the hydro-alcoholic extract of *Prunus avium* leaf and its antioxidant activity. The presence of essential metabolites was confirmed by preliminary phytochemical screening for glycosides, flavonoids, alkaloids, steroids, volatile oils, and tannins, indicating the plant's ethnomedicinal value. To maximise the separation and visibility of constituent substances, TLC was done using three solvent systems of varied polarity. Under 254 nm and 365 nm UV light, distinct well-resolved bands were observed, indicating a different range of polar and mid-polar phytochemicals. Further antioxidant activity was assessed using DPPH and ABTS radical scavenging assays. The extensive chromatographic and phytochemical profiles obtained in the study highlight the potential of *P. avium* as a valid phytotherapeutic candidate, setting the stage for future bioactive-guided reclusion and anticancer testing.

**Keywords:** *Prunus avium*, sweet cherry, Chromatographic content, TLC analysis, hydroalcoholic extract.

## 1. INTRODUCTION

*Prunus* belongs to the Rosaceae family, specifically the Amygdalaceae subfamily. It includes over 430 species of evergreen and deciduous trees and shrubs, primarily located in the temperate zones of northern hemisphere. *Prunus avium* (Sweet cherry) is a temperate region fruit that is also most popular in regions rich in phytoconstituents, which are responsible for various medicinal properties. They have a lower glycemic index than other fruits and vegetables. Sweet cherry is also said to be a good source of vitamin C & other vitamins also giving them an advantage over other fruits and vegetables. High amounts of water, lower levels of fat, mainly saturated fat, which is also cholesterol-free, and a calorie deficit. The principal phytoconstituents responsible for pharmacological actions in the *Prunus* genus are saponins, sterols, alkaloids, terpenoids, flavonoids, tannins, and phenolic acids <sup>1,2</sup>.

Sweet cherry also contain several phenolic chemicals, including phenolic acids and flavonoids (anthocyanins, flavan-3-ols, and flavanols) and (hydroxycinnamic

derivatives) which have been linked to their antioxidant activity <sup>3,4</sup>. Polyphenols were quantified using HPLC in an ultrasound-assisted *P. avium* extract and Ferulic acid, quercetin, Coumaric, Gallic acid and syringic acids were found <sup>5,6</sup>. *Prunus Avium* includes diverse flavonoids such Aequinoctin, Chrysins, Galangin, Naringenin, Jaceidin, Tectochrysin, Dihydrotectochrysin and Kaempferol, as well as phenolic acids like Caffeic acid, p-Coumeric acid, o-Coumaric acid, and Chlorogenic acid <sup>7</sup>. *Prunus Avium* contains many chemical constituents with varying pharmacological activity, including antioxidant, antimicrobial, anti-fibrotic, anti-tumor, anti-estrogen, and anti-cancer, respectively <sup>1</sup>.

## 2. MATERIALS AND METHODS

### 2.1 Preparation of plant material:

The Leaves of *Prunus avium* were collected from the natural habitat in Prayagraj, Uttar Pradesh, India and then shade dried to grind in a coarse powder.

## 2.2 Process of Extraction for Phytochemical / Phytoconstituent of *Prunus avium*

Extraction of phytoconstituent was done by using the soxhlet apparatus extraction technique from *Prunus avium* leaves. The dried and powdered plant material (about 50 g) was accurately weighed and placed in a thimble made of Whatman filter paper. The thimble was inserted into the Soxhlet extractor attached to a round-bottom flask containing 250 mL of mixture of water and alcohol (50:50)

solvent. The extraction was carried out at the solvent's boiling temperature for 6–8 hours until the solvent in the siphon tube of the extractor became colorless, indicating complete extraction of the phytochemicals. After extraction, the solvent was concentrated under reduced pressure using a rotary evaporator at to obtain a thick, viscous crude extract. The extract was then transferred into a pre-weighed glass dish and kept in a vacuum oven to remove any residual solvent and kept in a desiccator for further use (Figure 1) <sup>8</sup>.

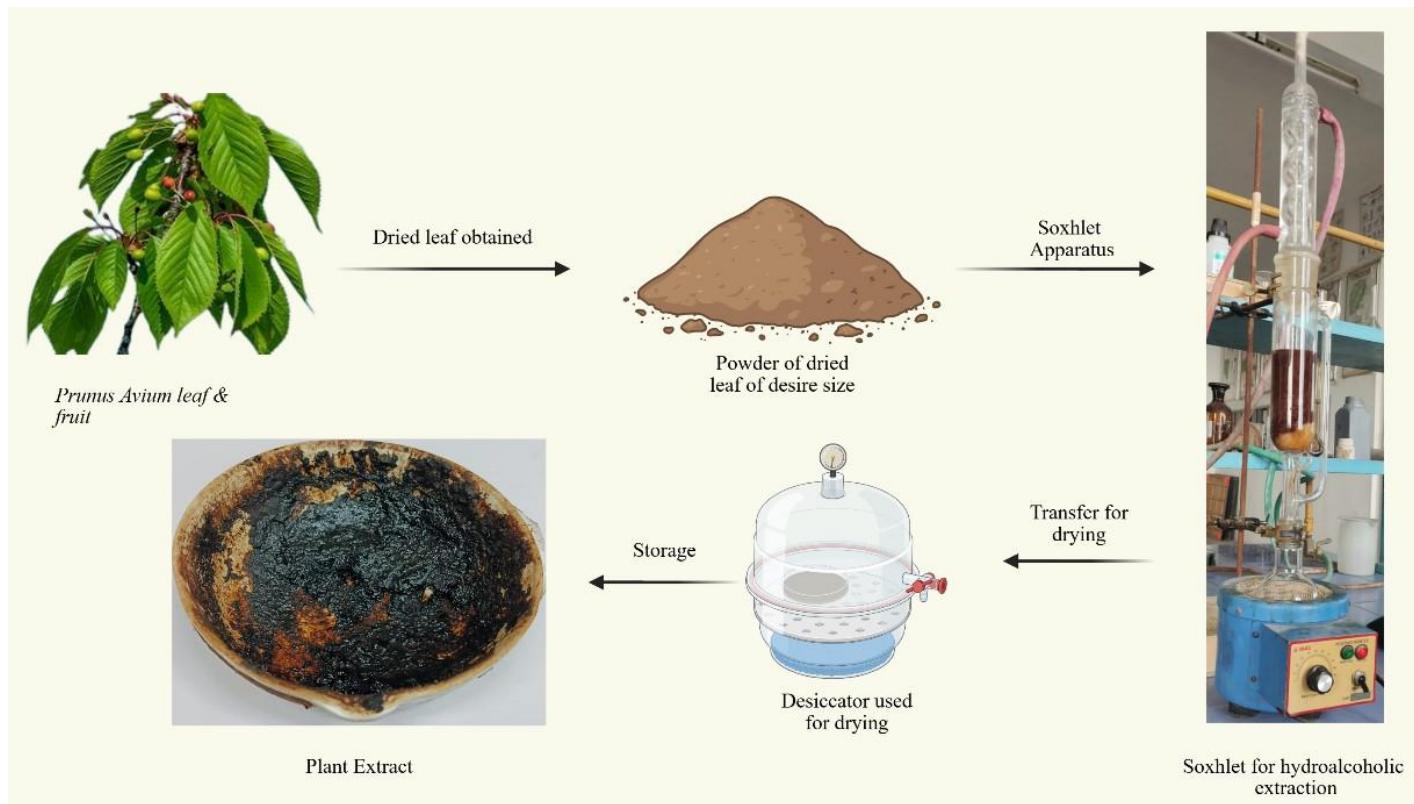


Figure 1: Extraction process of *Prunus avium* leaves extract. <https://BioRender.com>

## 2.3 Phytochemical Analysis

Different phytochemical tests were done on crude hydroalcoholic extract of the leaves of *prunus* to identify the phenols, alkaloids, glycosides, volatile oils, flavonoids, balsams, terpenes, tannins, and resins <sup>9</sup>.

## 2.4 Preparation of Thin layer Chromatography (TLC) plate & profiling of *Prunus avium* extract

To create a fingerprint of phytochemicals found in the hydroalcoholic extract, TLC was done. TLC plates from Merck the stationary phase consisted of pre-coated silica gel were used. A small volume of 1-1.5  $\mu$ L of the hydroalcoholic extract was employed upon the plate by glass capillary, keeping in mind that spot was roughly ~1 cm above from the plate's lower side. To ensure reproducibility and constant migration, spot size and spacing were uniformly distributed among all sample <sup>10,11</sup>.

Three different solvents were taken: 1- toluene: ethyl acetate (7:3, v/v), 2- methanol : ethyl acetate (2:8, v/v) 3- toluene : ethyl acetate (3:7, v/v) <sup>12</sup>. Spots were visualized using UV light of wavelengths of 254 and 365 nm <sup>13,14</sup>.

## 2.5 Antioxidant activity

### 2.5.1 DPPH scavenging activity:

In a 96-well microplate, 5  $\mu$ L of various test compound stock solutions (as detailed in the Excel sheet) were combined with 0.1mL of 0.1mM DPPH solution. The assay was conducted in triplicate, while duplicate blanks were prepared using 0.2mL of DMSO/ Methanol and 5  $\mu$ L of test compounds at different concentration (as listed in the Excel sheet). The plate was then incubated in the dark for 30 minutes. Post incubation, absorbance was measured at 495nm using an iMark Microplate Reader (Bio-Rad) to assess DPPH decolorization. A reaction mixture containing 20  $\mu$ L of deionized water served as the control. The radical scavenging activity was calculated as percent inhibition relative to the control, and the IC<sub>50</sub> values were determined using GraphPad Prism 6 <sup>15,16</sup>.

### 2.5.2 ABTS Radical Scavenging Ability:

ABTS (SRL-Chem-Cat no.-28042) radicles were prepared by mixing APS (2.45 mM) and ABTS (7mM) solution, which was diluted 100X to prepare ABTS free radical reagent. Add 10 $\mu$ L of different stock of the standard (Ascorbic Acid

-SD Fine- F13A/0413/1106/62, Concentration as per mentioned in excel sheet) and samples (As per mention in excel sheet) to the 200 $\mu$ l of ABTS free radical reagent in 96 well plate and incubated at RT for 10 min in dark. After incubation measure absorbance of the decolorization at 750nm using a microplate reader (iMark, BioRad). Results were presented with respect to negative control. IC<sub>50</sub> was calculated using Software Graph Pad Prism 6<sup>17,18</sup>.

### 3 RESULT

#### 3.1 Phytochemical analysis

It showed the diverse classes of phytochemicals as glycosides, Alkaloid, steroids, flavonoids, starch and

saponins in the different chemical tests.

#### 3.2 TLC Profiling

Bright fluorescent spots were visible under 365 nm UV light, which suggested the presence of flavonoids and terpenoids. In the Toluene: Ethyl Acetate (7:3) three spots were seen, methanol: ethyl acetate (2:8) solvent system showed total 5 spots and Toluene: Ethyl Acetate (3:7) showed 9 visible fluorescent spots, range of R<sub>f</sub> values from 0.41 to 0.81, indicating the diverse phytochemical profile, potentially encompassing phenolics compounds and sterols. (Figure 2)

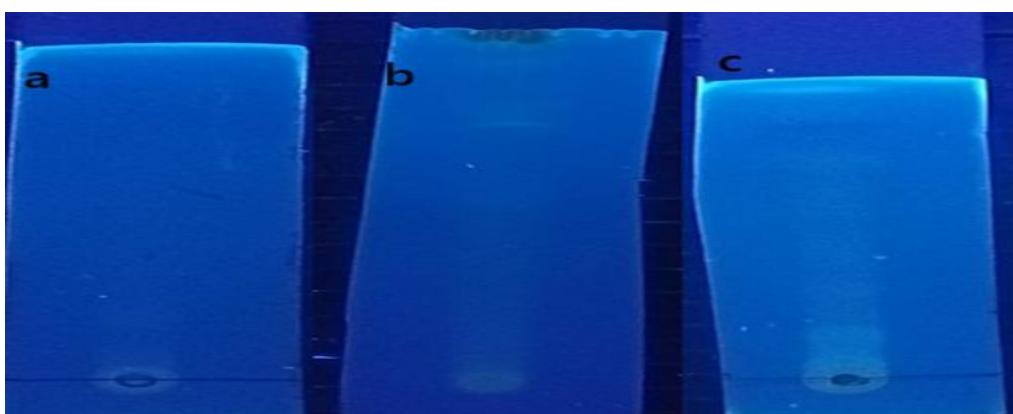


Figure 2: TLC profiling of *Prunus avium* bark employing multiple solvent systems and visualizing under 365nm UV light.  
(a) Toluene: Ethyl Acetate, 7:3 (b) Methanol: Ethyl Acetate, 2:8 (c) Toluene: Ethyl Acetate, 3:7.

#### 3.3 Antioxidant activity

DPPH radicle scavenging activity was observed in *Prunus avium* at IC<sub>50</sub> = 423.1 ± 0.041  $\mu$ g/ml (Figure 3) while ABTS scavenging activity was observed at IC<sub>50</sub> = 148.5 ± 0.92  $\mu$ g/ml (Figure 4).

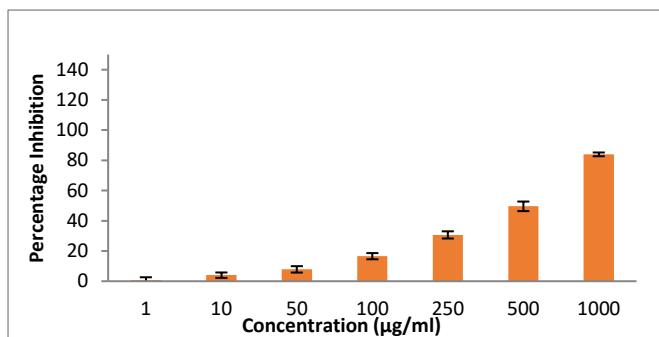


Figure 3: DPPH scavenging activity of *Prunus avium* (Data are represented in Mean ± SEM)

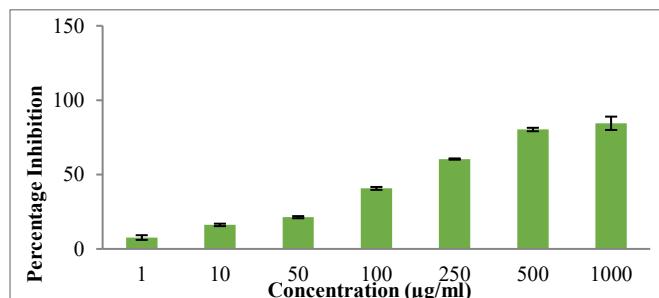


Figure 4: ABTS Radical Scavenging Ability of *Prunus avium* (Data are represented in Mean ± SEM)

### 4. DISCUSSION

The preliminary phytochemical screening of *Prunus avium* bark extract shown the presence of different classes of bioactive secondary plant metabolites, such as glycosides, alkaloids, steroids, flavonoids, starch, and saponins. These bioactive compounds are well known for its pharmacological properties, mostly antioxidant, anti-inflammatory, antimicrobial, and anticancer properties. Flavonoids and saponins, for example, are known to show strong free radical scavenging activity, however steroids and alkaloids have been linked with anti-inflammatory and antimicrobial potential. TLC profile confirmed the chemical variety of *P. avium*. Light color fluorescent spots were seen in 365 nm UV light, suggesting the presence of flavonoids and terpenoids, both. The chromatographic separation using different solvent systems shows TLC profiles: Toluene: Ethyl Acetate (7:3) shows 3 different spots. Methanol: Ethyl Acetate (2:8) shows 5 different spots. Toluene: Ethyl Acetate (3:7) shows 9 different spots. The experimental R<sub>f</sub> values 0.41 to 0.81, shows various phytochemical profile with different polarity. The higher number of spots in the Toluene: Ethyl Acetate (3:7) system signifies that this solvent system provided better resolution meant for separating various components. According to different literature, phenolic compounds, flavonoids, terpenoids and sterols are associated with antioxidant activity<sup>19-22</sup>.

TLC thus proved to be an effective preliminary tool to visualize the complex chemical composition of the extract. The potential antioxidant activity of *P. avium* bark extract

was evaluated by using two standards in vitro assays: DPPH free radical scavenging and ABTS radical cation decolorization assays. The DPPH assay of the extract showed an  $IC_{50}$  value of  $423.1 \pm 0.041 \mu\text{g/ml}$ , and the ABTS assay exhibited an  $IC_{50}$  value of  $148.5 \pm 0.92 \mu\text{g/ml}$ . The lower  $IC_{50}$  value in ABTS compared to DPPH for *P. avium* shows that the extract may be more effective against the ABTS radical cation as compared to DPPH radicals. Flavonoids and phenolics serve as hydrogen or electron donors, neutralizing reactive oxygen species (ROS) and thus mitigating oxidative stress<sup>23,24</sup>. Nevertheless, the crude extract's  $IC_{50}$  values indicate that it contains a diverse array of compounds with variable antioxidant potential.

## 5. CONCLUSION

Qualitative screening alongside TLC analysis confirmed the presence of a wide spectrum of bioactive phytochemicals in the hydroalcoholic extract of *Prunus avium* leaves. The presence of several secondary metabolites, including flavonoids, steroids, and saponins with established pharmacological actions, demonstrates this medicinal plant's therapeutic potential. Effective TLC analysis using varied solvent systems produces a comprehensive profile of the phytoconstituents, indicating substantial chemical diversity and variations in compound mobility influenced by solvent polarity. Antioxidant activity results substantiate the observations of *P. avium*'s traditional medicinal usage and warrant further exploration as a source of natural chemicals for the development of anticancer medicines.

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**Consent for publication:** The publication of the material in print, online or other media formats as determined by the publisher.

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## REFERENCES

1. Agrawal S, Kumar A, Singh AK, Singh H, Thareja S, Kumar P. A comprehensive review on pharmacognosy, phytochemistry and pharmacological activities of eight potent *Prunus* species of Southeast Asia. *J Tradit Chin Med.* 2024;44(3):620-628. <https://doi.org/10.19852/j.cnki.jtcm.2024.03.002>
2. Bento C, Goncalves AC, Silva B, Silva LR. Peach (*Prunus persica*): Phytochemicals and health benefits. *Food Rev Int.* 2022;38(10):1703-1734. <https://doi.org/10.1080/87559129.2020.1837861>
3. Chezanoglu E, Mourtzinos I, Goula AM. Sweet cherry and its by-products as sources of valuable phenolic compounds. *Trends Food Sci Technol.* 2024;145:104367. <https://doi.org/10.1016/j.tifs.2024.104367>
4. Goncalves AC, Bento C, Silva B, Simoes M, Silva LR. Nutrients, bioactive compounds and bioactivity: The health benefits of sweet cherries (*Prunus avium* L.). *Curr Nutr Food Sci.* 2019;15(3):208-227. <https://doi.org/10.2174/1573401313666170925154707>
5. Jesus F, Goncalves AC, Alves G, Silva LR. Health benefits of *Prunus avium* plant parts: An unexplored source rich in phenolic compounds. *Food Rev Int.* 2022;38(2):118-146. <https://doi.org/10.1080/87559129.2020.1854781>
6. Kashyap P, Riari CS, Jindal N. Polyphenol bio-accessibility and antioxidant activity of in vitro digested ultrasound-assisted Meghalayan cherry (*Prunus nepalensis*) pomace extract. *Biomass Convers Biorefinery.* 2023;13:14071-14085. <https://doi.org/10.1007/s13399-021-02150-0>
7. Nunes AR, Goncalves AC, Falcão A, Alves G, Silva LR. *Prunus avium* L. (sweet cherry) by-products: A source of phenolic compounds with antioxidant and anti-hyperglycemic properties-A review. *Appl Sci.* 2021;11(17):8516. <https://doi.org/10.3390/app11188516>
8. Kumar A, Nirmal P, Kumar M, Jose A, Tomer V, Oz E, Proestos C, Zeng M, Elobaid T, Sneha K, Faith Oz. Major phytochemicals: Recent advances in health benefits and extraction methods. *Molecules.* 2023;28(3):887. <https://doi.org/10.3390/molecules28020887> PMid:36677944 PMCid:PMC9862941
9. Amir M, Zafar A, Ahmad R, Ahmad W, Sarafraz M, Khalid M, Ghoneim MM, Alshehri S, Wahab S, Ahmad S. Quality control standardization, contaminant detection and in vitro antioxidant activity of *Prunus domestica* Linn. fruit. *Plants.* 2022;11(5):706. <https://doi.org/10.3390/plants11050706> PMid:35270176 PMCid:PMC8912893
10. Móricz ÁM, Ott PG. Separation and detection of apricot leaf triterpenes by high-performance thin-layer chromatography combined with direct bioautography and mass spectrometry. *J Chromatogr A.* 2022;1675:463167. <https://doi.org/10.1016/j.chroma.2022.463167> PMid:35635866
11. Ockun MA, Gercek YC, Demirsoy H, Demirsoy L, Macit I, Oz GC. Comparative evaluation of phenolic profile and antioxidant activity of new sweet cherry (*Prunus avium* L.) genotypes in Turkey. *Phytochem Anal.* 2022;33(4):564-576. <https://doi.org/10.1002/pca.3110> PMid:35122339
12. Zarzycki PK, Ślączka MM, Zarzycka MB, Włodarczyk E, Baran MJ. Application of micro-thin-layer chromatography as a simple fractionation tool for fast screening of raw extracts derived from complex biological, pharmaceutical and environmental samples. *Anal Chim Acta.* 2011;688(2):168-174. <https://doi.org/10.1016/j.aca.2011.01.001> PMid:21334482
13. Lapa-Guimarães J, Pickova J. New solvent systems for thin-layer chromatographic determination of nine biogenic amines in fish and squid. *J Chromatogr A.* 2004;1045(1-2):223-232. <https://doi.org/10.1016/j.chroma.2004.06.014> PMid:15378899
14. Mahmoud SA, Abbas AEF. Greenness, whiteness, and blueness assessment with spider chart solvents evaluation of HPTLC-densitometric method for quantifying a triple combination anti-*Helicobacter pylori* therapy. *Sustain Chem Pharm.* 2024;37:101412. <https://doi.org/10.1016/j.scp.2023.101412>
15. Nenadis N, Tsimidou MZ. DPPH (2,2-di(4-tert-octylphenyl)-1-picrylhydrazyl) radical scavenging mixed-mode colorimetric assay(s). *Meas Antioxid Act Capacit Recent Trends Appl.* 2018;141-164. <https://doi.org/10.1002/9781119135388.ch8>
16. Sanna D, Delogu G, Mulas M, Schirra M, Fadda A. Determination of free radical scavenging activity of plant extracts through DPPH assay: An EPR and UV-Vis study. *Food Anal Methods.* 2012;5(4):759-766. <https://doi.org/10.1007/s12161-011-9306-1>
17. Zheng L, Zhao M, Xiao C, Zhao Q, Su G. Practical problems when using ABTS assay to assess the radical-scavenging activity of peptides: Importance of controlling reaction pH and time. *Food Chem.* 2016;192:288-294. <https://doi.org/10.1016/j.foodchem.2015.07.015> PMid:26304349
18. Gulcin İ, Alwasel SH. DPPH radical scavenging assay. *Processes.* 2023;11(8):2248. <https://doi.org/10.3390/pr11082248>
19. Saeed RA, Khan MI, Butt MS, Faisal MN. Phytochemical screening of *Prunus avium* for its antioxidative and anti-mutagenic potential against DMBA-induced hepatocarcinogenesis. *Front Nutr.*

2023;10:1132356. <https://doi.org/10.3389/fnut.2023.1132356>  
PMid:37266135 PMCid:PMC10231329

20. Usenik V, Fajt N, Mikulic-Petkovsek M, Slatnar A, Stampar F, Veberic R. Sweet cherry pomological and biochemical characteristics influenced by rootstock. *J Agric Food Chem*. 2010;58(8):4928-4933. <https://doi.org/10.1021/jf903755b> PMid:20337477

21. Pietta PG (2000). Flavonoids as antioxidants. *Journal of Natural Products*, 63(7), 1035-1042. <https://doi.org/10.1021/np9904509>  
PMid:10924197

22. Rizvi S, Raza ST, & Ahmed F (2018). The role of plant-derived sterols and triterpenes in oxidative stress and inflammation: A review. *Free Radical Research*, 52(12), 1267-1281.

23. Saleh HA, Yousef MH, Abdelnaser A. The anti-inflammatory properties of phytochemicals and their effects on epigenetic mechanisms involved in TLR4/NF-κB-mediated inflammation. *Front Immunol*. 2021;12:606069. <https://doi.org/10.3389/fimmu.2021.606069> PMid:33868227  
PMcid:PMC8044831

24. Theansungnoen T, Nitthikan N, Wilai M, Chaiwut P, Kiattisin K, Intharuksa A. Phytochemical analysis and antioxidant, antimicrobial, and antiaging activities of ethanolic seed extracts of four *Mucuna* species. *Cosmetics*. 2022;9(1):14. <https://doi.org/10.3390/cosmetics9010014>