

## Exploring the Gut Microbiome's Influence on Peptic Ulcer Disease: Mechanistic Insights, Pharmacological Implications, and Emerging Therapeutic Strategies

<sup>1</sup> Deepannita Roy Mukherjee, <sup>2</sup> Sayak Tanbir, <sup>3</sup> Sohel Mondal, <sup>4</sup> Soumik Tarafder, <sup>5</sup> Dona Biswas, <sup>6</sup> Soumyadeb Dutta, <sup>7</sup> Saikat Santra, \* <sup>8</sup> Pinki Biswas

<sup>1</sup> Department of Pharmacology, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN - 741222(WB)

<sup>2</sup> Department of Pharmacology, Maulana Abul Kalam Azad University of technology (WB) Haringhata Farm, West Bengal 741249

<sup>3</sup> Department of Pharmacology, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN-741222 (WB)

<sup>4</sup> Department of Pharmaceutical Chemistry, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN-741222 (WB)

<sup>5</sup> Department of Pharmaceutics, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN-741222(WB)

<sup>6</sup> Department of Pharmacology, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN - 741222

<sup>7</sup> Department of Pharmaceutics, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN-741222(WB)

<sup>8</sup> Department of Pharmaceutics, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN - 741222(WB)

### Article Info:



#### Article History:

Received 23 Jan 2025

Reviewed 09 March 2025

Accepted 27 March 2025

Published 15 April 2025

### Cite this article as:

Mukherjee DR, Tanbir S, Mondal S, Tarafder S, Biswas D, Dutta S, Santra S, Biswas P, Exploring the Gut Microbiome's Influence on Peptic Ulcer Disease: Mechanistic Insights, Pharmacological Implications, and Emerging Therapeutic Strategies, Journal of Drug Delivery and Therapeutics. 2025; 15(4):209-218 DOI: <http://dx.doi.org/10.22270/jddt.v15i4.7088>

### \*Address for Correspondence:

Pinki Biswas, Department of Pharmaceutics, JRSET College of Pharmacy, Panchpota, Chakdaha, Nadia, PIN - 741222(WB)

### Abstract

**Background:** The gastrointestinal disorder Peptic Ulcer Disease (PUD) leads to mucosal damage in either stomach or duodenal tissue because of acid-peptic injury. The available evidence demonstrates that *Helicobacter pylori* (*H. pylori*) infection and nonsteroidal anti-inflammatory drug (NSAID) usage serve as proven ulcer causes but new research shows the gut microbiome as contributing to ulcer development and therapeutic approaches.

**Objective:** The purpose of this section is to examine the standard *H. pylori* ulcer development process while examining host immune responses through gut microbial alterations and their effect on PUD disease progression.

**Methods:** This study reviewed multiple research articles to examine how bacteria affect epithelial cells while studying inflammatory pathways as well as microbial metabolites particularly short-chain fatty acids (SCFAs). The review examined different pharmacy-based and natural therapies from the perspective of their ability to modulate the microbiome.

**Results:** The research shows Non-*H. pylori* bacteria damage gastric mucosal tissue by activating pro-inflammatory cytokines which leads to gastric homeostasis disturbances because of SCFA production. The ulcer formation gets worse because dysbiosis makes the intestines more sensitive to oxidative stress while simultaneously making the protective mucosal layer weaker. Antibiotics together with proton pump inhibitors affect gut microbial composition but natural treatments including curcumin combined with ginger and probiotics both contribute to microbial recovery and healing of ulcers.

**Conclusion:** Previous research about the gut microbiome's role in PUD pathogenesis forms a base for future development of tailored therapeutic strategies. Combining microbiome-based therapeutic methods with traditional medical protocols produces better management strategies for ulcers with improved healthcare results for patients.

**Keywords:** Peptic Ulcer Disease, Gut Microbiota, Microbial Dysbiosis, Immunoglobulin A (IgA), Gastrointestinal Inflammation, Mucosal Homeostasis

### 1. Introduction

Peptic ulcer disease (PUD) is mucosal break (ulcer) of the inner lining of the gastrointestinal tract caused by acid peptic digestion found when the ulcer extends beyond the muscularis mucosae to the submucosa. It occurs most commonly in the stomach and first part of the duodenum, but can be found distally in the esophagus, distally in the duodenum, and in the Meckel's diverticulum with heterotopic gastric mucosa<sup>1</sup>. Abdominal pain is

common; it is described as a burning or gnawing sensation, not usually severe, and is associated with bloating, nausea, and changes in appetite. Infection with *Helicobacter pylori* and the use of nonsteroidal anti-inflammatory drugs (NSAIDs), particularly on a long term, are the primary causes of PUD<sup>2</sup>. *H. pylori* is diagnosed with the help of a combination of medical history, endoscopy, and imaging and laboratory tests. Treatment strategies include medication – proton pump inhibitors (PPIs) to

reduce stomach acid, antibiotics to clear out the *H. pylori*, and lifestyle changes to reduce the risk of ulcers. Untreated, PUD can be complicated by severe complications including gastrointestinal bleeding, perforation of stomach or intestinal walls, and gastric outlet obstruction<sup>3</sup>. Therefore, PUD management is crucial for enhancing the quality of life and preventing complications but only effective. Early diagnosis and treatment can result in recovery and reduce a psychiatric disorder's recurrence<sup>4</sup>.

## 2. Traditional role of *Helicobacter pylori* in PUD

*Helicobacter pylori* (*H. pylori*) play a traditional and significant role in the development of peptic ulcer disease (PUD). This gram-negative bacterium colonizes the gastric mucosa and is considered one of the primary etiological agents of both gastric and duodenal ulcers<sup>5</sup>. Here are the key aspects of its role in PUD:

**2.1 Colonization:** *H. pylori* can survive in the acidic environment of the stomach by producing urease, an enzyme that converts urea into ammonia, neutralizing stomach acid and allowing the bacteria to thrive in the gastric epithelium<sup>6</sup>.

**2.2 Induction of Inflammation:** The presence of *H. pylori* induces chronic inflammation (chronic gastritis) in the stomach lining. The bacterium elicits an immune response that leads to the release of pro-inflammatory cytokines and immune cells, causing damage to the gastric mucosa<sup>7</sup>.

**2.3 Mucosal Damage:** *H. pylori* produce several virulence factors, including cytotoxins, which can directly injure the epithelial cells of the stomach. This damage disrupts the protective mucosal, chronically inflamed mucosa more susceptible to acid-peptic injury and prone to peptic ulceration<sup>8</sup>.

**2.4 Ulcer Formation:** The inflammatory response and mucosal damage caused by *H. pylori* contribute to the development of ulcers. Gastric and duodenal ulcers occur when the balance between protective factors (like mucus and bicarbonate) and aggressive factors (such as acid and pepsin) is disrupted<sup>1,9</sup>.

## 3. Expanding understanding of the gut microbiome in gastrointestinal health

In the gastrointestinal tract exists a complex microbiological system which functions fundamentally for human health, scientific investigations of the microbiome during recent times have successfully revealed new information about how this ecosystem operates in health systems and disease contexts including PUD and various gastrointestinal diseases<sup>10</sup>.

New discoveries about gut microbiome produce essential findings about its operation and structure together with its effects on health and digestive conditions<sup>11</sup>.

**3.1 Diversity and Composition:** The gut microbiome maintains multiple bacteria alongside archaea and viruses along with fungi which compose its structural variety. Multiple health functions require an equilibrium within the microbiome due to specific functions provided by different microorganisms which include nutrient

breakdown together with immune response management<sup>12</sup>.

**3.2 Role in Digestion:** During digestion, gut microorganisms break down complex fibers and carbohydrates because human enzymes lack this ability. The substrates undergo fermentation by these microorganisms to create short-chain fatty acids (SCFAs) that both generate energy for intestinal cells while simultaneously displaying anti-inflammatory properties. Microbes produce all the enzymes required for synthesizing vitamins such as B1, B9, B12 and K<sup>13</sup>.

**3.3 Immune Function:** Through mechanisms the gut microbiome establishes important regulation of the immune system capabilities. This process enables the correct identification of dangerous pathogens apart from beneficial substances to stop misdirected immune responses which result in inflammatory bowel disease and food allergies. The absence of a healthy gut microbiome makes individuals prone to develop two specific bacterial infections that affect GI tract: *C. difficile* and *H. pylori*<sup>13</sup>.

**3.4 Interaction with Pathogens:** A healthy gut microbiome protects individuals from pathogenic bacteria through its interactive mechanism including *H. pylori*. Healthy bacteria defend against pathogenic microbes through resource acquisition and substance production to decrease disease development including PUD and infections<sup>14</sup>.

**3.6 Influence on Metabolism:** The gut microbiome is involved in metabolic processes, including the synthesis of vitamins and the regulation of lipid and glucose metabolism. Dysbiosis (imbalance in the microbiome) has been linked to obesity, diabetes, and metabolic syndrome<sup>15</sup>.

**3.7 Gut-Brain Axis:** The microbiome communicates with the central nervous system through the gut-brain axis, influencing mood, stress responses, and behaviour. This connection highlights the microbiome's potential role in mental health and neurological conditions<sup>16</sup>.

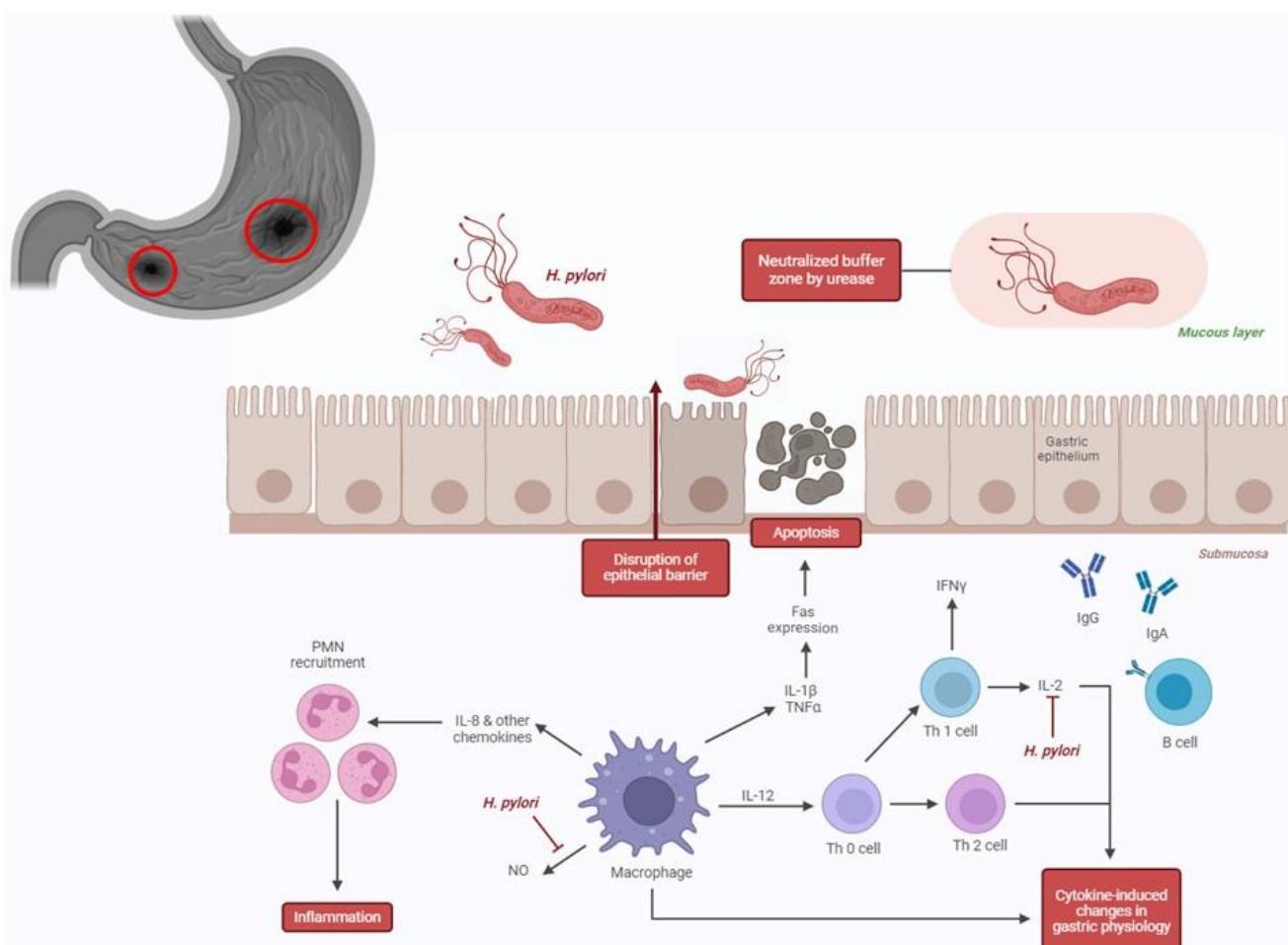
**3.8 Therapeutic Potential:** Understanding the gut microbiome opens new avenues for therapeutic interventions. Probiotics, prebiotics, and dietary modifications can help restore microbiome balance, promoting gastrointestinal health and potentially alleviating conditions like PUD and IBD<sup>17</sup>.

## 4. Mechanism of Gut Bacteria in Peptic Ulcer Pathogenesis

Tissue underlying peptic ulcers becomes exposed after the breakdown of a protective mucosal layer enables gastric acid to damage the vulnerable area. *H. pylori* serves as the most recognized bacterial factor in creating ulcers yet gut bacteria without *H. pylori* also significantly contributes to ulcer development<sup>18</sup>. The bacteria damage gastric mucosa through changes to gut microbiota ecology and mucin breakdown leading to breakdown of tight junction proteins. Bacteria set off inflammation upon activation of Toll-like receptors (TLRs) on gastric epithelial cells which result in the release of pro-inflammatory cytokines such as IL-1 $\beta$ , IL-6 and TNF- $\alpha$ <sup>19</sup>. The ulcer-forming process becomes accelerated because bacterial

induced production of reactive oxygen species (ROS) and oxidative stress combined with inflammation damages gastric cells. The gastric pH and mucosal permeability are altered when bacterial metabolites, particularly short-chain fatty acids (SCFAs) such as acetate and propionate butyrate trigger risk increased susceptibility to ulcers. The bacterial imbalance known as dysbiosis increases both stomach acid levels and

degrades protective defense systems while permitting excessive bacterial growth<sup>20</sup>. The identification of novel bacterial interactions with *H. pylori* provides important insights into peptic ulcer development. However, *H. pylori* continue to be the primary pathogenic factor which suggests new therapeutic targets based on gut microbiota management<sup>21,22</sup>.



**Figure 1: The diagram of *H. pylori* pathogenesis** where urease neutralizes acid, disrupting the epithelial barrier and triggering apoptosis. Macrophages activate Th1 and Th2 cells, leading to cytokine release, inflammation, and immune evasion. Persistent immune responses weaken the mucosal barrier, promoting ulcer formation<sup>23,24</sup>.

## 5. Gut Microbial Dysbiosis and Its Role in Peptic Ulcer Disease (PUD)

Peptic ulcer disease (PUD) is a disease of pathogenesis which is related with gut microbial dysbiosis, abnormality of the microbial community. However, even conditions of gut microbiota disruption may worsen ulcer formation, impede healing and promote treatment resistance in the presence of *Helicobacter pylori* (*H. pylori*) as the key bacterial factor<sup>25</sup>. A healthy gut microbiome maintains mucosal integrity, modulates immune responses, and affects the regulation of gastric acid secretion. But when dysbiosis means problems with these protective mechanisms, the possibility of injuring the gastric mucous is enhanced<sup>26</sup>. This weakness of mucosal barrier is a consequence of loss of beneficial microbes like *Lactobacillus* and *Bifidobacterium*, which produce short chain fatty acids (SCFAs) required to

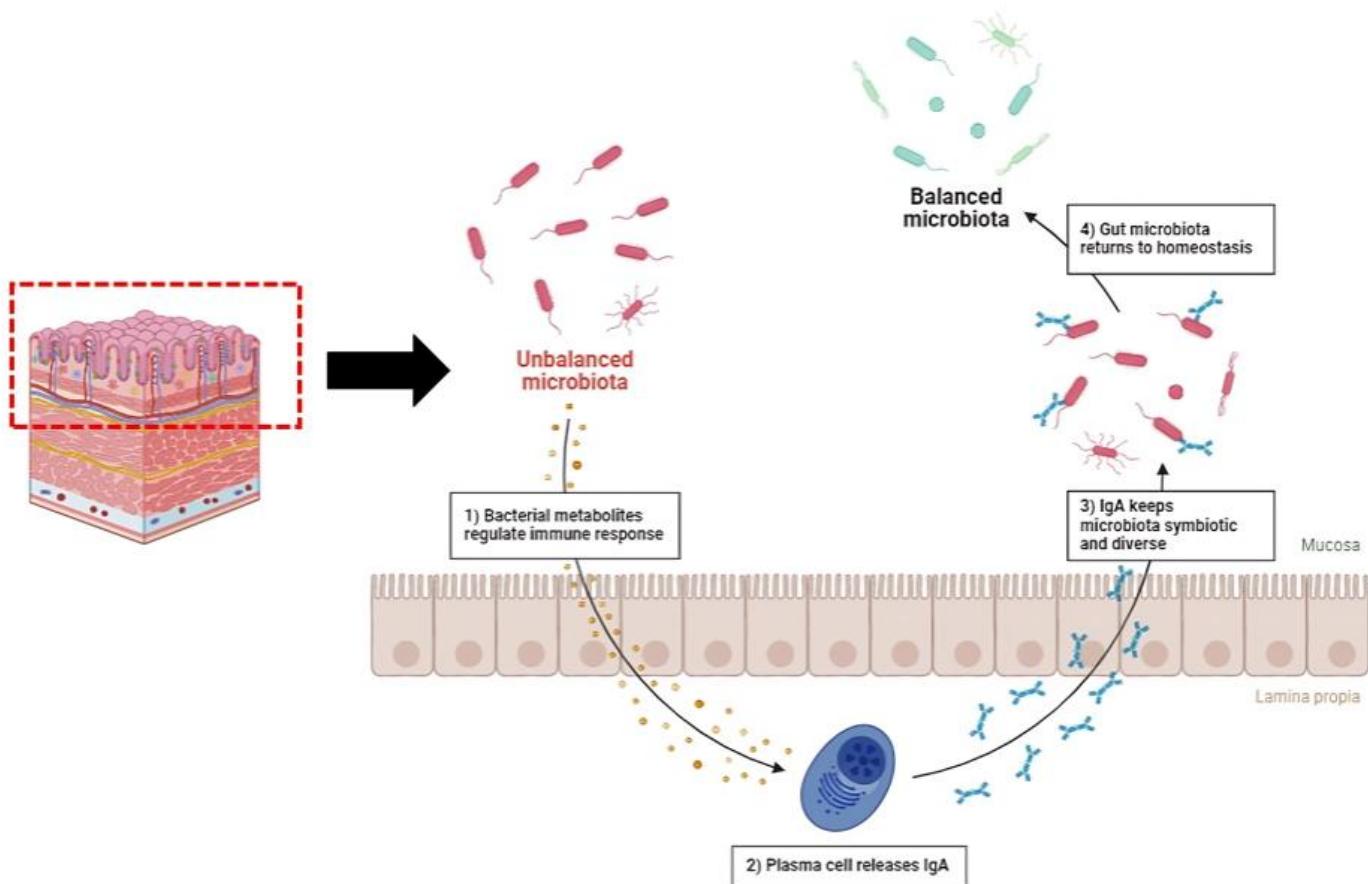
maintain gastric homeostasis. Increased epithelial permeability as well as makes the stomach lining more susceptible to acid related injury<sup>27</sup>.

Also, holistic dysbiosis promotes the overgrowth of opportunistic pathogens, such as *Escherichia coli*, *Klebsiella*, *Enterococcus* and *Clostridium*<sup>28</sup>. They release virulence factors, trigger mucin loss, and produce proinflammatory cytokines, including IL-1 $\beta$ , IL-6 and TNF  $\alpha$ , which result in chronic inflammatory and oxidative stress<sup>28</sup>. Moreover, microbial imbalance changes gastric signalling patterns of acid secretion to further aggravate mucosal damage. Drugs are directly influenced by dysbiosis, in terms of metabolism and efficacy. PUD treatment antibiotics in the treatment of PUD influence absorption of PPIs and antibiotics by the gut microbiota. A reduced drug effectiveness and

participation in antibiotic resistance via an imbalanced microbiome may impair *H. pylori* eradication<sup>29</sup>.

For dysbiosis in PUD management, one should address the disequilibrium by using probiotics, prebiotics, and microbiome friendly therapies. Like *Lactobacillus* and *Bifidobacterium*, probiotics help quell pathogenic bacteria, build up the mucosal barrier, and decrease the level of inflammation<sup>30</sup>. Dietary modification consisting of increase Fiber intake stimulates the growth of beneficial bacteria that also increases SCFA production.

Antimicrobial and anti-inflammatory natural compounds like curcumin, polyphenols, and flavonoids are microbiome friendly alternatives to conventional treatments<sup>31</sup>. Future therapeutic approaches may involve personalized microbiome profiling to develop targeted interventions that optimize gut health and improve PUD outcomes. By integrating microbiome-based strategies with standard medical treatments, it is possible to enhance ulcer healing, reduce recurrence, and improve overall gastrointestinal health<sup>32</sup>.



**Figure 2: Role of IgA in Regulating Gut Microbiota and Its Impact on Peptic Ulcer Disease** IgA regulates the gut microbiome, which is crucial in preventing gut microbial dysbiosis—a key factor in peptic ulcer disease (PUD). Dysbiosis occurs when harmful bacteria outnumber beneficial ones, triggering inflammation and mucosal damage. The immune system responds by releasing IgA, which helps maintain microbial balance by preventing harmful bacterial overgrowth. When IgA functions properly, gut microbiota return to homeostasis, reducing inflammation and protecting against ulcers<sup>33</sup>.

## 6. Bacterial Interactions with Gastric Epithelial Cells

Gut bacteria can interact directly with the gastric epithelial cells, disrupting the epithelial barrier, cellular damage, and ulcer formation<sup>4</sup>. While *H. pylori* is well-known for its ability to adhere to gastric epithelial cells via specific adhesins, non-*H. pylori* bacteria can also colonize the stomach and contribute to mucosal injury<sup>18</sup>.

**6.1 Adhesion and Colonization:** Certain non-*H. pylori* bacteria, such as *Streptococcus*, *Enterococcus*, and *Lactobacillus*, can colonize the gastric mucosa. These bacteria may adhere to epithelial cells through different

adhesions and surface proteins, which allow them to persist in the stomach despite the acidic environment<sup>34</sup>.

**6.2 Barrier Disruption:** The presence of gut bacteria on the epithelial surface can disrupt tight junctions between cells. This disruption increases the permeability of the gastric mucosa, allowing stomach acids and other irritants to penetrate the epithelial layer, causing inflammation and ulceration<sup>35</sup>.

**6.3 Cytotoxic Effects:** Some bacterial strains, such as cytotoxins or proteases, produce toxins or enzymes that damage epithelial cells. This cytotoxic effect can lead to

cell death, triggering tissue damage, inflammation, and eventually ulcer formation<sup>36</sup>.

## 7. Inflammatory Responses Triggered by Non-*H. pylori* Bacteria

The interaction between gut bacteria and the host's immune system is critical in peptic ulcer pathogenesis. Even without *H. pylori* and other bacterial species can induce an inflammatory response contributing to ulcer formation<sup>37</sup>.

**7.1 Pattern Recognition Receptors (PRRs):** Non-*H. pylori* bacteria can activate pattern recognition receptors like Toll-like receptors (TLRs) on gastric epithelial cells and immune cells. The activation of TLRs leads to the production of pro-inflammatory cytokines, such as interleukin-1 (IL-1), interleukin-6 (IL-6), and tumour necrosis factor-alpha (TNF- $\alpha$ ). These cytokines can promote inflammation, leading to tissue damage and ulcer development<sup>38</sup>.

**7.2 Neutrophil and Macrophage Activation:** The inflammatory response recruits immune cells such as neutrophils and macrophages to the site of infection. The activated immune cells release reactive oxygen species (ROS) and proteolytic enzymes, which can damage the gastric mucosa and contribute to ulcer formation<sup>39</sup>.

**7.3 Microbial Dysbiosis:** Imbalances in gut microbiota composition, also known as dysbiosis, can exacerbate inflammation. For example, increasing pathogenic bacteria or decreasing protective commensal bacteria may enhance the inflammatory response, leading to chronic gastritis and peptic ulcer development<sup>40</sup>.

## 8. Role of Metabolites (e.g., Short-Chain Fatty Acids) in Ulcer Formation

Metabolites produced by gut bacteria, such as short-chain fatty acids (SCFAs), can also influence the pathogenesis of peptic ulcers. SCFAs are primarily produced through fermentation of dietary fibers by gut microbiota, including acetate, propionate, and butyrate<sup>41</sup>.

**8.1 Protective Effects of SCFAs:** SCFAs, especially butyrate, have anti-inflammatory and protective effects on the gastric mucosa. They can help maintain the integrity of the epithelial barrier by enhancing mucus production and stimulating the proliferation of epithelial cells. However, when dysbiosis occurs, the beneficial effects of SCFAs may be reduced, leading to increased susceptibility to ulceration<sup>17,42</sup>.

**8.2 Butyrate and Mucosal Repair:** Butyrate can promote mucosal healing by inducing the expression of genes associated with cell growth and repair. It also has the potential to inhibit the expression of pro-inflammatory cytokines, thus reducing inflammation and aiding in the resolution of peptic ulcers<sup>43</sup>.

**8.3 Dysbiosis and Altered SCFA Production:** Changes in gut microbiota composition may alter the production of SCFAs. For instance, an overgrowth of certain bacteria may increase the production of toxic metabolites, such as ammonia or secondary bile acids, which can disrupt the gastric barrier and lead to ulcer formation. Reducing butyrate-producing bacteria can also impair mucosal healing and increase the risk of chronic ulcers<sup>44</sup>.

**Table: 1** Drug Use and Its Impact on Gut Microbiome in Peptic Ulcer Disease

Drug Category	Examples	Purpose in PUD	Impact on Gut Microbiome	Mitigation Strategies	Ref
<b>Proton Pump Inhibitors (PPIs)</b>	Omeprazole, Pantoprazole, Esomeprazole	Reduce gastric acid secretion to promote ulcer healing	Increases gastric pH, leading to overgrowth of bacteria like <i>Enterococcus</i> and <i>Streptococcus</i>	Use only when necessary; monitor gut microbiome health	45
<b>H2 Receptor Antagonists (H2RAs)</b>	Ranitidine (withdrawn), Famotidine	Reduce acid production as an alternative to PPIs	Less impact on gut microbiota than PPIs but may still alter microbial composition	Prefer H2RAs over PPIs for long-term use when possible	46
<b>Antibiotics (for <i>H. pylori</i> eradication)</b>	Amoxicillin, Clarithromycin, Metronidazole, Levofloxacin	Kill <i>H. pylori</i> to prevent ulcer recurrence	Reduces microbial diversity, disrupts gut flora, and increases risk of antibiotic resistance	Combine with probiotics to restore beneficial bacteria	47
<b>Bismuth-containing compounds</b>	Bismuth subsalicylate, Bismuth subcitrate	Forms protective coating, has antimicrobial effects against <i>H. pylori</i>	Mildly alters microbiota but may help maintain microbial balance	Used in <i>H. pylori</i> eradication therapy	48
<b>Nonsteroidal Anti-inflammatory Drugs (NSAIDs)</b>	Ibuprofen, Aspirin, Naproxen	Pain relief and anti-inflammatory	Damages gut lining, alters microbiome	Use gastroprotective agents (PPIs, misoprostol); prefer	49

(risk factors for PUD)		effects but can cause ulcers	composition, increases gut permeability	COX-2 inhibitors if necessary	
<b>Cytoprotective Agents</b>	Sucralfate, Misoprostol	Sucralfate coats ulcers: Misoprostol stimulates mucus and bicarbonate secretion	Minimal effect on gut microbiota	Can be used to prevent NSAID-induced	50

## 9. Natural Drugs and Phytochemicals in Modulating Gut Microbiome for Peptic Ulcer Treatment

Peptic ulcers are primarily caused by the bacterium *Helicobacter pylori* (*H. pylori*) and the prolonged use of nonsteroidal anti-inflammatory drugs (NSAIDs). A key factor in both their development and treatment is the role of the gut microbiome<sup>51</sup>. Natural drugs and phytochemicals are increasingly studied for their potential to modulate gut health and combat infections like *H. pylori*. Here's how some prominent natural compounds play a role:

### 9.1 Role of Natural Compounds in Gut Health

**9.1.1 Curcumin (from turmeric):** Curcumin is a potent anti-inflammatory and antioxidant compound. It has been shown to enhance gut health by modulating the gut microbiome, promoting the growth of beneficial bacteria, and reducing gut inflammation. Curcumin's role in inhibiting the adhesion of *H. pylori* to the gastric mucosa and reducing inflammatory responses makes it useful in preventing or treating ulcers. It also helps to reduce the oxidative stress caused by *H. pylori*, further supporting gut integrity<sup>52</sup>.

**9.1.2 Ginger:** Ginger contains bioactive compounds like gingerols and shogaols that have anti-inflammatory, antioxidant, and antimicrobial effects. Ginger helps maintain gut health by reducing inflammation and modulating gut motility. Studies have shown that ginger extracts can inhibit the growth of *H. pylori* and improve gastric mucosal health by increasing mucus secretion, which acts as a protective layer against acid and bacterial invasion<sup>53</sup>.

**9.1.3 Garlic:** Garlic contains allicin, a compound with potent antimicrobial properties. Garlic has shown significant activity against *H. pylori* and other gut pathogens that contribute to ulcer formation. It also helps balance the gut microbiome by promoting beneficial bacteria while suppressing harmful ones. The antibacterial, antifungal, and immune-boosting properties of garlic make it effective in preventing bacterial overgrowth and enhancing gut health<sup>53</sup>.

### 9.2 Phytochemicals and Their Antibacterial Activity against *H. pylori* and Other Ulcer-related Bacteria

**9.2.1 Polyphenols:** These are a class of phytochemicals found in fruits, vegetables, and herbs like green tea, berries, and olive oil. Polyphenols have demonstrated antibacterial activity against *H. pylori* by disrupting bacterial membranes, inhibiting its growth, and reducing its ability to colonize the stomach lining. Epigallocatechin gallate (EGCG), a polyphenol in green tea, has been

shown to suppress *H. pylori* growth and reduce gastric inflammation<sup>54</sup>.

**9.2.2 Flavonoids:** Flavonoids are potent antioxidants found in plants like chamomile, liquorice, and apples. Their antibacterial activity against *H. pylori* includes inhibiting bacterial enzymes and suppressing its urease activity, which is essential for *H. pylori* survival in the acidic stomach environment. Flavonoids like quercetin and rutin are effective in reducing gastric acid secretion, promoting mucus production, and enhancing gastric mucosal defence<sup>55</sup>.

**9.2.3 Tannins:** Found in foods like tea, coffee, and berries, tannins possess antibacterial properties that have been found effective against *H. pylori*. Tannins inhibit bacterial adhesion to the stomach lining and reduce inflammation associated with ulcers<sup>56</sup>.

## 10. Plant-based Therapies to Restore Microbiome Balance and Promote Ulcer Healing

**10.1 Probiotics and Prebiotics:** Probiotic supplements containing beneficial bacteria (e.g., *Lactobacillus* and *Bifidobacterium* species) help restore microbiome balance disrupted by *H. pylori* infections or the use of antibiotics in ulcer treatment. Probiotics have shown the ability to suppress *H. pylori* growth, modulate immune responses, and enhance the healing of gastric mucosa. Prebiotics, which are fibers that feed beneficial bacteria, can help sustain a healthy gut environment<sup>57</sup>.

**10.2 Liquorice (*Glycyrrhiza glabra*):** Deglycyrrhizinated licorice (DGL) is often used as a natural remedy for ulcers. It helps promote the secretion of protective mucus in the stomach and acts as an anti-inflammatory agent. Licorice can also suppress *H. pylori* growth and enhance the healing of ulcers.

**10.3 Aloe Vera:** Aloe vera has soothing and healing properties that can support the healing of the gastric lining. It contains bioactive compounds with anti-inflammatory and antimicrobial effects. Aloe vera has been shown to promote mucus production, which protects the stomach lining from the harmful effects of gastric acid and bacteria<sup>58</sup>.

**10.4 Slippery Elm (*Ulmus rubra*):** Slippery elm contains mucilage, a gel-like substance that coats and soothes the stomach lining. It has been used in herbal medicine to treat ulcers by protecting the stomach from acid and aiding in the healing of inflamed tissues<sup>59</sup>.

## 10. Clinical Applications of Various Probiotics, Prebiotics and Symbiotic

Probiotics, prebiotics, and symbiotics play a vital role in managing peptic ulcer disease (PUD) by restoring gut microbiota balance, reducing inflammation, and promoting mucosal healing.

**10.1 Probiotics** such as *Lactobacillus acidophilus*, *Bifidobacterium bifidum*, and *Saccharomyces boulardii* help suppress *H. pylori*, enhance immune responses, and reduce antibiotic side effects. They also improve gut barrier integrity and reduce inflammation<sup>57</sup>.

**10.2 Prebiotics** like inulin, fructose oligosaccharides (FOS), and resistant starch stimulate beneficial bacteria, enhance short-chain fatty acid (SCFA) production, and support epithelial repair<sup>60</sup>.

**10.3 Symbiotic**, a combination of probiotics and prebiotics, further enhance gut health by improving microbial colonization, boosting immune function, and accelerating ulcer healing. These therapies serve as effective adjuncts to conventional PUD treatment, improving eradication success rates while minimizing gastrointestinal side effects<sup>61</sup>.

## 11. Gut Microbiota and Obesity

The gut microbes maintain the metabolic equilibrium of the host. Another study, for example, found that the gut microbiota of adult population with type 2 diabetes differs from that of non-diabetic adults, and that health of the subjects may improve by modulating their gut microflora by probiotics and prebiotics administration. Although these findings and the linkage between obesity and diabetes and abdominal fat presence, few studies have been directed at identifying putative correlation between composition of the microbiota and occurrence of inflammation and metabolic changes in humans with obesity<sup>62</sup>. Patients with diabetes mellitus were studied in which those individuals had a lower number of *Faecalibacterium prausnitzii* and higher levels of inflammatory markers. Large changes in the abundances of various bacteria from different taxa, however, were found to be associated with obesity<sup>63</sup>.

Individuals with obesity have lower Bifidobacteria population (and most other organisms in the group of Firmicutes) as compared to lean individuals. As we also found in patients with type 2 diabetes mellitus to nondiabetic patients. These results indicate that Bifidobacteria might also have an involvement in the genesis of obesity and associated comorbidities. Bacteria used prebiotics such as inulin-type fructus as energy substrates when the mice were fed prebiotics such as inulin-type fructus. Levels of lipopolysaccharide, glucose tolerance, and fat mass developed were inversely correlated, and Bifidobacteria number increased much. Additionally, the prebiotic approach inhibited overexpression of many proadiposity and proinflammatory host genes<sup>64</sup>.

## 11. Faecal Microbiota Transplantation

Studies show that IBD causes changes to gut microbiota diversity and includes increased pathogen levels alongside decreased commensal bacteria populations. For this reason health experts view FMT as an emerging IBD therapeutic option. FMT employs donor fecal material as a treatment method to return intestinal

microbiota back to its normal state after pathology-induced changes in the recipient patient. FMT exists in two different delivery forms: the lower gastrointestinal tract and the upper gastrointestinal tract. Enema and colonoscopy and sigmoidoscopy belong to the former category while endoscopy and nasoduodenal tube and capsule ingestion constitute the latter. Research continues to evaluate the effectiveness of FMT therapy despite its proven ability in *Clostridium difficile* infection recurrence<sup>65</sup>.

## 12. Future Perspectives

In the areas of peptic ulcer disease (PUD), future research seeks to develop novel therapies with microbiome and personalized medicine as well as highly advanced diagnostics. Such microbiome targeted therapies include probiotics, prebiotics and symbiotic etc., which are expected to restore the obliterated microbial balance and to improve gut health<sup>66</sup>. Moreover, there are also safer and more effective treatment options, including microbiome friendly antibiotics and phytochemicals in novel drug formulations. Genomic and Microbiome profiling will apply to personalize medicine with personalized therapies based on individual susceptibility and microbial composition<sup>67</sup>. Early detection and monitoring on microbial imbalances will be facilitated by advanced diagnostic tools such as artificial intelligence (AI) and non-invasive biomarkers. Precise ways to get rid of harmful bacteria while preserving the healthy bacteria may be provided by CRISPR-based microbiome editing as well as by genetically engineered probiotics<sup>68</sup>. In addition, mucosal vaccines targeting *H. pylori* may provide a long-term prevention, and FMT as a therapy for patients with resistant cases. Further optimization of patient outcomes will continue to be achieved with holistic treatment models incorporating dietary interventions and gut brain axis therapies. Future work will also expand clinical trials and develop regulatory frameworks to validate proof of safety and efficacy of these microbiome approaches to prevention and management of PUD<sup>69</sup>.

## 13. Conclusion

Peptic Ulcer Disease (PUD) is a persistent condition affected by *Helicobacter pylori* (*H. pylori*), nonsteroidal anti-inflammatory drugs (NSAIDs), and gut microbial interactions. While advances in microbiome research have enhanced the understanding of PUD pathogenesis based on microbial diversity, inflammation and metabolites (short chain fatty acids), traditional treatments remain a problem. While there are antibiotics, proton pump inhibitors (PPIs) and H2 receptor antagonists for addressing the ulcers, they also affect the microbial balance of your gut causing dysbiosis and other complications.

This calls for searching for therapeutic alternatives that are microbiome friendly. Natural compounds including curcumin, polyphenols, flavonoids and probiotics modulate gut microbiota, reduce inflammation, and improve mucosal protection and are promising. In addition, such peanuts might assist in restoring microbial

homeostasis while ulcer healing while causing little side effects.

To optimize PUD treatment, future innovations in microbiome engineering and personalized medicine will be important. Interventions tailored for individual microbial compositions can be based on advanced diagnostics such as next generation sequencing and metabolomics. Restoring gut homeostasis will likely be achieved by further revolutionizing treatment from the emerging therapies like genetically engineered probiotics and fecal microbiota transplantation (FMT).

In order to bridge the conventional management of PUD and microbiome-based treatments one has to approach with a multidisciplinary research approach that integrates gastroenterology, microbiology and precision medicine. By focusing on microbiome health as much as possible, along with other treatments, healthcare providers can increase the success rates in recovery, reduce recurrence of ulcers, and minimize long term complications. A microbiome focused shift toward integrative corner of ulcer care is likely to boost efficacy and maintainability of ulcer management, and lead the way for next generation, patient centred ulcer management.

**Conflict of Interest:** The authors declare no potential conflict of interest with respect to the contents, authorship, and/or publication of this article.

**Author Contributions:** All authors have equal contribution in the preparation of manuscript and compilation.

**Source of Support:** Nil

**Funding:** The authors declared that this study has received no financial support.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** The data supporting in this paper are available in the cited references.

**Ethical approval:** This study does not involve experiments on animals or human subjects.

## References

1. Harsha C, Banik K, Bordoloi D, Kunnumakkara AB. Antiulcer properties of fruits and vegetables: A mechanism based perspective. *Food and Chemical Toxicology*, 2017; 108:104-119. <https://doi.org/10.1016/j.fct.2017.07.023>
2. Smith ME, Morton DG. The stomach. In: *The Digestive System*. 2nd ed. Elsevier; 2010:51-69. <https://doi.org/10.1016/B978-0-7020-3367-4.00004-9>
3. Wallace JL. Recent advances in gastric ulcer therapeutics. *Current Opinion in Pharmacology*, 2005; 5(6):573-577. <https://doi.org/10.1016/j.coph.2005.06.004>
4. Malik TF, Gnanapandithan K, Singh K. Peptic ulcer disease. In: *Helicobacter pylori*. Springer; 2023:635-639. <https://doi.org/10.1007/978-981-97-0013-4-54>
5. Malfertheiner P, Mégraud F, O'Morain C, et al. Current concepts in the management of Helicobacter pylori infection: The Maastricht III consensus report. *Gut*, 2007; 56(6):772-781. <https://doi.org/10.1136/gut.2006.101634>
6. Elbehiry A, Marzouk E, Aldubaib M, et al. Helicobacter pylori infection: Current status and future prospects on diagnostic, therapeutic and control challenges. *Antibiotics*, 2023; 12(2):191. <https://doi.org/10.3390/antibiotics12020191>
7. Hooi JKY, Lai WY, Ng WK, et al. Global prevalence of Helicobacter pylori infection: Systematic review and meta-analysis. *Gastroenterology*, 2017; 153(2):420-429. <https://doi.org/10.1053/j.gastro.2017.04.022>
8. Kayali S, Manfredi M, Gaiani F, et al. Helicobacter pylori, transmission routes and recurrence of infection: State of the art. *Acta Biomedica*, 2018; 89:72-76. <https://doi.org/10.23750/abm.v89i8-s.7947>
9. Parikh NS, Ahlawat R. Helicobacter pylori. *StatPearls*. Published online August 8, 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK534233/>
10. Schloss PD, Handelsman J. Status of the microbial census. *Microbiology and Molecular Biology Reviews*, 2004; 68(4):686-691. <https://doi.org/10.1128/MMBR.68.4.686-691.2004>
11. Bull MJ, Plummer NT. Part 1: The human gut microbiome in health and disease. *Integrative Medicine: A Clinician's Journal*, 2014; 13(6):17. Available from: <https://PMC.ncbi.nlm.nih.gov/articles/PMC4566439/>
12. Oami T, Chihade DB, Coopersmith CM. The microbiome and nutrition in critical illness. *Current Opinion in Critical Care*, 2019; 25(2):145-149. <https://doi.org/10.1097/MCC.0000000000000582>
13. Leonard JM, Del Toro D. Defining the microbiome components (bacteria, viruses, fungi) and microbiome geodiversity. *Surgical Infections*, 2023; 24(3):208. <https://doi.org/10.1089/sur.2023.014>
14. Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. *Cell*, 2006; 124(4):837-848. <https://doi.org/10.1016/j.cell.2006.02.017>
15. Mazmanian SK, Kasper DL. The love-hate relationship between bacterial polysaccharides and the host immune system. *Nature Reviews Immunology*, 2006; 6(11):849-858. <https://doi.org/10.1038/nri1956>
16. Dethlefsen L, McFall-Ngai M, Relman DA. An ecological and evolutionary perspective on human-microbe mutualism and disease. *Nature*, 2007; 449(7164):811-818. <https://doi.org/10.1038/nature06245>
17. Round JL, Mazmanian SK. The gut microbiome shapes intestinal immune responses during health and disease. *Nature Reviews Immunology*, 2009; 9(5):313. <https://doi.org/10.1038/nri2515>
18. Akin OY, Tsou VM, Werner AL. *Gastrospirillum hominis*-associated chronic active gastritis. *Fetal and Pediatric Pathology*, 1995; 15(3):429-435. <https://doi.org/10.3109/15513819509026978>
19. Akada JK, Shirai M, Takeuchi H, Tsuda M, Nakazawa T. Identification of the urease operon in *Helicobacter pylori* and its control by mRNA decay in response to pH. *Molecular Microbiology*, 2000; 36(5):1071-1084. <https://doi.org/10.1046/j.1365-2958.2000.01918.x>
20. Akhiani AA, Pappo J, Kabok Z, et al. Protection against *Helicobacter pylori* infection following immunization is IL-12-dependent and mediated by Th1 cells. *Journal of Immunology*, 2002; 169(12):6977-6984. <https://doi.org/10.4049/jimmunol.169.12.6977>
21. Kusters JG, Van Vliet AHM, Kuipers EJ. Pathogenesis of *Helicobacter pylori* infection. *Clinical Microbiology Reviews*, 2006; 19(3):449-490. <https://doi.org/10.1128/CMR.00054-05>
22. Achtman M, Suerbaum S. Sequence variation in *Helicobacter pylori*. *Trends in Microbiology*, 2000; 8(2):57-58. [https://doi.org/10.1016/S0966-842X\(99\)01676-5](https://doi.org/10.1016/S0966-842X(99)01676-5)
23. Ferlay J, Colombet M, Soerjomataram I, et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. *International Journal of Cancer*, 2019; 144(8):1941-1953. <https://doi.org/10.1002/ijc.31937>

24. Reyes VE. *Helicobacter pylori* and its role in gastric cancer. *Microorganisms*, 2023; 11(5):1312. <https://doi.org/10.3390/microorganisms11051312>

25. Kurilshikov A, Medina-Gomez C, Bacigalupo R, et al. Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nature Genetics*, 2021; 53(2):156-165. <https://doi.org/10.1038/s41588-020-00763-1>

26. Zhao JH, Stacey D, Eriksson N, et al. Genetics of circulating inflammatory proteins identifies drivers of immune-mediated disease risk and therapeutic targets. *Nature Immunology*, 2023; 24(9):1540-1551. <https://doi.org/10.1038/s41590-023-01588-w>

27. He Y, Koido M, Sutoh Y, et al. East Asian-specific and cross-ancestry genome-wide meta-analyses provide mechanistic insights into peptic ulcer disease. *Nature Genetics*, 2023; 55(12):2129-2138. <https://doi.org/10.1038/s41588-023-01569-7>

28. Mujadzic H, Noorani S, Riddle PJ, et al. Ulcer bleeding in the United States: Epidemiology, treatment success, and resource utilization. *Digestive Diseases and Sciences*, 2024; 69(6):1963-1971. <https://doi.org/10.1007/s10620-024-08322-y>

29. Li W, Lv BM, Quan Y, Zhu Q, Zhang HY. Associations between serum mineral nutrients, gut microbiota, and risk of neurological, psychiatric, and metabolic diseases: A comprehensive Mendelian randomization study. *Nutrients*, 2024; 16(2):244. <https://doi.org/10.3390/nu16020244>

30. Sanders ME, Merenstein DJ, Reid G, Gibson GR, Rastall RA. Probiotics and prebiotics in intestinal health and disease: From biology to the clinic. *Nature Reviews Gastroenterology & Hepatology*, 2019; 16(10):605-616. <https://doi.org/10.1038/s41575-019-0173-3>

31. Agirman G, Yu KB, Hsiao EY. Signaling inflammation across the gut-brain axis. *Science*, 2021; 374(6571):1087-1092. <https://doi.org/10.1126/science.abi6087>

32. Li J, Cai H, Zhang Y, et al. Dysbiosis of gut microbiota is associated with pathogenesis of peptic ulcer diseases through inflammatory proteins: A Mendelian randomization study. *Medicine (Baltimore)*, 2024; 103(39):e39814. <https://doi.org/10.1097/MD.00000000000039814>

33. Monteiro RC, Rafeh D, Gleeson PJ. Is there a role for gut microbiome dysbiosis in IgA nephropathy? *Microorganisms*, 2022; 10(4):683. <https://doi.org/10.3390/microorganisms10040683>

34. Backert S, Clyne M, Tegtmeyer N. Molecular mechanisms of gastric epithelial cell adhesion and injection of CagA by *Helicobacter pylori*. *Cell Communication and Signaling*, 2011; 9:28. <https://doi.org/10.1186/1478-811X-9-28>

35. Wroblewski LE, Peek RM. Targeted disruption of the epithelial-barrier by *Helicobacter pylori*. *Cell Communication and Signaling*, 2011; 9:29. <https://doi.org/10.1186/1478-811X-9-29>

36. Alzahrani S, Lina TT, Gonzalez J, Pinchuk IV, Beswick EJ, Reyes VE. Effect of *Helicobacter pylori* on gastric epithelial cells. *World Journal of Gastroenterology*, 2014; 20(36):12767. <https://doi.org/10.3748/wjg.v20.i36.12767>

37. Suzuki H, Moayyedi P. *Helicobacter pylori* infection in functional dyspepsia. *Nature Reviews Gastroenterology & Hepatology*, 2013; 10(3):168-174. <https://doi.org/10.1038/nrgastro.2013.9>

38. Warren JR, Marshall B. Unidentified curved bacilli in gastric epithelium in active chronic gastritis. *The Lancet*, 1983; 321(8336):1273-1275. [https://doi.org/10.1016/S0140-6736\(83\)92719-8](https://doi.org/10.1016/S0140-6736(83)92719-8)

39. Wroblewski LE, Peek RM, Wilson KT. *Helicobacter pylori* and gastric cancer: Factors that modulate disease risk. *Clinical Microbiology Reviews*, 2010; 23(4):713-739. <https://doi.org/10.1128/CMR.00011-10>

40. Ali A, AlHussaini KI. *Helicobacter pylori*: A contemporary perspective on pathogenesis, diagnosis and treatment strategies. *Microorganisms*, 2024; 12(1):222. <https://doi.org/10.3390/microorganisms12010222>

41. Kim MS, Kim Y, Choi H, et al. Transfer of a healthy microbiota reduces amyloid and tau pathology in an Alzheimer's disease animal model. *Gut*, 2020; 69(2):283-294. <https://doi.org/10.1136/gutjnl-2018-317431>

42. Palm NW, de Zoete MR, Flavell RA. Immune-microbiota interactions in health and disease. *Clinical Immunology*, 2015; 159(2):122-127. <https://doi.org/10.1016/j.clim.2015.05.014>

43. Shin Y, Han S, Kwon J, et al. Roles of short-chain fatty acids in inflammatory bowel disease. *Nutrients*, 2023; 15(20):4466. <https://doi.org/10.3390/nu15204466>

44. Lynch SV, Pedersen O. The human intestinal microbiome in health and disease. *The New England Journal of Medicine*, 2016; 375(24):2369-2379. <https://doi.org/10.1056/NEJMra1600266>

45. Freedberg DE, Lebwohl B, Abrams JA. The impact of proton pump inhibitors on the human gastrointestinal microbiome. *Clinics in Laboratory Medicine*, 2014; 34(4):771. <https://doi.org/10.1016/j.cll.2014.08.008>

46. Nugent CC, Falkson SR, Terrell JM. H2 blockers. *Journal of Clinical Gastroenterology*, 2024; 5:143-148. <https://doi.org/10.1097/00004836-198312001-00014>

47. Zaman T, Haq A, Ahmad R, et al. The role of probiotics in the eradication of *Helicobacter pylori* and overall impact on management of peptic ulcer: A study involving patients undergoing triple therapy in Bangladesh. *Cureus*, 2024; 16(3):e56283. <https://doi.org/10.7759/cureus.56283>

48. Alkim H, Koksal AR, Boga S, Sen I, Alkim C. Role of bismuth in the eradication of *Helicobacter pylori*. *American Journal of Therapeutics*, 2017; 24(6):E751-E757. <https://doi.org/10.1097/MJT.0000000000000389>

49. Tai FWD, McAlindon ME. Non-steroidal anti-inflammatory drugs and the gastrointestinal tract. *Clinical Medicine (Northfield IL)*, 2021; 21(2):131. <https://doi.org/10.7861/clinmed.2021-0039>

50. Ballinger A. Cytoprotection with misoprostol: Use in the treatment and prevention of ulcers. *Digestive Diseases*, 1994; 12(1):37-45. <https://doi.org/10.1159/000171435>

51. Zhang W, Lian Y, Li Q, et al. Preventative and therapeutic potential of flavonoids in peptic ulcers. *Molecules*, 2020; 25(20):4626. <https://doi.org/10.3390/molecules25204626>

52. Kunnumakkara AB, Hegde M, Parama D, et al. Role of turmeric and curcumin in prevention and treatment of chronic diseases: Lessons learned from clinical trials. *ACS Pharmacology & Translational Science*, 2023; 6(4):447-471. <https://doi.org/10.1021/acptsci.2c0012>

53. Mao QQ, Xu XY, Cao SY, et al. Bioactive compounds and bioactivities of ginger (*Zingiber officinale* Roscoe). *Foods*, 2019; 8(6):185. <https://doi.org/10.3390/foods8060185>

54. Sharifi-Rad M, Roberts TH, Matthews KR, et al. Ethnobotany of the genus *Taraxacum*—phytochemicals and antimicrobial activity. *Phytotherapy Research*, 2018; 32(11):2131-2145. <https://doi.org/10.1002/ptr.6157>

55. Sharifi-Rad M, Nazaruk J, Polito L, et al. *Matricaria* genus as a source of antimicrobial agents: From farm to pharmacy and food applications. *Microbiological Research*, 2018; 215:76-88. <https://doi.org/10.1016/j.micres.2018.06.010>

56. de Jesus NZT, de Souza Falcão H, Gomes IF, et al. Tannins, peptic ulcers and related mechanisms. *International Journal of Molecular Sciences*, 2012; 13(3):3203-3228. <https://doi.org/10.3390/ijms13033203>

57. Lukic J, Chen V, Strahinic I, et al. Probiotics or pro-healers? The role of beneficial bacteria in tissue repair. *Wound Repair and Regeneration*, 2018; 25(6):912-922. <https://doi.org/10.1111/wrr.12607>

58. Rahnama M, Mehrabani D, Japoni S, Edjtehadi M, Saberi-Firooz M. The healing effect of licorice (*Glycyrrhiza glabra*) on *Helicobacter pylori*-infected peptic ulcers. *Journal of Research in Medical Sciences*, 2013; 18(6):532-535. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3818629/>

59. Slippery elm. *LiverTox: Clinical and Research Information on Drug-Induced Liver Injury*. National Institute of Diabetes and Digestive and Kidney Diseases. Published January 5, 2024. <https://www.ncbi.nlm.nih.gov/books/NBK599741/>

60. Skoufou M, Tsigalou C, Vradelis S, Bezirtzoglou E. The networked interaction between probiotics and intestine in health and disease: A promising success story. *Microorganisms*. 2024;12(1):194. <https://doi.org/10.3390/microorganisms12010194>

61. Markowiak P, Ślizewska K. Effects of probiotics, prebiotics, and synbiotics on human health. *Nutrients*. 2017;9(9):1021. <https://doi.org/10.3390/nu9091021>

62. Blottière HM. The gut microbiota and obesity. In: *Energy Balance and Obesity*. 2017. Accessed March 24, 2025. <https://www.ncbi.nlm.nih.gov/books/NBK565809/>

63. Le Chatelier E, Nielsen T, Qin J, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013;500(7444):541–546. <https://doi.org/10.1038/nature12506>

64. Crudele L, Gadaleta RM, Cariello M, Moschetta A. Gut microbiota in the pathogenesis and therapeutic approaches of diabetes. *eBioMedicine*. 2023;97:104821. <https://doi.org/10.1016/j.ebiom.2023.104821>

65. Boicean A, Birlutiu V, Ichim C, Anderco P, Birsan S. Fecal microbiota transplantation in inflammatory bowel disease. *Biomedicines*. 2023;11(4):1016. <https://doi.org/10.3390/biomedicines11041016>

66. Choi KW, Chen CY, Stein MB, et al. Assessment of bidirectional relationships between physical activity and depression among adults: A 2-sample Mendelian randomization study. *JAMA Psychiatry*. 2019;76(4):399–408. <https://doi.org/10.1001/jamapsychiatry.2018.4175>