

**Figure 2: Chemical structures of related substances (Impurities) of Apalutamide**

## MATERIALS AND METHODS

Apalutamide and other related substances IMP-1, IMP-2, IMP-3 and IMP-4 were of AR grade. Waters Alliance HPLC system with PDA/UV detector (Model No. 2996 and 2695), Digital Ultra Sonicator (Labman) (Model No. LMUC-3), analytical balance (Sartorius) (Model No. BT224S) and pH Meter (Polmon) (Model No. LP139SA) were used for the study. Ammonium phosphate (SRL), Ortho phosphoric Acid (Avra) HPLC grade Acetonitrile (Merck), hydrochloric acid, sodium hydroxide and hydrogen peroxide (30% w/v) were used. Milli-Q water and Millipore 0.45  $\mu$ m membrane filters were used for the entire study.

### Procedure

#### Preparation of mobile phase

1.32 grams of Ammonium phosphate was weighed and transferred into a 1000 mL volumetric flask, dissolved in Milli-Q water and pH was adjusted to 7.0 with Ammonia solution and made up to the volume with the diluent (Acetonitrile). A

mixture of Ammonium phosphate buffer solution and Acetonitrile is used as the mobile phase on gradient mode.

#### Preparation of standard stock solutions

3.0 mg of each of IMP-1, IMP-2, IMP-3, IMP-4 and 2.0 mg of Apalutamide standard were weighed and transferred into a 100 mL volumetric flask, dissolved and made up to the volume with diluent (Acetonitrile) and further dilutions were made as per the requirement.

#### Preparation of test solution

10.0 mg of each of IMP-1, IMP-2, IMP-3, IMP-4 and Apalutamide samples were weighed, transferred into a 10 mL volumetric flask and diluted with diluent.

#### Method validation

#### Optimized chromatographic conditions

A mixture of Ammonium phosphate buffer solution and Acetonitrile (30: 70, v/v) was used as the mobile phase on gradient mode for the chromatographic study (Flow rate: 1.0

mL/min; UV detection: 243 nm; Injection volume: 20  $\mu$ L). Waters Alliance HPLC system with PDA/UV detector (Model No. 2996 and 2695) was used for the chromatographic study and the run time was 40 min.

### Linearity

Drug solutions containing Apalutamide and its impurities (5-40  $\mu$ g/mL) were prepared n from their stock solutions and injected into the HPLC system (n=3) and the corresponding chromatograms were recorded. The peak area of Apalutamide and its impurities (IMP-1, IMP-2, IMP-3 IMP-4) were noted and calibration curves were drawn by plotting the concentration of Apalutamide and its impurities (IMP-1, IMP-2, IMP-3 IMP-4) solutions on the x-axis and the corresponding mean peak area on the y-axis.

### Precision and Accuracy studies

Method precision was evaluated by test sample spiked with Apalutamide IMP-I, IMP-2, IMP-3 and IMP-4 at specification level with respect to the test sample concentration by injecting six different test preparations.

The accuracy of the method was proved by checking the recovery of known impurities. The test solution was spiked with the known impurities at LOQ, 50%, 100% and 200% level and the % RSD was calculated.

### Assay of Apalutamide tablets

Apalutamide is available as tablets (Label claim: 60 mg) with brand names APATIDE, APNAT, APALUCIDE, PRYOR, ERLEADA etc from different pharmaceutical companies. 20 tablets of two different companies were accurately weighed, powdered and powder equivalent to 25 mg of Apalutamide was carefully transferred to two separate 25 ml volumetric flasks and Apalutamide was extracted with HPLC grade acetonitrile, sonicated and filtered. These solutions were diluted further

with the mobile phase and 20  $\mu$ L of each of these formulation solutions was injected in to the system (n=3) and the average peak area was calculated from the respective chromatograms and thereby the amount of Apalutamide was calculated from the calibration curve.

### Stress degradation studies

Stress degradation studies of Apalutamide solution was carried out to confirm whether the known impurities are the degradation products or not.

For acidic degradation, the sample was treated with 2N hydrochloric acid at 80°C for 4 hours and then neutralized with 2N sodium hydroxide. For alkali degradation, the sample was treated with 2N NaOH at 80°C for 8 hours and then neutralized with 2N hydrochloric acid. For oxidative degradation, the sample was treated with 0.001N KMnO<sub>4</sub> on bench top for 30 min.

After degradation treatments the samples were cooled to room temperature, diluted with the diluent and injected for chromatographic analysis.

## RESULTS AND DISCUSSION

The authors have developed a new validated stability indicating RP-HPLC method for the quantification of Apalutamide and its related substances on gradient mode using Waters Alliance system (Model no. 2996 and 2695) with Inertsil ODS-3 (250  $\times$  4.6 mm, 5  $\mu$ m) column (PDA detector) (Column temperature 35°C) and a mixture of Ammonium phosphate buffer solution and Acetonitrile (UV detection 271 nm) was chosen for the present study. The injection volume was 20  $\mu$ L and the run time was 40 min. The analytical methods developed earlier for the estimation of Apalutamide summarised in Table 1 and the gradient program was given in Table 2.

**Table 1: Review of literature**

Method	Mobile phase (v/v)	Column	$\lambda$ (nm)	Comment	Ref
HPLC	0.01 M disodium phosphate dihydrate buffer (pH 4.20 $\pm$ 0.05): Acetonitrile	Luna Omega Polar C 18	225	Related substances (Gradient mode)	3
HPLC LC-MS/MS	10 mM KH <sub>2</sub> PO <sub>4</sub> (pH 3.5): Acetonitrile	Atlantis dC <sub>18</sub>	270	Design of experiments (Gradient mode)	4
LC-ESI-MS/MS	0.1% Formic acid: Acetonitrile (20:80)	Inertsil C18	--	Human plasma Canagliflozin as internal standard (Isocratic mode)	5
RP-HPLC	Ammonium phosphate buffer (pH 7.0): Acetonitrile (30: 70)	Inertsil ODS-3	243	4 Impurities (Gradient mode)	Present method

**Table 2: Gradient program**

Time (minutes)	Acetonitrile (%)	Buffer (%)
0.01	80	20
9	75	25
20	30	70
25	20	80
30	80	20
35	80	20

### Method validation<sup>6-8</sup>

#### Linearity, Precision and Accuracy

Linearity was conducted by preparing the five levels of linearity solutions for Apalutamide and for known impurities. The LOD and LOQ results were shown in Table 3 (Figure 2). The retention times of Apalutamide IMP-I, IMP-2, IMP-3 and IMP-4 were observed as 22.69, 3.48, 5.68, 16.34 and 16.83 min.

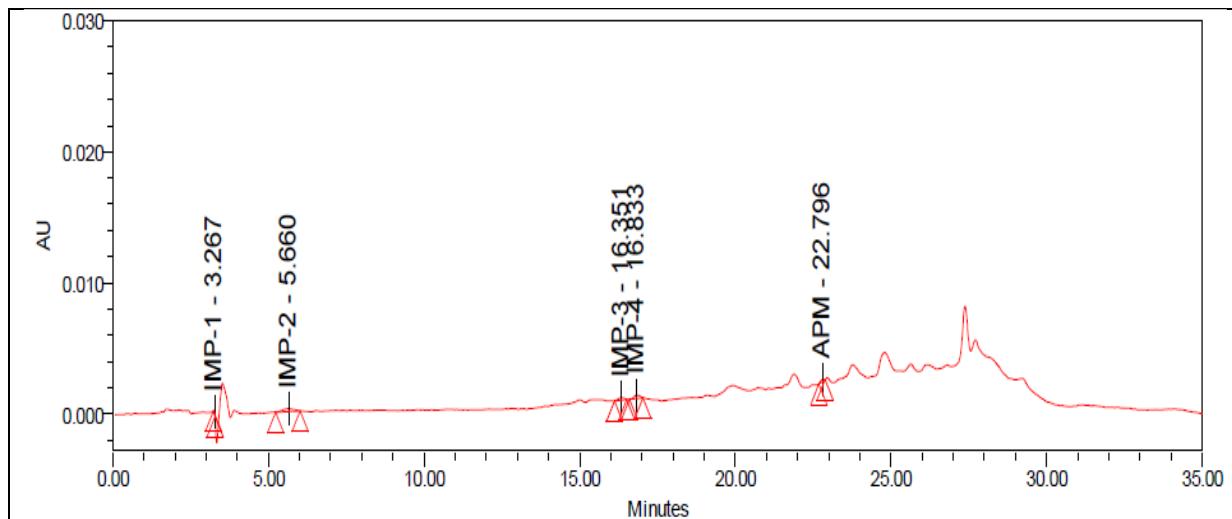


Figure 2A: Chromatogram for LOD solution

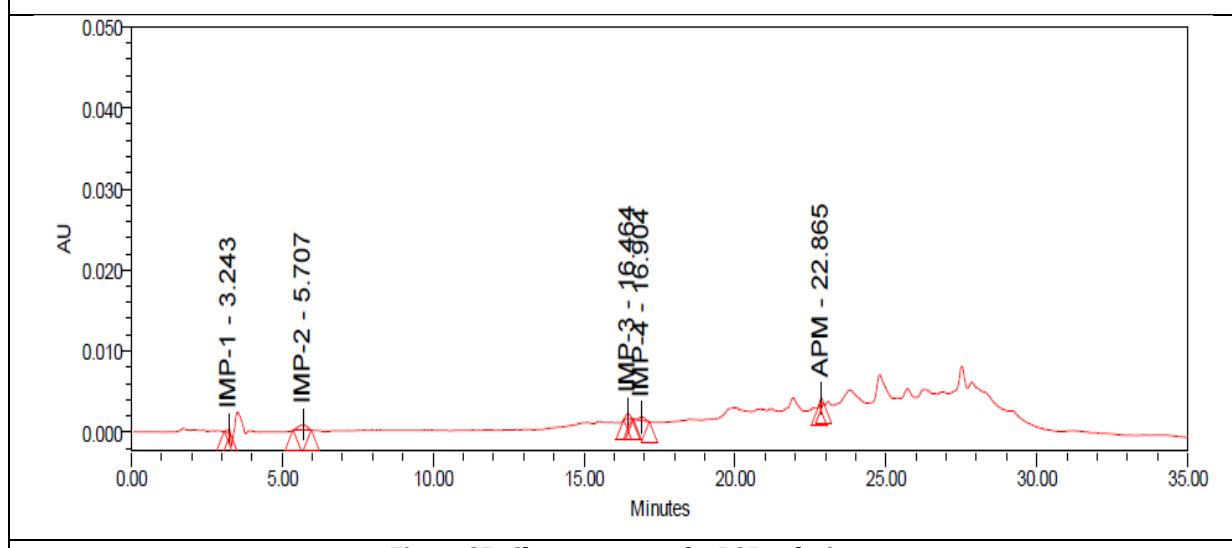


Figure 2B: Chromatogram for LOD solution

Table 3: Results of LOD &amp; LOQ

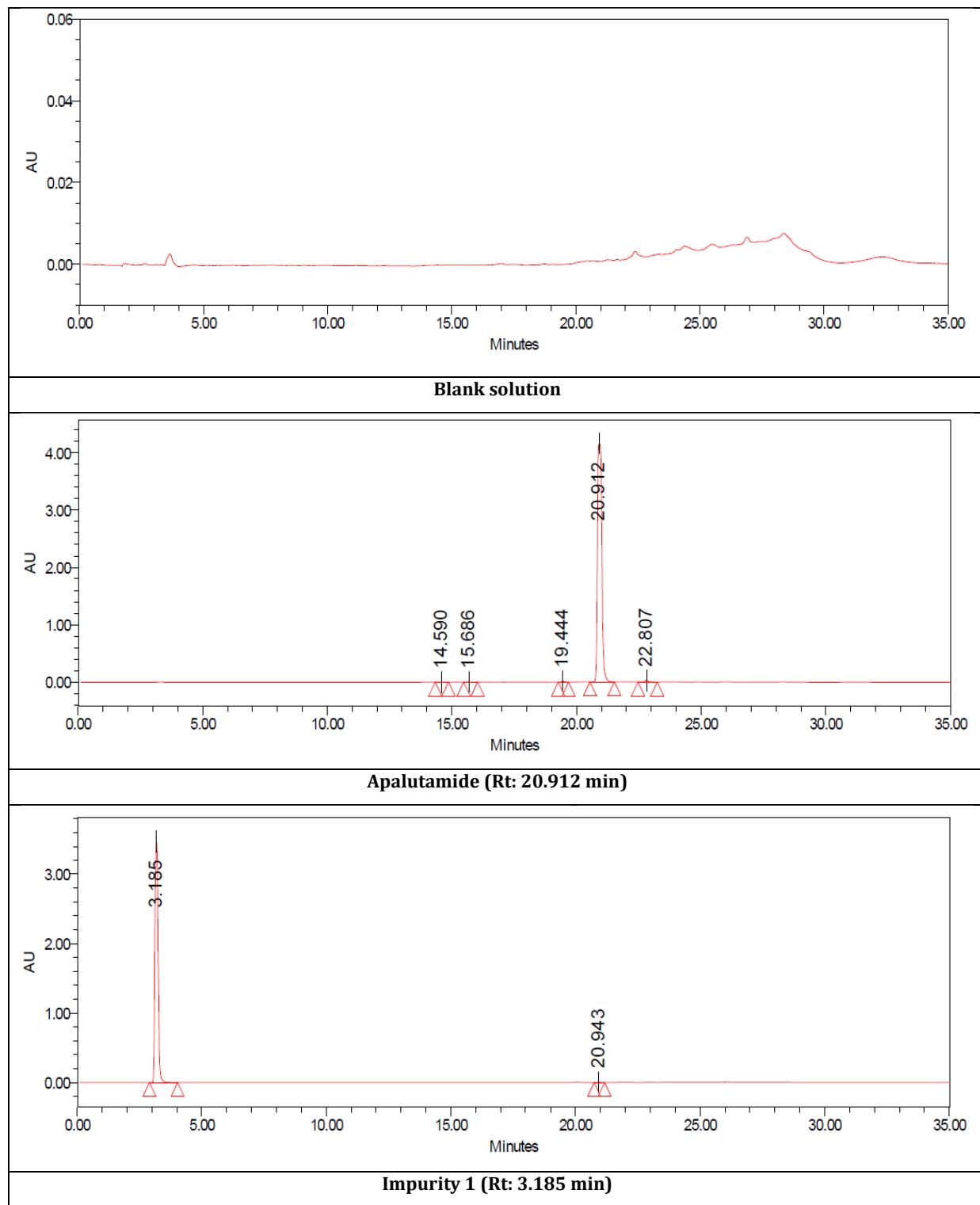
Injection	Limit of quantification				
	Peak area				
	IMP-1	IMP-2	IMP-3	IMP-4	Apalutamide
1	2985	13396	6332	6421	3956
2	2634	15407	7234	6530	4793
3	2856	13956	6395	6352	4025
4	2901	13263	7020	6329	4562
5	2790	13156	7325	6989	4489
6	2923	14434	6452	6524	4562
<b>Mean</b>	<b>2848</b>	<b>13935</b>	<b>6793</b>	<b>6421</b>	<b>4398</b>
%RSD	<b>3.96</b>	<b>5.68</b>	<b>6.06</b>	<b>3.40</b>	<b>6.90</b>
Limit of detection					
Injection	Peak area				
	IMP-1	IMP-2	IMP-3	IMP-4	Apalutamide
1	391	5713	2565	3227	1123
2	379	5625	2495	3312	1209
S/N Ratio					
LOQ	14.2	15.3	15.0	10.0	15.9
LOD	4.4	4.2	3.7	4.4	3.0

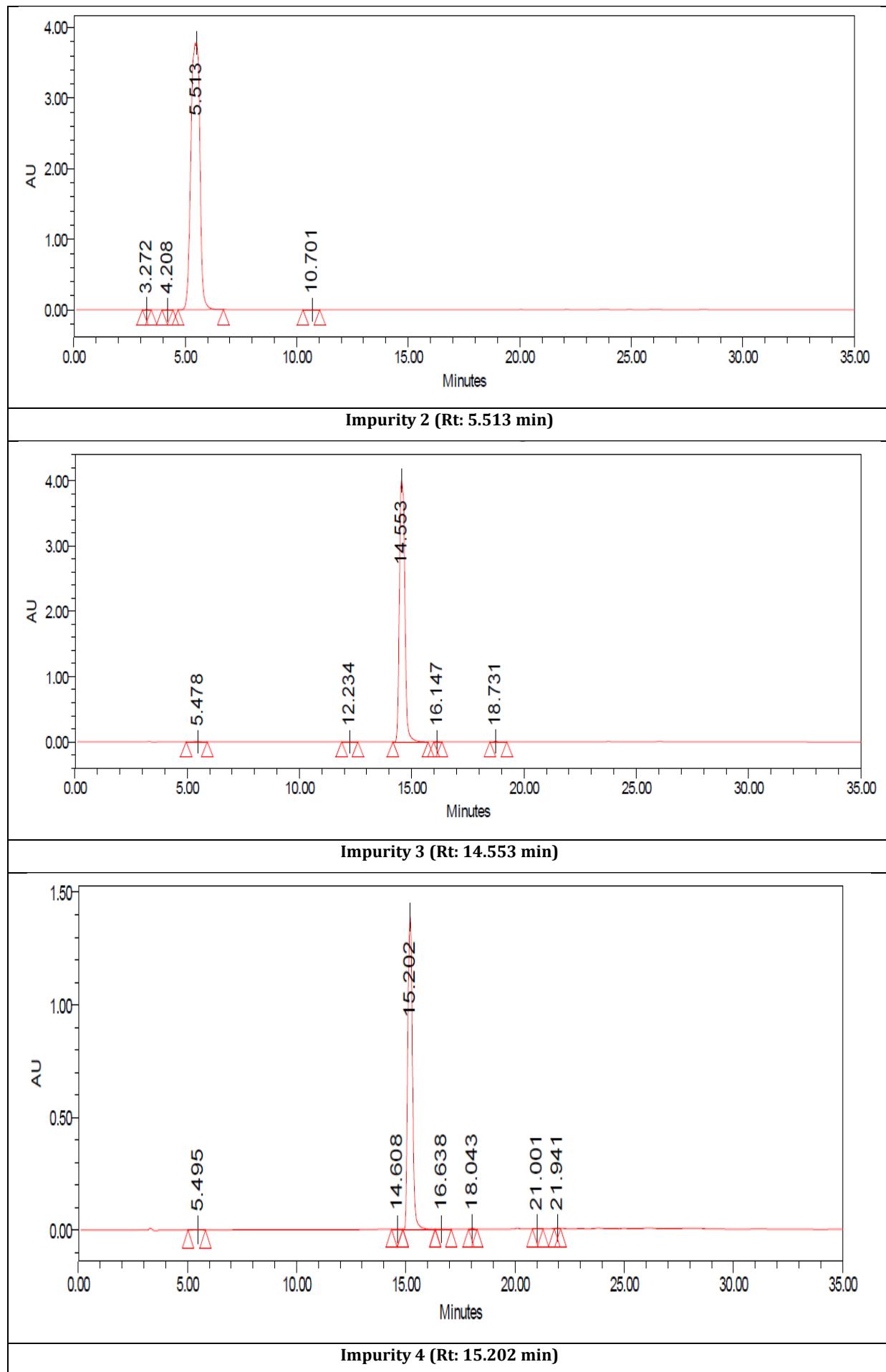
The representative chromatograms of Apalutamide and its four impurities were shown in Figure 3. A calibration graph was drawn by taking the concentration of Apalutamide IMP-I, IMP-2, IMP-3 and IMP-4 on the x axis and the corresponding peak area on the y axis (Figure 4).

**Table 4: Linearity of Apalutamide and impurities**

Conc. ( $\mu\text{g/ml}$ )	Apalutamide	IMP-1	IMP-2	IMP-3	IMP-4
	Mean peak area				
5	17781	13249	50088	44793	18933
10	36453	47474	101464	85426	39794
20	74932	93859	204377	167120	80599
30	113295	143835	290240	260196	131188
40	150509	193274	408962	341451	165925
Rt (min)	22.69	3.48	5.68	16.34	16.83

\*Mean of three replicates





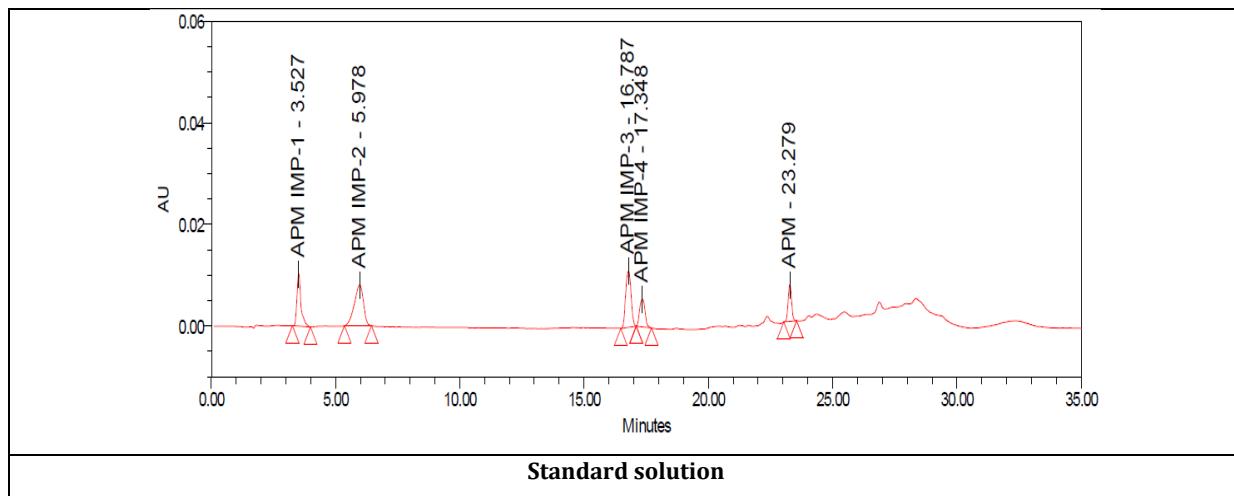


Figure 3: Representative chromatograms of Apalutamide and its impurities

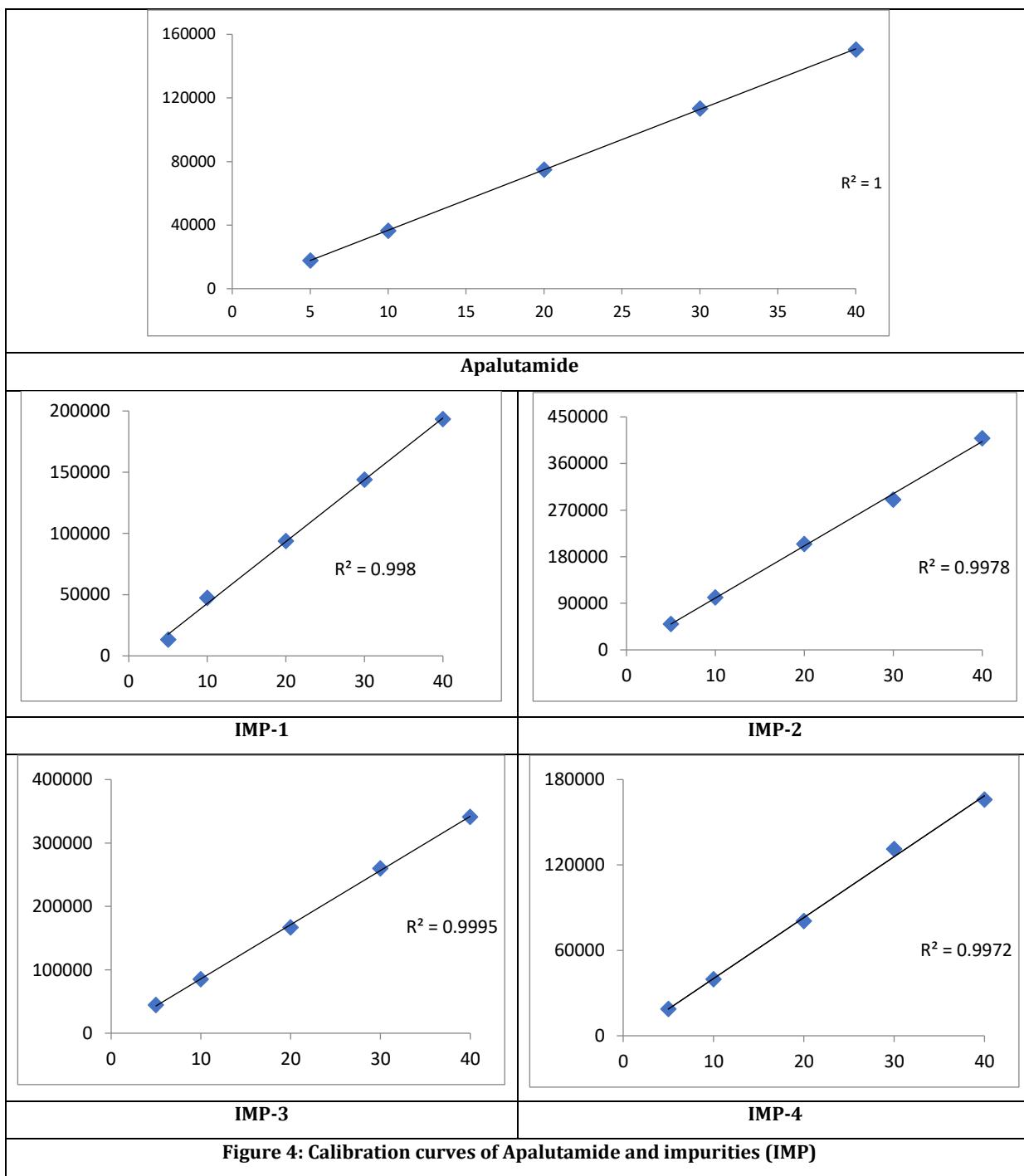


Figure 4: Calibration curves of Apalutamide and impurities (IMP)

The precision results of the impurities were shown in Table 5. The accuracy of the method was proved by checking the recovery of known impurities. Test solution was spiked with

known impurities at LOQ, 50%, 100% and 200% level and the obtained results were summarized in Table 6-9.

**Table 5: Precision study**

S. No.	IMP-1 (%)		IMP-2 (%)		IMP-3 (%)		IMP-4 (%)	
	Analyst 1	Analyst 2						
1	0.152	0.151	0.150	0.151	0.152	0.153	0.154	0.151
2	0.154	0.153	0.151	0.153	0.153	0.151	0.150	0.153
3	0.150	0.154	0.153	0.154	0.150	0.153	0.152	0.152
4	0.153	0.150	0.150	0.154	0.150	0.150	0.152	0.150
5	0.152	0.153	0.153	0.150	0.152	0.150	0.153	0.154
6	0.150	0.151	0.151	0.153	0.150	0.152	0.151	0.152
<b>Mean</b>	0.150	0.150	0.150	0.150	0.150	0.150	0.150	0.150
<b>%RSD</b>	<b>0.96</b>	<b>0.93</b>	<b>0.82</b>	<b>0.98</b>	<b>0.80</b>	<b>0.83</b>	<b>0.85</b>	<b>0.85</b>

**Table 6: Accuracy study of Impurity-1**

	IMP-1 % Recovery			
	% Recovery at LOQ Level	% Recovery at 50 % Level	% Recovery at 100 % Level	% Recovery at 200 % Level
	95.5	99.9	99.5	98.9
	96.1	100.2	100.2	100
	91.2	98.2	98.9	98.9
	93.9	99.3	99.6	99.6
	94.6	100.5	99.9	100.9
	92.1	99.4	99.9	99.4
<b>Mean %Recovery</b>	93.90	99.6	99.67	99.6
<b>Mean %RSD</b>	<b>1.87</b>	<b>0.75</b>	<b>0.41</b>	<b>0.69</b>

**Table 7: Accuracy study of Impurity-2**

	IMP-2 % Recovery			
	% Recovery at LOQ Level	% Recovery at 50 % Level	% Recovery at 100 % Level	% Recovery at 200 % Level
	97.6	99.7	100.5	99.5
	95.2	100.6	98.5	98.6
	99.1	98.0	99.3	99.9
	98.5	99.5	98.9	100.3
	98.5	100.8	100.1	99.6
	100.0	99.5	98.7	100.9
<b>Mean %Recovery</b>	98.2	99.7	99.3	99.8
<b>Mean %RSD</b>	<b>1.53</b>	<b>1.10</b>	<b>0.74</b>	<b>0.71</b>

**Table 8: Accuracy study of Impurity-3**

	IMP-3 % Recovery			
	% Recovery at LOQ Level	% Recovery at 50 % Level	% Recovery at 100 % Level	% Recovery at 200 % Level
95.1	98.1	99.3	100.5	
96.8	99.6	98.2	100.9	
97.0	99.0	100.2	99.1	
97.8	98.8	99.8	98.2	
98.0	100.9	100.8	99.8	
96.8	100.9	99.8	100.9	
<b>Mean %Recovery</b>	96.9	99.6	99.7	99.9
<b>Mean %RSD</b>	<b>0.97</b>	<b>1.05</b>	<b>0.81</b>	<b>0.99</b>

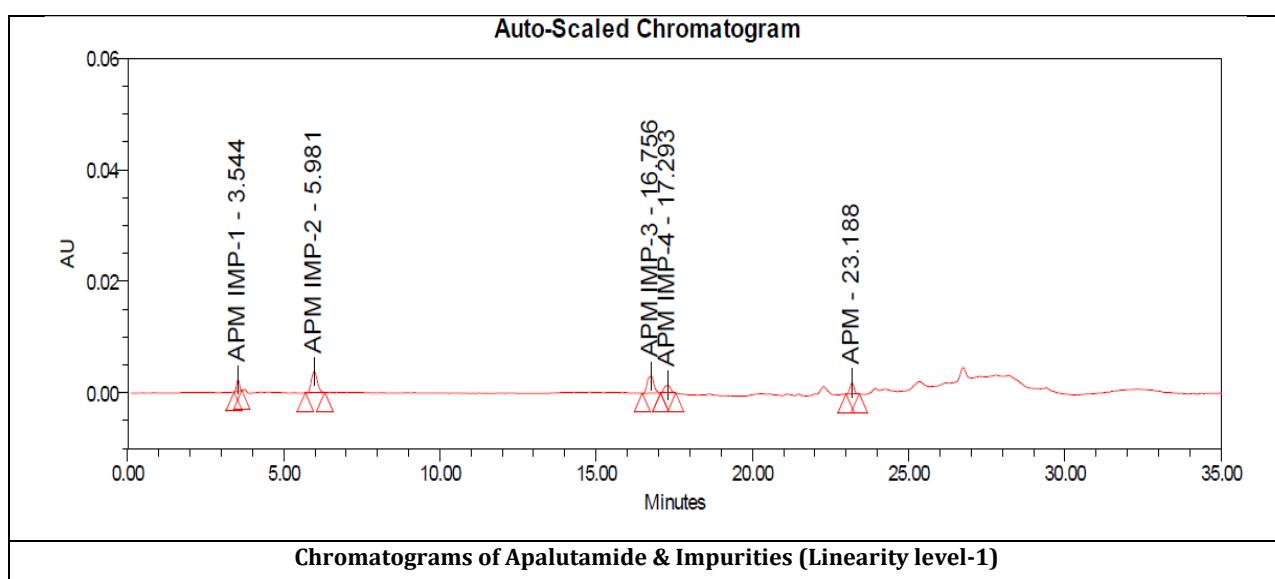
**Table 9: Accuracy study of Impurity-4**

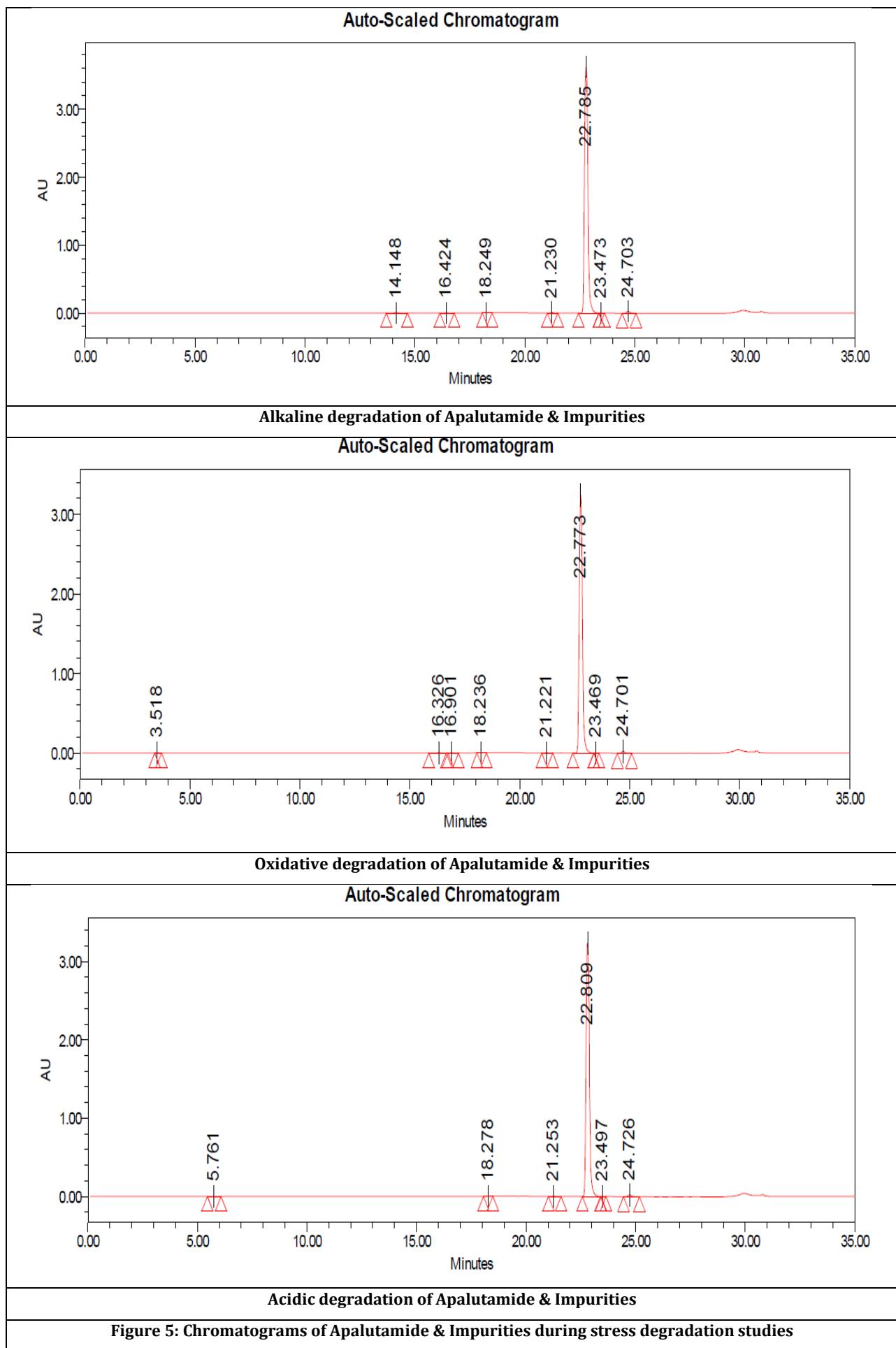
	IMP-4 % Recovery			
	% Recovery at LOQ Level	% Recovery at 50 % Level	% Recovery at 100 % Level	% Recovery at 200 % Level
99.3	100.2	99.7	100.5	
98.2	99.8	98.6	100.6	
99.9	98.5	98.5	99.9	
100.5	98.6	98.2	98.9	
99.5	99.6	99.9	100.9	
98.6	98.6	99.9	98.7	
<b>Mean % Recovery</b>	99.3	99.2	99.1	99.9
<b>Mean % RSD</b>	<b>0.77</b>	<b>0.68</b>	<b>0.72</b>	<b>0.85</b>

### Stress degradation studies

The pure drug Apalutamide API was eluted at  $22.69 \pm 0.1$  mins with theoretical plates ( $>2000$  and tailing factor  $<1.5$  which are within the acceptable criteria. The % degradation was found to be less than 2% and the system suitability parameters were

within the acceptable criteria. During the degradation studies of Apalutamide such as acidic, alkaline and oxidative degradation there is no interference of the drug peak indicating that the proposed method is highly specific and the resolution is more than 2.0 and the representative chromatograms were shown in Figure 5 and the results were shown in Table 10.





**Table 10: Stress degradation studies (ND: Not detected; NA: Not applicable)**

Degradation condition	IMP-1 (%)	IMP-2 (%)	IMP-3 (%)	IMP-4 (%)	Total impurities (% w/w)	% Net degradation
Unstressed sample	ND	ND	ND	ND	1.07	NA
Acidic degradation 2N HCl /80°C for 4 hrs	ND	0.06	ND	ND	0.97	0.97
Alkaline degradation 2N NaOH /80°C for 8 hrs	ND	ND	0.12	ND	1.29	1.29
Oxidative degradation 10% H <sub>2</sub> O <sub>2</sub> degradation at 80°C for 1hr	0.13	ND	ND	ND	1.27	1.27

## CONCLUSION

A simple, rapid, accurate and precise RP-HPLC method for the analysis of Apalutamide and its related substances has been developed and validated as per ICH guidelines. It is concluded that the Apalutamide related substances and preservative content method is validated and suitable for its intended purpose and it is suitable for the testing of in-vitro, related substances and preservative content samples of Apalutamide suspension 100 mg/mL and tablets during routine quality control and stability testing.

## ACKNOWLEDGEMENT

The authors are grateful to NATCO Pharma for supplying Apalutamide and related substances as gift samples.

## CONFLICTS OF INTEREST

The authors declare no conflict of interest.

## REFERENCES

1. Al-Salama ZT, "Apalutamide: First Global Approval" Drugs, 2018; 78: 699-705. <https://doi.org/10.1007/s40265-018-0900-z> PMid:29626324
2. Rathkopf DE, "Phase I study of ARN-509, a novel antiandrogen, in the treatment of castration-resistant prostate cancer" Journal of Clinical Oncology, 2013; 31(28): 3525-3530. <https://doi.org/10.1200/JCO.2013.50.1684> PMid:24002508 PMcid:PMC3782148
3. Bandaru LGR, Konduru N, Kowtharapu LP, Regulagadda S, Kanuparthi PR and Gundla R, "Development and validation of Apalutamide-related substances method in solid dosage forms using HPLC" Biomed Chromatography, 2023; 37(4): e5576. <https://doi.org/10.1002/bmc.5576> PMid:36573285
4. Lakka NS, Kuppan C, Vadagam N, Reddamoni SY, Muthusamy C, "Degradation pathways and impurity profiling of the anticancer drug Apalutamide by HPLC and LC-MS/MS and separation of impurities using Design of Experiments", Biomed Chromatography, 2023; 37(2): e5549. <https://doi.org/10.1002/bmc.5549> PMid:36409057
5. Sai Uday Kiran G and Sandhya P, "Method development and validation for the analysis of Apalutamide in human plasma by LC-MS/MS" International Journal of Current Research and Review, 2022; 14(4): 74-79. <https://doi.org/10.31782/IJCRR.2022.14413>
6. ICH Q2 (R1) Validation of analytical procedures: Text and methodology (2005).
7. ICH Q1A (R2) Stability testing of new drug substances and products (2003).
8. ICH Q3A (R2) Impurities of new drug substances (2006).