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Review Article

Contrasting Evidences Between High Cholesterol Levels and The Risk of Cardiovascular Disease: A Review

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Abstract



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Introduction: The cholesterol hypothesis had been kept alive for decades by reviewers who used statistics that excluded the results from unsuccessful trials and ignored numerous contradictory observations. High dietary cholesterol intake had been associated with development of cardiovascular diseases (CVD) and mortality, for over several decades, without a direct link between the CVD and high serum cholesterol level. Hence, people avoided healthy cholesterol-rich diets due to the fear of developing CVD. The relationship between elevated plasma cholesterol and CVD and criteria for appropriate methods for screening patients with elevated cholesterol had remained a source of medical debates. Lack of decrease in overall mortality rates in patients without clinical coronary disease in whom aggressive lowering of cholesterol was achieved might have contributed to the lack of consensus on this most important issue.

Methodology: With information derived from search engines, such as Elsevier, Springer, PubMed, Science Direct, Medline, Google Scholar and a library search for articles published in peer-reviewed journals.

Results: Several research results showed that association between total serum cholesterol, its components and CVD is weak, absent or inverse implying that consuming healthy high cholesterol diets may not be harmful to health.

Conclusion: This review provides evidence contrasting the links between elevated plasma cholesterol and CVD, and demonstrated that elevated cholesterol concentrations, rather, improve quality of life and life expectancy. In addition, most prevalent methods for cholesterol quantification in biological samples and foods, utilizing new technologies, such as Ambient Ionization Mass Spectrometry are summarized, along with other components of cholesterol.

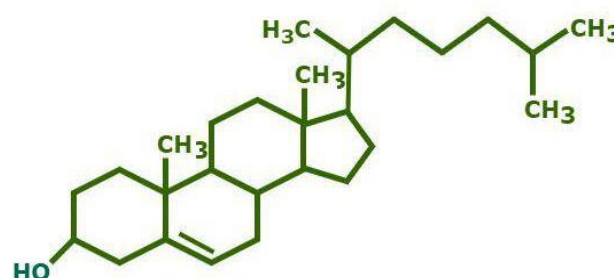
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Introduction

Cholesterol, a fat-like or waxy organic molecule made of sterol backbone with hydroxy group and 8 carbon atoms had been reported in the blood stream, body organs, nerve fibres and all animal products^{1,2,3,4}. Available reports^{5,6,7,8,9,10} showed that cholesterol, which composed of about 30% of all animal cell membranes, was required to build and modulate membrane fluidity over the range of physiological temperatures, maintain membranes integrity, cells flexibility, signaling and nerve conductivity. More than 90% of cellular cholesterol, located in the plasma membrane^{11,12}, served as a precursor of all steroid hormones including adrenal gland hormones, cortisol and aldosterone, progesterone, estrogens and testosterone and their derivatives^{13,14,15}, vitamin D and bile salt, responsible for intestinal digestion of dietary fats and absorption of fat-soluble vitamins A, D, E and K^{16,17,18,19}.

Cholesterol could either be synthesized *de novo* by many cells of the body (endogenous) or supplied from the diet (exogenous) and for the latter, the major dietary factors leading to hypercholesterolaemia include high intakes of

cholesterol itself, saturated fats and excessive calories. The liver was reported as one of the major sites of endogenous synthesis of cholesterol, from a wide variety of foods, especially saturated fats²⁰.



Formular: C₂₇H₄₆O Molecular mass: 386.65

Figure1: Structure of cholesterol,

The liver had long been reported to tightly regulate its pool of cholesterol by influencing the input from dietary cholesterol, the biosynthesis, secretion and uptake from plasma

lipoproteins, the conversion of cholesterol into bile, and the reuptake of biliary cholesterol and bile acids from the intestine to the liver²¹. All animal-based foods contained cholesterol in varying amounts²² from the ability of all animal cells to manufacture cholesterol. Major dietary sources of cholesterol were listed as red meat, egg yolks and whole eggs, liver, kidney, giblets, fish oil, butter and human breast milk²³. According to Sonawane *et al.*²³ and De Smet *et al.*²⁴, synthesis of cholesterol in plant cells occurred as a precursor for other compounds, such as phytosterols and steroidal glycoalkaloids, with cholesterol remaining in plant foods only in minor amounts or absent. Some plant foods, such as avocado, flax seeds and peanuts, contain phytosterols, competing with cholesterol for absorption in the intestines, thereby reducing the absorption of both dietary and bile cholesterol²⁵. Cholesterol and other blood fats are carried in the blood by proteins and after the combination of these proteins and fats, lipoproteins were the end products²⁶. There are 2 main types of Lipoproteins: low density lipoprotein (LDL) or 'bad cholesterol' and high-density lipoprotein (HDL) or 'good cholesterol'. The LDL was reported to carry cholesterol from the liver, through the blood stream to the cells and arteries of the body^{27,28} a phenomenon that was incriminated in promoting atherosclerosis by stimulating production of haematopoietic stem/progenitor cells²⁹. Hence, laboratory measurement of HDL was considered more significant for clinical diagnosis. Whereas, HDL removes cholesterol from the blood stream rather than depositing it in the arteries³⁰.

Physiology of Cholesterol

The esterification of fat via chylomicrons led to introduction of cholesterol into the blood³¹, which causes its poor absorption by the gut. The body also compensated for absorption of ingested cholesterol by reducing its own cholesterol synthesis³². For these reasons, cholesterol in food, 7 to 10 hours after ingestion, has little, if any effect on concentrations of cholesterol in the blood. However, during the first 7 hours after ingestion, as absorbed fats are being distributed around the body within extracellular water by the various lipoproteins, the concentrations increased³³ with the liver excreting it via bile in a non-esterified form into the digestive tract. Behrman and Gopalan³⁴ reported that about 50% of the excreted cholesterol was reabsorbed by the small intestine back into the bloodstream. According to Jones³⁵, increased dietary cholesterol absorption led to decreased endogenous synthesis, thus, maintaining cholesterol balance. While dietary triglycerides were almost completely absorbed, only 30-50% of the cholesterol present in the intestinal lumen was absorbed^{36,37,38}.

Pathophysiology of cholesterol

Atherosclerotic process commenced with endothelial damage, dysfunction of endothelial cells and consequential increase in LDL particle which permeated through the vascular wall and got trapped by cellular matrix in the intima with smooth muscle cells migration into the lesion ultimately encapsulation and protection of the newly formed plaque from exposure occurred; the plaque may occlude or reduce laminal flow of blood causing ischaemia or ruptured to form thrombus and could lead to unstable angina and/or acute myocardial infarction (AMI)^{39,40}.

Contrasting Research on Cholesterol Values and the Controversies

Early last century, Anichkov observed that rabbits fed pure cholesterol dissolved in sunflower oil developed atherosclerotic lesions, whereas the control rabbits fed just sunflower oil did not^{41,42,43}; the latter was largely rejected with no followed-up attempt. Serious research on the role of

cholesterol in atherosclerosis got under way when laboratories later tried to reproduce the results using dogs and rats but failed to link cholesterol rich diet with the development of atherosclerosis, because dogs and other carnivores metabolize cholesterol differently from rabbits and other herbivores. This led to the dismissal of Anichkov's results speculating that rabbits were not a good model for human physiology and not relevant to humans and other animals. For half a century, a high level of total cholesterol (TC) or LDL cholesterol (LDL-c) was considered as the major cause of atherosclerosis and cardiovascular diseases (CVD) which implied that individuals with high TC should have a higher risk of death from CVD⁴⁴. The hypothesis that high TC causes CVD was introduced in the 1960s by Framingham Heart Study group with a 30-year follow-up study which reported that for each 1 mg/dL drop in TC per year, there was an 11% increase in coronary heart disease (CHD) and total mortality⁴⁵. In contrast, about 3 years later, the American Heart Association and the U.S. National Heart, Lung and Blood Institute jointly published a summary, concluding that a 1.0% reduction in an individual's TC resulted in an approximately 2.0% reduction in CHD risk⁴⁶. This apparent controversy is supported by the notion that high blood cholesterol levels mainly caused CVD was found debatable because people with low levels were just as atherosclerotic as those with high levels and their risk of developing CVD was reported to be the same or higher⁴⁷. Much earlier, Landé and Sperry⁴⁸ found, in correlation with age, that unselected low TC people were as atherosclerotic as high TC people. Indeed, there was a lack of association between LDL-c and degree of atherosclerosis⁴⁹. In addition, an Austrian study that included 67,413- and 82,237- men and women, respectively, follow up for many years, revealed that TC was weakly associated with CHD mortality for men, except for those within the 50 to 64 years bracket, while for women, it was weakly associated among those below 50 years old with no association after that age⁵⁰. There was no association between TC and other CVD-induced mortality, except that low TC was inversely associated with CVD mortality for women above the age of 60 years. If high LDL-c causes CVD, LDL-c of untreated patients with CVD should be higher than normal. If the speculation that high LDL-c caused CVD is accepted, it would be expected that LDL-c of untreated CVD patients should be higher than normal, but in contrast, a large American study, that included almost 140,000 patients with AMI revealed that their LDL-c at the time of admission into the hospital, was actually lower than normal⁵¹. Al-Mallah *et al.*⁵² reported similar results, but lowered the patients' LDL-c even more, a 3-year follow-up revealed that total mortality among those with LDL-c below 105 mg/dL (2 mmol/L) was twice as high compared to those with a higher LDL-c, adjustment for confounding variables (14.8% vs. 7.1%, $p = 0.005$), notwithstanding. Weak association between TC and degree of atherosclerosis occurred in patients admitted into a hospital⁵³ who might have included patients with familial hypercholesterolemia (FH). As the percentage of such patients in cardiology department was much higher than that obtained in the general population, a bias might have been introduced. If high TC constituted the major cause of atherosclerosis, an exposure-response should be expected to follow in cholesterol-lowering drug trials; for example, the arteries of those whose lipid values were lowered should benefit the more. The calculated exposure-response in a review of 16 angiographic exercise induced cholesterol-lowering trials, revealed a correlation in only one present. Lewington⁵⁴, in a meta-analysis performed by the prospective studies collaboration, there was an association between TC and cardiovascular mortality in all ages and both sexes; the risk decreased with increasing age and became minimal after the age of 80 years. This appears contrasting, since atherosclerosis and CVD were considered mainly the diseases

of elderly, which implied that the cholesterol hypothesis, expectedly, would predict that the disease should be more prevalent in the elderly than in the young, which disagreed with the above finding.

Epidemiological analysis revealed that lack of association existed between dietary cholesterol and/or egg intake and CVD risk, generally in population^{55,56} in contrast to a consistent relationship that existed between egg intake and CVD in Diabetics^{15,55,56}; this risk in the diabetics was ascribed to phosphatidylcholine content of eggs⁵⁷ and not the dietary cholesterol that are more poorly absorbed in obese and insulin-resistant populations compared to lean individuals⁵⁸. The phosphatidylcholine intake was linked to the gut microbial-dependent production of trimethylamine N-oxide (TMAO), a metabolite that promoted atherosclerosis in hyperlipidemic mouse model with an association with CVD risk in human cohort studies⁵⁹. Indeed, a systematic review and meta-analysis of 17 cohort studies examining the relationship between dietary cholesterol and CVD⁶⁰ dietary cholesterol was significantly associated with heart disease, ischaemic stroke or haemorrhagic stroke. In addition, no association existed between LDL-c and coronary calcification in a study that included 304 patients⁶¹. However, a notable exception occurred in 1779 healthy individuals without conventional risk factors for CVD, where LDL-c was significantly higher among those with subclinical atherosclerosis (125.7 vs.117.4 mg/dL), an association that did not prove causation, as mental stress, for instance, was able to raise cholesterol by 10–50% within half an hour^{62,63}, in addition to causing atherosclerosis by mechanisms other than an increase in LDL-c; for instance, via hypertension and increased platelet aggregation. In Japan, the prevalence of heterozygous FH was found to be 5.7% (1/17.5) in a total of 359 patients with acute coronary syndrome, but more prominent in younger patients who were less than 60 years old (7.8%)⁶⁴.

However, Zhong *et al.*⁶⁵ reported associations of dietary cholesterol (egg consumption) with incident CVD and mortality using 29 615 adults pooled from 6 prospective cohort studies in the US with a median follow-up of 17.5 years; each additional 300 mg of dietary cholesterol consumed per day was significantly associated with higher risk of incident CVD and each additional half an egg consumed per day was significantly associated with higher risk of incident and all-cause mortality in a dose-response manner. This study had several limitations. First, appropriate interpretation of the study findings required consideration of measurement error for self-reported diet data. Instead of using different dietary assessment tools to create heterogeneities for data analyses, the study relied on single measurement of egg and dietary cholesterol consumption. Exposure misclassification might be of concern, but results were similar when censoring participants at different time points⁶⁵.

In animals, cholesterol is produced endogenously with only about 25% of the serum cholesterol derived from the diet. An average 70 kg adult synthesizes about 850 mg cholesterol per day. If this individual is to consume 400 mg/day of dietary cholesterol and absorb 60% that amounts to only 22% of cholesterol coming from the diet. Furthermore, the values are skewed even more towards cholesterol biosynthesis in overweight and obese individuals⁵⁸ studies evaluating the association of low LDL-c with safety outcomes, Faselis *et al.*⁶⁶ found that achieving an LDL-c of 40-50 mg/dL seems to be safe, and importantly might offer CV beneficial effects.

The writing committee of the studies of Lewington⁵⁴ and Collins *et al.*⁶⁷, published a meta-analysis of 61 prospective observational studies that involved about 900,000 adults with the conclusion that TC was associated with CHD mortality in

all ages and both sexes. In contrast, Ravnkov *et al.*⁶⁸, found the opposite in a systematic review of 19 cohort studies that involved more than 68,000 elderly individuals above 60 years of age. In a larger cohort study consisting 118,160 subjects greater than 50 years, individuals with the highest LDL-c levels were found to live even longer than those on statin treatment⁶⁹. In addition, Hamazaki *et al.*⁷⁰ reported that numerous Japanese epidemiological studies found high LDL-c was not a risk factor for CHD mortality in women of any age, and further, there was an inverse association between TC and all-cause mortality, irrespective of age and sex. Benn⁷¹ used genetic variants in proprotein convertase subtilisin-kexin type 9 (PCSK9) and a variant in HMGCR, to examine the causal effect of low LDL-c levels on risk of neurological disease and did not observe any associations with increased risk of Alzheimer's disease, vascular dementia, any dementia, and Parkinson's disease in 111,194 individuals from the general population. Data on 380 genetic variants from the International Genomics of Alzheimer's Project were, however, suggestive of a causal effect of low LDL-c levels in reducing the risk of Alzheimer's disease.

Researchers have over the years, repeatedly fed laboratory animals large amounts of cholesterol in their diets^{72,73,74}, expecting this to produce vascular accidents. It did not, which showed that despite the presence of cholesterol in atheromatous plaques these lesions are not caused by eating cholesterol. These animal studies have demonstrated that inflammatory processes tightly regulate the developmental processes of lesions, with significant contributions from both adaptive and innate immune processes^{75,76,77,78}. Other supports for the above hypotheses were derived from the fact that CVD might be caused by infections and that LDL directly bound to inactivate a broad range of micro-organisms and their toxic products^{44,79,80}. Another explanation for an inverse association between LDL-c and mortality was the report that high LDL-c, protected against cancer, as many cancers were caused by viruses⁸¹. Furthermore, cholesterol lowering experiments on rodents resulted in cancer⁸². In addition, 9 cohort studies including more than 1440,000 individuals followed for 10-30 years found an inverse association between cancer and TC, measured at a start of the study and more patients with cancer were on cholesterol lowering treatment, irrespective of their age and sex⁵³. For these reasons, it will be of value to state that there is an inverse association or lack of association between cholesterol and CVD and mortality.

Measurements of Cholesterol Level

Various analytical methods for determination of cholesterol levels (Table 1) have been developed including Gas Chromatography-Mass Spectrometry (GC-MS), Gas Chromatography Isotope-Dilution Mass Spectrometry (GC-ID-MS), Liquid chromatography-mass spectrometry (LC-MS), Electrospray Ionization Tandem Mass Spectrometry (ESI-MS), Electrospray Ionization tandem mass spectrometry (ESI-MS), Matrix-assisted laser desorption/ionization-time of flight (MALDI-MS), and Matrix-assisted laser desorption/ionization-mobility mass spectrometry (MALDI-IM-MS), with serum and plasma as the acceptable specimens^{83,84} and Ambient ionization mass spectrometry techniques (AIMS). The AIMS had been proposed as alternative approaches for high-throughput screening as they did not require tedious sample pretreatments. However, with the new diagnostic technique or imaging modality such as the performance of intravascular photoacoustic imaging, which is an emerging plaque detection technique that provides lipid specific chemical information from an arterial wall with great optical contrast and long acoustic penetration depth⁸⁵. Direct Analysis in Realtime Mass Spectrometry (DART-MS) was developed for rapid, cost-

effective and increase the understanding of physiological processes and disease pathology towards a more precise absolute quantitation of cholesterol. Matrix-Assisted Laser Desorption/Ionization Mass Spectrometry Imaging (MALDI-MSI) and Desorption Electrospray Ionization Mass Spectrometry Imaging (DESI-MSI) techniques were used in

tissue sections, to observed spatial distribution of cholesterol and its derivatives. Mass spectrometry-based molecular imaging increases the understanding of physiological processes and pathology towards to a more precise absolute quantitation of molecules in it.

Table 1: Methods for cholesterol measurements

Method	Principles	Ref
Modified Abell-Kendall method	Saponification and colour development	86
Fluorometric-enzymatic assay	Hydrosis of cholesterol to produce hydrogen peroxide (H ₂ O ₂) which can be quantified	87
GC-MS	Separation of cholesterol using the density and dispersal in gas medium and quantification by mass spectroscope	88, 89
GC-ID-MS	Comparing the sample with a standard cholesterol of known concentration which is an isotopically labelled cholesterol in gas medium and quantification using mass spectroscope	90
LC-MS	Separation of cholesterol using the density and distribution or dispersal in a liquid phase and quantification with mass spectroscope	91
ESI-MS	Conversion of cholesterol to ionic form and quantification	90
MALDI-MS	Removal of cholesterol, its dehydration, ion mobility and ultimate quantification using mass spectroscopy	92
MALDI-IM-MS	Coating of cholesterol with metal to enhance its ion mobility and quantification with mass spectroscope	93
DESI-MSI	Formation of cholesterol ion and quantification	94
DART-MS	Dehydration of cholesterol and the spectrometric quantification	95,96
MALDI-MSI	Embedding cholesterol in a matrix the spectrometric imaging and quantification in tissues	97
DESI-MSI	Reaction of cholesterol with Betained aldehyde to produce coloured a background in organs that can be imaged	98

Conclusion

The multifaceted path by which cholesterol became linked to CHD is one of the greatest biomedical stories of the 20th century. The current review does not support the notion that dietary cholesterol increases the risk of CVD. The idea that high cholesterol levels in the blood constitute the main cause of CVD is doubtful because people with low levels become just as atherosclerotic as people with high levels and their risk of suffering from CVD is the same or higher. In reality, the accelerated vascular damage seen in CVD is likely to be a consequence of more brittle arterial cell walls, as cholesterol to be a component of them which modulates their fluidity, conferring flexibility and hence resistance to damage from the ordinary hydrodynamic blood forces. In the absence of efficient receptors for LDL cholesterol, cells will be unable to use this component adequately for the manufacture of normally resilient arterial cell walls, resulting in accelerated arteriosclerosis. Eating high cholesterol diet is harmless, shown by its failure to produce vascular accidents in laboratory animals, but its avoidance causes malnutrition from lack of fat-soluble vitamins, especially vitamin D. Analytical differences may account for variations in laboratory results that may account for variations in interpretations.

Conflicts of interest

None.

References

- Vella CA, Kravitts L, Janot JM, A review of the impact of exercise on cholesterol levels. IDEA Health & Fitness Source, 2011; 19(10): 48-54.
- Laurence GD, Dietary fats and health: dietary recommendations in the context of scientific evidence. Advance in nutrition, 2013; 4(3): 294-302. <https://doi.org/10.3945/an.113.003657> PMID:23674795 PMCID:PMC3650498
- McLaughlin MA, Cardiovascular care made incredible easy! Lippincott Williams & Wilkins, 2019.
- Wali H, Rehman FU, Umar A, Ahmed S, Cholesterol degradation and production of extracellular cholesterol oxidase from *Bacillus pumilus* W1 and *Serratia marcescens* W8. Biomed research international, 2019; pp 9. <https://doi.org/10.1155/2019/1359528> PMID:31183360 PMCID:PMC6512041
- Ohvo-Rekilä H, Ramstedt B, Leppimäki P, Slotte JP, "Cholesterol interactions with phospholipids in membranes". Progress in Lipid Research, 2002; 41 (1): 66-97. [https://doi.org/10.1016/S0163-7827\(01\)00020-0](https://doi.org/10.1016/S0163-7827(01)00020-0) PMID:11694269
- Cortes VA, Busso D, Maiz A, Arteaga A, Nervi F, Rigotti A, Physiological and pathological implications of cholesterol. Frontiers in Bioscience, 2014; 19: 416-428. <https://doi.org/10.2741/4216> PMID:24389193
- Maekawa M, Fairn GD, Complementary probes reveal that phosphatidylserine is required for the proper transbilayer distribution of cholesterol. Journal of Cell Science, 2015; 128: 1422-1433. <https://doi.org/10.1242/jcs.164715> PMID:25663704

8. Alphonse PAS, Jones PJH, Revisiting human cholesterol synthesis and absorption: the reciprocity paradigm and its key regulators. *Lipids*, 2016; 51: 519-536. <https://doi.org/10.1007/s11745-015-4096-7> PMID:26620375
9. Hendrani AD, Adesiyun T, Quispe R, Jones SR, Stone NJ, Blumenthal RS, Martin SS, Dyslipidemia management and primary prevention of cardiovascular disease: Current guidelines and strategies. *World Journal of Cardiology*, 2016; 26(2): 201-210. <https://doi.org/10.4330/wjc.v8.i2.201> PMID:26981215 PMCid:PMC4766270
10. Cartocci V, Servadio M, Trezza V, Pallottini V, Can cholesterol metabolism modulation affect brain function and behavior? *Journal of Cellular Physiology*, 2017; 232: 281-286. <https://doi.org/10.1002/jcp.25488> PMID:27414240
11. Goluszko P, Nowicki B, Membrane cholesterol: a crucial molecule affecting interactions of microbial pathogens with mammalian cells. *Infection and Immunity*, 2005; 73: 7791-7796. <https://doi.org/10.1128/IAI.73.12.7791-7796.2005> PMID:16299268 PMCid:PMC1307024
12. Song Y, Kenworthy AK, Sanders CR, Cholesterol as a co-solvent and a ligand for membrane proteins. *Protein Science*, 2014; 23: 1-22. <https://doi.org/10.1002/pro.2385> PMID:24155031 PMCid:PMC3892294
13. Lloyd-Jones DM, Goff DC, Stone, NJ, Treatment of blood cholesterol to reduce risk for atherosclerotic cardiovascular disease. *Annals of Internal Medicine*, 2016; 164(2): 135-136. <https://doi.org/10.7326/L15-0521> PMID:26784487
14. Mauvais-Jarvis F, Sex and gender factors affecting metabolic homeostasis, diabetes and obesity. Springer International Publishing, Cham, 2017; vol.1043. <https://doi.org/10.1007/978-3-319-70178-3>
15. Benjamin BK, Qiu C, Han Z, Lu W, Sun G, Qin X, Wang X, Wang X, Li R, Liang P, The Association between type-2 diabetes duration and major adverse cardiac events after percutaneous coronary intervention. *Journal of Diabetes Research*, 2021; 1-9. <https://doi.org/10.1155/2021/7580486>
16. Boyer JL, Bile formation and secretion. *Comprehensive Physiology*, 2013; 3: 1035-1078. <https://doi.org/10.1002/cphy.c120027> PMID:23897680 PMCid:PMC4091928
17. Faridi KF, Lupton JR, Martin SS, Banach M, Quispe R, Kulkarni K, Vitamin D, deficiency and non-lipid biomarkers of cardiovascular risk. *Archives of Medical Science*, 2017; 13: 732-737. <https://doi.org/10.5114/aoms.2017.68237> PMID:28721139 PMCid:PMC5510501
18. Harvard Health, "How it's made: Cholesterol production in your body - Harvard Health" 2018.
19. Ibrahim MA, Jialal I, Hypercholesterolemia stat pearls, 2019; 6(1): 16-23.
20. Hemler EC, Hu FB, Plant-based diet s for cardiovascular disease prevention: all plant foods are not created equal. *Current atherosclerosis reports*, 2019; 21(5): 18. <https://doi.org/10.1007/s11883-019-0779-5> PMID:30895476
21. Arnold DR, Kwiterovich PO, Cholesterol: Absorption, Function, and Metabolism. in *Encyclopedia of Food Sciences and Nutrition (Second Edition)*. 2003; 1226-1237. <https://doi.org/10.1016/B0-12-227055-X/00225-X>
22. United States Department of Agriculture (USDA), Cholesterol content in foods, rank order per 100 g; In: *USDA Food Composition Databases*, 2019.
23. Sonawane PD, Pollier J, Panda S, Szymanski J, Massalha H, Yona M, "Plant cholesterol biosynthetic pathway overlaps with phytosterol metabolism". *Nature Plants*, 2016; 3(1): 16205. <https://doi.org/10.1038/nplants.2016.205> PMID:28005066
24. De Smet E, Mensink RP, Plat J, Effects of plant sterols and stanols on intestinal cholesterol metabolism: suggested mechanisms from past to present. *Molecular Nutrition & Food Research*, 2012; 56(7): 1058-72. <https://doi.org/10.1002/mnfr.201100722> PMID:22623436
25. European Food Safety Authority (EFSA), "Scientific opinion on the substantiation of health claims related to plant sterols and plant stanols and maintenance of normal blood cholesterol concentrations", 2010.
26. Ogru E, Libinaki R, West SM, U.S. patient application, 2009; No. 11/817,453.
27. Badimon L, Chiva-Blanch G, Lipid metabolism in dyslipidemia and familial hypercholesterolemia. In: *The Molecular Nutrition of Fats*, 2019; 307-322. <https://doi.org/10.1016/B978-0-12-811297-7.00024-X>
28. Num-Adom SM, Ogbuagu NE, Aluwong T, Adamu S, Umar IA, Esievo KAN, Dyslipidaemia of alloxan-induced type 1 diabetes mellitus in dogs is attenuated by ethanolic extract of *Anoigessus leiocarpus* stem bark. *Diabetes and Obesity International Journal*, 2022; 7(3): 1-9. <https://doi.org/10.23880/doi-16000260>
29. Esievo KAN, *Veterinary Clinical Pathology*, 1st ed. Spectrum Books, Ibadan; United Kindom Limited, Jersey; Safari Books (Exprt). 2017; Pp 171.
30. Shaish A, Horari Y, Cohen H, Schonfeld G, Harats D, Application of Dunalella in atherosclerosis 20. *The Alga Dunaliella*, 2019. <https://doi.org/10.1201/9780429061639-20>
31. Di-Ciaula A, Garruti G, Lunardi Baccetto R, Molina-Molina E, Bonfrate L, Wang DQ, portincasa P, Bile acid physiology. *Annals of Hepatology*, 2017; pp. 4-14. <https://doi.org/10.5604/01.3001.0010.5493> PMID:29080336
32. Dubois C, Armand M, Mekki N, Portugal H, Pauli AM, Bernard PM, Lafont H Lairon D, "Effects of increasing amounts of dietary cholesterol on postprandial lipemia and lipoproteins in human subjects". *Journal of Lipid Research*, 1994; pp. 35. [https://doi.org/10.1016/S0022-2275\(20\)39946-6](https://doi.org/10.1016/S0022-2275(20)39946-6) PMID:7868978
33. Cohn JS, Kamili A, Wat E, Chung RW, Tandy S, "Dietary phospholipids and intestinal cholesterol absorption". *Nutrients*, 2010; 2 (2): 116-27. <https://doi.org/10.3390/nu2020116> PMID:22254012 PMCid:PMC3257636
34. Behrman EJ, Gopalan V., Scovell WM, "Cholesterol and Plants". *Journal of Chemical Education*, 2005; 82 (12): 1791. <https://doi.org/10.1021/ed082p1791>
35. Jones PJ Dietary cholesterol and the risk of cardiovascular disease in patients: A review of the Harvard Egg Study and other data. *International Journal of Clinical Practice*, 2009; 63: 1-8. <https://doi.org/10.1111/j.1742-1241.2009.02136.x> PMID:19751443
36. Huff MW, Pollex RL, Hegele RA, NPC1L1: evolution from pharmacological target to physiological sterol transporter. *Arteriosclerosis Thrombosis Vascular Biology*, 2006; 26: 2433-2438. <https://doi.org/10.1161/01.ATV.0000245791.53245.ee> PMID:16973966
37. Levy E, Spahis S, Sinnott D, Peretti N, Maupas-Schwalm F, Delvin E, Lambert M, Lavoie MA, Intestinal cholesterol transport proteins: an update and beyond. *Current Opinion in Lipidology*, 2007; 18: 310-318. <https://doi.org/10.1097/MOL.0b013e32813fa2e2> PMID:17495606
38. Wang DQ Regulation of intestinal cholesterol absorption. *Annual Review of Physiology*, 2007; 69: 221-248. <https://doi.org/10.1146/annurev.physiol.69.031905.160725> PMID:17002594
39. Attard R, Dingli P, Doggen CJM, Cassar K, Farrugia R, Wettinger SB, The impact of passive smoking o inflammation, lipid profile and the risk of myocardial infarction. *Open Heart*, 2017; 4: 1-8. <https://doi.org/10.1136/openhrt-2017-000620> PMID:28878948 PMCid:PMC5574419
40. Hu T, Jialal I, *Physiology, Cholesterol* In: Treasure Island (FL). StatPearls Publishing, 2019.
41. Anichkov NN, Experimental arteriosclerosis in animals. In: Cowdry EV, editor, *Arteriosclerosis: A survey of the problem*. New York: MacMillan Publishing, 1933; pp. 271-322.
42. Anichkov NN A history of experimentation on arterial arteriosclerosis in animals. In: Blumenthal HT, editor, *Cowdry's arteriosclerosis: a survey of the problem*. 2nd ed. Springfield, (IL): Charles C. Thomas Publishing; 1967; pp. 21-46.

43. Kuijpers PMJC, History in medicine: the story of cholesterol, lipids and cardiology. *e-Journal of Cardiology Practice*, 2021; 19(9).
44. Ravnskov U, Is atherosclerosis caused by high cholesterol? *Open Journal of Medicine*, 2002; 95: 397-403. <https://doi.org/10.1093/qjmed/95.6.397> PMID:12037248
45. Anderson KM, Castelli WP, Levy D, Cholesterol and mortality. 30 years of follow-up from the framingham study. *Journal of the American Medical Association*, 1987; 257: 2176-2180. <https://doi.org/10.1001/jama.1987.03390160062027> PMID:3560398
46. LaRosa JC, Hunnigake D, Bush D, The cholesterol facts. A summary of the evidence relating dietary fats, serum cholesterol, and coronary heart disease. A joint statement by the American heart association and the national heart, lung, and blood institute. *Circulation*, 1990; 81: 1721-1733. <https://doi.org/10.1161/01.CIR.81.5.1721> PMID:2184951
47. Vega GL, Grundy SM, Current trends in non-HDL cholesterol and LDL cholesterol levels in adults with atherosclerotic cardiovascular disease. *Journal of Clinical Lipidol*, 2019; 13: 563-567. <https://doi.org/10.1016/j.jacl.2019.05.012> PMID:31248654
48. Landé KE, Sperry WM, Human atherosclerosis in relation to the cholesterol content of the blood serum. *Archives of Pathology*, 1936; 22: 301-312.
49. Hecht HS, Superko HR, Electron beam tomography and national cholesterol education program guidelines in asymptomatic women. *Journal of America College of Cardiology*, 2001; 37: 1506-1511. [https://doi.org/10.1016/S0735-1097\(01\)01211-6](https://doi.org/10.1016/S0735-1097(01)01211-6) PMID:11345357
50. Ulmer H, Kelleher C, Diem G, Concin H, "Why Eve is not Adam: prospective follow-up in 149650 women and men of cholesterol and other risk factors related to cardiovascular and all-cause mortality". *Journal of Women's Health (Larchmt)*, 2004; 13(1): 41-53. <https://doi.org/10.1089/154099904322836447> PMID:15006277
51. Sachdeva A, Cannon CP, Deedwania PC, Lipid levels in patients hospitalized with coronary artery disease: an analysis of 136,905 hospitalizations in get with the guidelines. *American Heart Journal*, 2009; 157:111-117. <https://doi.org/10.1016/j.ahj.2008.08.010> PMID:19081406
52. Al-Mallah MH, Hatahet H, Cavalcante JL, Low admission LDL-cholesterol is associated with increased 3-year all-cause mortality in patients with non-ST segment elevation myocardial infarction. *Cardiology Journal*, 2009; 16: 227-233.
53. Ravnskov U, McCully KS, Infections may be causal in the pathogenesis of atherosclerosis. *American Journal of Medical Sciences*, 2012; 344: 391-394. <https://doi.org/10.1097/MAJ.0b013e31824ba6e0> PMID:22495581
54. Lewington S, Blood cholesterol and vascular mortality by age, sex, and blood pressure: a meta-analysis of individual data from 61 prospective studies with 55 000 vascular deaths. *Lancet*, 2007; 370(9602): 0-1839. [https://doi.org/10.1016/S0140-6736\(07\)61778-4](https://doi.org/10.1016/S0140-6736(07)61778-4) PMID:18061058
55. Rong Y, Chen L, Zhu T, Egg consumption and risk of coronary heart disease and stroke: dose-response meta-analysis of prospective cohort studies. *British Medical Journal*, 2013; 346, e8539. <https://doi.org/10.1136/bmj.e8539> PMID:23295181 PMID:PMC3538567
56. Shin JY, Xun P, Nakamura Y, Egg consumption in relation to risk of cardiovascular disease and diabetes: a systematic review and meta-analysis. *American Journal of Clinical Nutrition*, 2013; 98: 146-159. <https://doi.org/10.3945/ajcn.112.051318> PMID:23676423 PMID:PMC3683816
57. Zheng Y, Li Y, Rimm EB, Hu FB, Albert CM, Rexrode KM, Manson JE, Qi L Dietary phosphatidylcholine and risk of all-cause and cardiovascular-specific mortality among us women and men. *American Journal of Clinical Nutrition*, 2016; 104, 173-180. <https://doi.org/10.3945/ajcn.116.131771> PMID:27281307 PMID:PMC4919531
58. Pihlajamäki J, Gylling H, Miettinen TA, Laakso M, Insulin resistance is associated with increased cholesterol synthesis and decreased cholesterol absorption in normoglycemic men. *Journal of Lipid Research*, 2004; 45: 507-512. <https://doi.org/10.1194/jlr.M300368-JLR200> PMID:14657199
59. Tang WH, Wang, Z, Levison BS, Koeth RA, Britt EB, Wu, Y, Hazen SL, Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *New England Journal of Medicine*, 2013; 368: 1575-1584. <https://doi.org/10.1056/NEJMoa1109400> PMID:23614584 PMID:PMC3701945
60. Berger S, Raman G, Vishwanathan R, Jacques PF, Johnson EJ Dietary cholesterol and cardiovascular disease: a systematic review and meta-analysis. *American Journal of Clinical Nutrition*, 2015; 102(2): 276-294. <https://doi.org/10.3945/ajcn.114.100305> PMID:26109578
61. Fernández-Friera LFV, López-Melgar B, Normal LDL-cholesterol levels are associated with subclinical atherosclerosis in the absence of risk factors. *JACC*. 2017; 70: 2979-2991. <https://doi.org/10.1016/j.jacc.2017.10.024> PMID:29241485
62. Dimsdale JE, Herd A, Variability of plasma lipids in response to emotional arousal. *Psychosomology Medicine*, 1982; 44:413-430. <https://doi.org/10.1097/00006842-198211000-00004> PMID:6758015
63. Rosenman RH, Relationships of neurogenic and psychological factors to the regulation and variability of serum lipids. *Stress Medicine*, 1993; 9: 133-140. <https://doi.org/10.1002/smi.2460090302>
64. Ohmura H, Fukushima Y, Mizuno A, Niwa K, Kobayashi Y, Ebina T, Kimura K, Ishibashi S, Estimated Prevalence of Heterozygous Familial Hypercholesterolemia in Patients with Acute Coronary Syndrome. *International Heart Journal*, 2017; 58(1): 88-94. <https://doi.org/10.1536/ihj.16-188> PMID:28123161
65. Zhong VW, Horn LV, Cornelis MC, Wilkins JT, Ning H, Carnethon MR, Greenland P, Mentz RJ, Tucker KL, Zhao L, Norwood AF, Lloyd Jones DM, Allen NB, Associations of dietary cholesterol or egg consumption with incident cardiovascular disease and mortality. *Journal of the American Medical Association*, 2019; 321(11): 1081-1095. <https://doi.org/10.1001/jama.2019.1572> PMID:30874756 PMID:PMC6439941
66. Faselis C, Imprialos K, Grassos H, Pittaras A, Kallistratos MM, AthanasiosIs very low LDL-C harmful? *Current Pharmaceutical Design*, 2018; 24(7): 3658-3664. <https://doi.org/10.2174/1381612824666181008110643> PMID:30295187
67. Collins R, Reith C, Emberson J, Interpretation of the evidence for the efficacy and safety of statin therapy. *Lancet*, 2016; 388: 2532-2561. [https://doi.org/10.1016/S0140-6736\(16\)31357-5](https://doi.org/10.1016/S0140-6736(16)31357-5) PMID:27616593
68. Ravnskov UD, Hama M, DHamazaki T, Hammarskjöld B, Hynes N, Kendrick M, Langsjoen PH, Malhotra A, Mascitelli L, McCully KS, Ogushi Y, Okuyama H, Rosch Paul J, Schersten T, Sultan S, Sundberg R, "Lack of an association or an inverse association between low-density-lipoprotein cholesterol and mortality in the elderly: a systematic review". *British Medical Journal Open*, 2016; 6(6): e010401. <https://doi.org/10.1136/bmjopen-2015-010401> PMID:27292972 PMID:PMC4908872
69. Bathum L, Depont CR, Engers PL, Association of lipoprotein levels with mortality in subjects aged 50+ without previous diabetes or cardiovascular disease: a population-based register study. *Scandalevia Journal of Primary Health Care*, 2013; 31:172-180. <https://doi.org/10.3109/02813432.2013.824157> PMID:23941088 PMID:PMC3750440
70. Hamazaki T, Okuyama TH, Ogushi NY, Hama KR, Towards a Paradigm Shift in Cholesterol Treatment: A Re-Examination of the Cholesterol Issue in Japan. 2015; 66(4). <https://doi.org/10.1159/000381654>
71. Benn M, Low LDL cholesterol, PCSK9 and HMGR genetic variation, and risk of Alzheimer's disease and Parkinson's disease: Mendelianrandomisation study. *BioMedical Journal*, 2017; 357: 1648. <https://doi.org/10.1136/bmj.j1648> PMID:28438747 PMID:PMC5421439

72. Cappell DF, Anderson JR, Muir's Text book of Pathology. 9th edn. London, Edward Arnold, 1924.
73. Adams DD, Seven deficiencies in Harrison's 16th edn. *Journal of Clinical and Laboratory Immunology*, 2005; 54:1-13.
74. Adams DD *Quarterly Journal of Medicine*, 2011; 104: 867-870. <https://doi.org/10.1093/qjmed/hcr087> PMID:21690178
75. Hasson GK, Immune mechanisms in atherosclerosis. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 2001; 21: 1876-1890. <https://doi.org/10.1161/hq1201.100220> PMID:11742859
76. Willerson JT, Ridker PM, Inflammation as a cardiovascular risk factor. *Circulation*, 2004; 109: II2-II10. <https://doi.org/10.1161/01.CIR.0000129535.04194.38>
77. Paoletti R, Gotto AM, Hajjar DP, Inflammation in atherosclerosis and implication for therapy. *Circulation*, 2004; 109: III20-III26. <https://doi.org/10.1161/01.CIR.0000131514.71167.2e>
78. Libby P, Theroux P, Pathophysiology of coronary heart disease. *Circulation*, 2005; 111: 3481-3488. <https://doi.org/10.1161/CIRCULATIONAHA.105.537878> PMID:15983262
79. Jacobs D, Blackburn H, Haggins M, Report of the conference on low blood cholesterol: Mortality association. *Circulation Journal*, 1992; 86: 1046-1060. <https://doi.org/10.1161/01.CIR.86.3.1046> PMID:1355411
80. Ravnskov U, McCully KS, Vulnerable plaque formation from obstruction of vasa vasorum by homocysteinylated and oxidized auto-antibodies. *Annals of clinical laboratory science*, 2009; 39: 3-16.
81. Read SA, Douglas MW, Virus induced inflammation and cancer development. *Cancer Letters*, 2014; 345: 174-181. <https://doi.org/10.1016/j.canlet.2013.07.030> PMID:23941825
82. Newman TB, Hulley SB, Carcinogenicity of Lipid-Lowering Drugs-Reply. *Journal of the American Medical Association*, 1996; 275(19):1481-1482. <https://doi.org/10.1001/jama.275.19.1481> PMID:8622220
83. National Cholesterol Education Program (NCEP), Executive summary of the third report of the Expert Panel on detection, evaluation and treatment of high blood cholesterol in adults (Adult Treatment Panel III). *Journal of the American Medical Association*, 2001; 285: 2486-2497. <https://doi.org/10.1001/jama.285.19.2486> PMID:11368702
84. Li LH, Dutkiewicz EP, Huang YC, Zhou HB, Hsu CC, Analytical methods for cholesterol quantification. *Journal of Food and Drug Analysis*, 2019; 27(2): 375-386 <https://doi.org/10.1016/j.jfda.2018.09.001> PMID:30987710 PMID:PMC9296199
85. Sung SO, Bahamans SCH, Andreade L, Stuart MFF, Brian C, Frequency-domain differential photo acoustic radar: theory and validation from ultrasensitive atherosclerotic plaque imaging. *Journal of Biomedical Optics*, 2019; 24(6): 66003. <https://doi.org/10.1117/1.JBO.24.6.066003> PMID:PMC6977017
86. Abell LL, Levy BB, Brodie BB, Kendall FE, A simplified method for the estimation of total cholesterol in serum and demonstration of its specificity. *Journal of Biological Chemistry*, 1952; 195: 357-366. [https://doi.org/10.1016/S0021-9258\(19\)50907-3](https://doi.org/10.1016/S0021-9258(19)50907-3)
87. dos Santos Ferreira CE, França CN, Correr CJ, Zucker ML, Andriolo A, Scartezini M, Clinical correlation between a point-of-care testing system and laboratory automation for lipid profile. *international journal of clinical chemistry*, 2015; 446: 263-266. <https://doi.org/10.1016/j.cca.2015.04.036> PMID:25952166
88. Lian K, Zhang P, Wang W, Dai T, Li E, Determination of total cholesterol in serum by gas chromatography-mass spectrometry. *Asian Journal of Chemistry*, 2014; 26: 2646-2648. <https://doi.org/10.14233/ajchem.2014.15780>
89. Beggio M, Cruz Hernandez C, Golay P A, Lee LY, Giuffrida F, Quantification of total cholesterol in human milk by gas chromatography. *Journal of Separation Science*, 2018; 41: 1805-1811. <https://doi.org/10.1002/jssc.201700833> PMID:29327426
90. Liebis G, Binder M, Schifferer R, Langmann T, Schulz B, Schmitz G, High throughput quantification of cholesterol and cholesteryl ester by electrospray ionization tandem mass spectrometry (ESI-MS/MS). *Biochim Biophys Acta Mol Cell Biol Lipids*, 2006; 1761: 121-128. <https://doi.org/10.1016/j.bbalip.2005.12.007> PMID:16458590
91. Kock R, Delvoux B, Greiling H, Determination of total cholesterol in serum by liquid chromatography-isotope dilution mass spectrometry. *Clinical Chemistry*, 1997; 43: 1896-1903. <https://doi.org/10.1093/clinchem/43.10.1896>
92. Hidaka H, Hanyu N, Sugano M, Kawasaki K, Yamauchi K, Katsuyama T, Analysis of human serum lipoprotein lipid composition using MALDI-TOF mass spectrometry. *Annals of Clinical & Laboratory Science*, 2007; 37: 213-221.
93. Xu L, Kliman M, Forsythe JG, Korade Z, Hmelo AB, Porter NA, Profiling and imaging ion mobility-mass spectrometry analysis of cholesterol and 7-dehydrocholesterol in cells via sputtered silver MALDI. *Journal of the American Society for Mass Spectrometry*, 2015 26: 924-933. <https://doi.org/10.1007/s13361-015-1131-0> PMID:25822928 PMID:PMC4608680
94. Wu C, Ifa DR, Manicke NE, Cooks RG, Rapid, direct analysis of cholesterol by charge labeling in reactive desorption electrospray ionization. *Analytical Chemistry*, 2009; 81: 7618-7624. <https://doi.org/10.1021/ac901003u> PMID:19746995 PMID:PMC2747033
95. Al-Balaa D, Rajchl A, Grégrová A, Ševčík R, Čížková H, DART mass spectrometry for rapid screening and quantitative determination of cholesterol in egg pasta. *Journal of Mass Spectrometry*, 2014; 49: 911-917. <https://doi.org/10.1002/jms.3465> PMID:25230188
96. Hsieh H-Y, Li L.-H, Hsu R-Y, Kao W-F, Huang Y-C, Hsu C-C, Quantification of endogenous cholesterol in human serum on paper using direct analysis in real time mass spectrometry. *Analytical Chemistry*, 2017; 89: 6146-6152. <https://doi.org/10.1021/acs.analchem.7b00943> PMID:28505411
97. Enthaler B, Pruns JK, Wessel S, Rapp C, Fischer M, Wittern K-P, Improved sample preparation for MALDI-MSI of endogenous compounds in skin tissue sections and mapping of exogenous active compounds subsequent to ex-vivo skin penetration. *Analytical and Bioanalytical Chemistry*, 2012; 402: 1159-1167. <https://doi.org/10.1007/s00216-011-5562-6> PMID:22139470