

Available online on 15.01.2023 at <http://jddtonline.info>

# Journal of Drug Delivery and Therapeutics

Open Access to Pharmaceutical and Medical Research

Copyright © 2023 The Author(s): This is an open-access article distributed under the terms of the CC BY-NC 4.0 which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited



Open Access Full Text Article



Research Article

## GCMS and FTIR analysis of ethanol and methanol leave extract of *Urena lobata* (Caesar weed) for bioactive phytochemical constituents

Collins Obinna Keke<sup>1</sup>\*, Winifred Njideka Nsofor<sup>1</sup>, Francis Koku Ras Kumabia<sup>2</sup>, Godian Chima Iloabuchi<sup>1</sup>, Juliet Chioma Ejiofor<sup>1</sup>, Olachi Lilian Osuagwu<sup>1</sup>

<sup>1</sup> Department of Biochemistry, School of Biological Sciences, Federal University of Technology, Owerri, Imo State, Nigeria

<sup>2</sup> Department of Science Education, St. Francis College of Education, Hohoe, Ghana

### Article Info:

### Abstract



#### Article History:

Received 17 Nov 2022  
Reviewed 23 Dec 2022  
Accepted 04 Jan 2023  
Published 15 Jan 2023

#### Cite this article as:

Keke CO, Nsofor WN, Kumabia FKR, Iloabuchi GC, Ejiofor JC, Osuagwu OL, GCMS and FTIR analysis of ethanol and methanol leave extract of *Urena lobata* (Caesar weed) for bioactive phytochemical constituents, Journal of Drug Delivery and Therapeutics. 2023; 13(1):99-115

DOI: <http://dx.doi.org/10.22270/jddt.v13i1.5722>

#### \*Address for Correspondence:

Collins Obinna Keke, Department of Biochemistry, School of Biological Sciences, Federal University of Technology, Owerri, Imo State, Nigeria

This study was designed to apply the highly sophisticated biological and chemical characterization techniques-(GC-MS) Gas chromatography-mass spectrometry and FTIR spectroscopy to screen for bioactive phytochemicals present in ethanol and methanol extracts of *U. lobata*. The leaves of the plant were collected from a farm in Umuode village in Osisioma Ngwa local government area of Abia state, Nigeria. Ethanol and methanol extracts of the leaves were prepared and analyzed using Buck M910 Gas chromatography system with HP-5MS column (30m in length x250µm in diameter x 0.25µm in thickness film). Analysis of functional groups in the phytochemicals were done using Buck scientific M530 USA FTIR.12 vibrational functional groups were revealed by the FTIR screening in the ethanol and methanol extracts of *U. lobata* leaves. Some of the functional groups were indicative of alcohols, phenols, aromatic compounds, unsaturated hydrocarbons, vinyl ethers, amines, isonitriles and aliphatic compounds. GC-MS analysis of the ethanol leaf extract revealed 41 bioactive compounds with the following being most abundant, 9-octadecenoic acid(16.8%), dodecanoic acid(13.43%),n-hexadecanoic acid(11.73%), octadecanoic acid (9.78%), 1-docosene(9.57%) while the methanol extract revealed 47 bioactive compounds with the following having the highest abundance, n-hexadecanoic acid (26.65%) and (9.11%), dodecanoic acid(6.89%), 1-docosene(6.06%), erucic acid(4.09%).These phytochemicals and many others present in the leaf have been reported to possess multiple therapeutic activities. This therefore explains the use of this leaf in ancient medicine to treat numerous disease conditions. It further implies that the leaf could be exploited for the formulation of therapeutic molecules.

**Keywords:** *Urena lobata*, GC-MS, FTIR, Spectroscopy, Phytochemicals, Functional group, Phytoconstituents

## INTRODUCTION

Plants, both in their natural state and as extracts, have provided significant benefits to human health for centuries. The pharmacological benefits of these plants come from the wide variety of secondary and primary metabolites they contain, each of which has its own complicated chemical makeup and is responsible for their pharmacological effects<sup>1</sup>. For their low or nonexistent price tags and lack of negative side effects, natural remedies routinely top the list of those recommended over today's synthetic pharmaceuticals. The chemical make-up of these plants is useful not only in traditional therapy but also in the characterization and finding of novel sources of phyto-components for pharmacological research and development<sup>2, 3</sup>. Recently, the use of Gas chromatography-mass spectrometry and FTIR spectroscopy for target identification of functional groups and determination of these different phyto components that are present in medicinal plants even in small concentrations has expanded substantially<sup>1</sup>. GC/MS remains superior among effective, rapid, and precise methods for identifying plant-based substances such as amino acids, alkaloids, esters, long-chain hydrocarbons, steroids, organic acids, nitro compounds and alcohols<sup>4,3</sup>.

The sub-shrub *Urena lobata* is a member of the Malvaceae family. They attain heights of 0.6-3m and diameters of 7cm at the base<sup>5</sup>. The plant is mostly found in the tropics and subtropics of South and North America, Philippines, Asia, Indonesia as well as in several African nations like Nigeria, Senegal, Ghana and the Democratic Republic of the Congo<sup>6</sup>. In Nigeria, *U. lobata* is also popular and known with different local names among different ethnic groups. The Igbos of Eastern Nigeria particularly those in Imo state call it Achara /Udo<sup>7</sup>the Hausas call it Rama-rama, in Efik, it is known as Rideri, the Binis and some other parts of Edo state call it Oronhon while the Yorubas of Western Nigeria call the plant different names such as; Ilasa-omode, Akeri, Ilasa-agbunrin<sup>8</sup>. *U. lobata* is employed in folkloric medicine in the treatment of dysentery, hematemesis, edema, carbuncle, bleeding due to trauma, leucorrhea, gonorrhea, fever, pain, cold, numbness resulting from rheumatism and other disease conditions<sup>9</sup>. The inhabitants of Nigeria's Katsina State utilize the leaves and petals of this plant to make local delicacies and also offer them to lactating women to increase their milk supply<sup>10</sup>. Numerous studies have reported significant pharmacological activities of various extracts of this plant parts to include anti-inflammatory<sup>11</sup>, anti-diabetic<sup>12</sup>, anti-tumor<sup>13</sup>, antimicrobial<sup>14</sup>, anti-oxidant and cytotoxic<sup>15</sup>, wound healing effect<sup>16</sup>, anti-

diarrheal<sup>11</sup>, neuro-pharmacological activities<sup>17</sup>, anti-fertility in female rats as a result of reduced myometrial and epimetrial thickness accompanied by reduced uterine diameter<sup>18</sup> as well as reversibly inhibiting spermatogenesis and steroidogenesis serving as male contraceptives<sup>19</sup>.

In order to better understand the therapeutic effects of *Urena lobata*, this study used GC/MS and FTIR to identify the many bioactive phytoconstituents and functional groups present in ethanol and methanol extracts of the plant.

## MATERIALS AND METHODS

### Collection and processing of plant materials

Healthy and mature leaves of the plant were freshly picked from a farm in Umuode village in Osisioma Ngwa L.G.A of Abia state, Nigeria and identified as *Urena lobata*.

### Extraction of plant material

The picked leaves underwent a thorough cleaning with distilled water before being air dried for two weeks. Using an electric blender (ES-BL-090/350W/220-240V/50Hz/China), the dry leave materials were crushed into a fine powder. Soxhlet extraction was performed on the dried and powdered materials using ethanol and methanol as extraction solvents. A rotary evaporator (Heidolph Rotavapor, Germany) was used to filter and concentrate the extracts. When the dried extracts were ready for analysis, they were put into airtight containers, corked, and kept in a refrigerator at 4°C.

### Gas Chromatography –Mass Spectrometry (GC-MS) Analysis

Using a BUCK M910 Gas chromatograph with an HP-5MS column (30 m in length, 250µm in diameter, and 0.25µm in film thickness), we were able to conduct the GC-MS analysis of the bioactive components. High-energy electrons were used in a system of electron ionization for spectroscopic detection by GC-MS (70 eV). Helium gas, purified to 99.995% purity, was employed as the carrier gas at a flow rate of 1 mL/min. We started with a temperature of 50°C, ramped it up by 3°C every minute, and held it there for around 10 minutes.

At last, a 10°C/min increase in temperature brought the total to 300 °C. In splitless mode, 1µm of the produced 1% extracts diluted with acetonitrile was injected. Based on the chromatographic peak area, the relative amount of bioactive chemicals in each extract was calculated and expressed as a percentage. Based on GC retention time on HP-5MS column and spectrum matching with software data of standards, bioactive chemicals were identified from various extracts (Replib and Mainlab data of GC-MS systems). The name, molecular weight and structure of the compounds of the test extracts were determined

### Fourier Transform Infrared Spectroscopic (FT-IR) Analysis

A Buck Scientific M530 USA FTIR was used for the analysis. A deuterated triglycine sulfate detector and a potassium bromide beam splitter were used in this device. The spectra were acquired and adjusted with the help of the Gram A1's software. About 1.0g of samples and 0.5ml of nujol were used; they were carefully mixed and then placed on the salt pellet. FTIR spectra were acquired during the measurement in frequency ranges of 4,000 - 600 cm<sup>-1</sup> and co-added at 32 scans and 4 cm<sup>-1</sup> resolution. Transmitter values were shown as FTIR spectra.<sup>20</sup>.

## RESULTS AND DISCUSSION

Among the 41 phytocompounds identified by GC-MS in the ethanol leave extract of *U. lobata* (Table 1), the following were

found to be most predominant: 9-Octadecenoic acid, (E)- (16.80%), Dodecanoic acid (13.43%), n-Hexadecanoic acid (11.73%), Octadecanoic acid (9.78%), 1-Docosene (9.57%), Cyclohexadecane, 1,2-diethyl- (4.85% and 2.03%), Oleic acid (4.74%), Phenol,3,5-bis(1,1-dimethylethyl) (2.42%), Tetradecanoic acid (2.14%). Conversely, the least abundant bioactive phytocompounds in the extract were Nonadecyltrifluoroacetate (0.12%), 6,11-Dimethyl-2, 6,10-dodecatrien-1-ol (0.16%), Decane (0.19%), Dodecyl acrylate (0.22%), 1-Eicosanol (0.22%) and Methyl stearate (0.24%). In a similar study, <sup>76</sup>Fadillah and his group from Jawa Barat, Indonesia investigated the active compounds in both 50% and 70% ethanol extracts of *U. lobata*. Their findings revealed 5 and 17 bioactive compounds in 50% and 70% ethanol extract of *U. lobata* leaves respectively. They went further to report high concentration of n-Hexadecanoic acid (19.97%) as well as Oleic acid (9.42%) in 50% ethanol leave extract of *U. lobata*. Again, the most prevalent bioactive constituent in the 70% ethanol leave extract of *U. lobata* was discovered to be n-Hexadecanoic acid (18.91%). These findings are comparable with ours, particularly in methanol extract which had, n-Hexadecanoic acid (26.65%) as the most abundant bioactive compound, nevertheless, their study revealed the presence of other bioactive phytocompounds which were not identified in ours and vice versa. As demonstrated in table 5, ethanol leave extract of *U. lobata* has been shown to contain a number of biologically active phytocompounds with diverse biological and pharmacological effects. 9-Octadecenoic acid has been reported to exhibit antimicrobial activities<sup>31</sup>.

Dodecanoic acid (also known as lauric acid), a medium-chain as well as saturated fatty acid found abundantly in oils of coconut and palm kernel is known to exhibit antibacterial and antioxidant activities<sup>32</sup>. Among many other bioactivities, n-hexadecanoic acid (also known as palmitic acid) has inflammatory, antioxidant, hypocholesteromic, and cancer-preventive properties<sup>38</sup>. Some common and natural sources of n-Hexadecanoic are palm oil, butter, palm kernel oil, milk, meat and cheese. 1-Docosene has antibacterial activities<sup>43</sup>. Albrathy also reported the antimicrobial activities of Oleic acid, a common monounsaturated fatty acid seen naturally in numerous animal and vegetable fats and oil and in plants such as *Gladiolus italicus* and *Prune mume*<sup>43</sup>.

In addition to the above mentioned biological roles, oleic acid has been shown to have antifungal, anti-inflammatory, antioxidant, and antibacterial properties<sup>41</sup>. The phenolic compound, Phenol,3,5-bis(1,1-dimethylethyl) possesses antibacterial and antimicrobial activities<sup>30,29</sup>. Least abundant bioactive compounds such as; 1-Eicosanol, methyl stearate, Dodecyl acrylate and 6,11-Dimethyl-2,6,10 dodecatrien-1-ol are not exempted in exhibiting different biological activities as many researchers have also demonstrated their biological activities. For example, 1-Eicosanol, an alcohol has been reported to possess antimalarial, antifungal, and antioxidant activities<sup>31</sup> while Methyl stearate has proven to be a potent anti-inflammatory, intestinal lipid metabolic regulator, GABA aminotransferase inhibitor, antinociceptive, anthelmintic, and a potent gastric inhibitor<sup>42</sup>. On the other hand, Dodecyl acrylate, a typical acrylate ester is known to be antioxidant and antimicrobial<sup>36</sup> while 6,11-Dimethyl-2,6,10 dodecatrien-1-ol has been researched to possess antimicrobial activities<sup>46</sup>.

The most bioactive phytocompounds (47) were found in the methanol extract of *U. lobata*. This was illustrated by the GC-MS chromatogram of the methanol extract, which is shown in Figure 2. The following were the prevailing bioactive phytocompounds identified; n-Hexadecanoic acid (26.65% and 9.11%), Dodecanoic acid (6.89%), 1-Docosene (6.06%), Erucic acid (4.09%), 1-Octadecene (3.54%), 9-Octadecenoic acid (3.45%), Tetradecanoic acid (3.01%) and

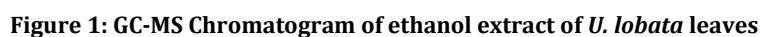
Diisooctylphthalate (2.97%). The least abundant compounds were; 3-Eicosene (2.97%), 6-Octadecanoic acid methyl ester (0.14%), Dodecane (0.15%), 1-Docosene (0.17%), Trifluoroacetic acid pentadecyl ester (0.18%), Cyclohexane, 1,1'-(1,4-butanediyl)bis (0.22%), Tridecanoic acid, 12-methyl-methyl ester (0.22%) and Cyclotetradecane (0.23%). These bioactive compounds and many more in methanol extract of *U. lobata* have been reported to possess many biological activities just like those identified in ethanol extract of the leaf. One of them is Erucic acid; an omega-9 and monosaturated fatty acid that is common among brassicaceae family of flowering plants has been reported to regulate mesenchymal stem cell differentiation into osteoblasts and adipocyte<sup>72</sup>. Another study reported it to have a broad-spectrum antiviral activity against Influenza A. Virus (IAV), anti-inflammatory and pro-inflammatory amplification effect as well as the ability to inhibit NF- $\kappa$ B and p38 MAPK<sup>73</sup>. 1-Octadecene, a long-chain hydrocarbon and an alkene which has been found in appreciable quantity in *Vaccinium macrocarpon*, a North American variant of cranberry known to be used by women in the treatment of recurrent urinary tract infection has also shown excellent antibacterial, antioxidant and anticancer activities<sup>39,59,60</sup>. 9-Octadecenoic acid has been reported to possess antimicrobial activities<sup>31</sup>. An ample amount of 9-Octadecenoic acid is found naturally in *Eleutherococcus sessiliflorus* and *Dipteryx lacunifera*, an oleaginous leguminous plant native to Piauí and Maranhão state in North East of Brazil<sup>77</sup>. Diisooctyl phthalate has antimicrobial, antifouling<sup>74</sup> and antibacterial activities<sup>75</sup>. 3-Eicosene has also shown to be antimicrobial, antihyperglycemic, cytotoxic, antioxidant and insecticidal<sup>35</sup>. Another fatty acid methyl ester (FAME) that has been shown to have important biological activities is 6-octadecanoic acid methyl ester. It has been researched to possess strong analgesic, anti-inflammatory and antipyretic activities<sup>68</sup>. Pantadecanoic acid, 9-Hexadecanoic acid and Heptadecanoic acid are among the fatty acids uniquely found in methanol extract of *U. lobata* leaves with tremendous biological significance. Pantadecanoic acid, an essential fatty acid found in butter whose biological activities promote long-term metabolic and cardiovascular health<sup>78</sup> has been shown to possess anticancer activity<sup>61</sup>. 9-Hexadecanoic acid from methanol extract of *Tribolium castaneum* also showed good antibacterial activity against *Escherichia coli*, a Gram-negative, enteropathogenic bacterium implicated in diarrheal disease and urinary tract infections<sup>65</sup>. Heptadecanoic acid has antioxidant properties<sup>66</sup>.






Tables 3 and 4 show the findings of the FTIR investigation of the ethanol and methanol leaf extracts of *U. lobata*. FTIR, a vibration spectroscopic technique<sup>79</sup> is renowned for its ability to pin-point important functional groups embedded in plant extracts, biological and synthetic compounds. For identifying types of bonds (functional groups) in compounds, FT-IR

remains unmatched<sup>80</sup>. In this study, the FTIR spectra of ethanol and methanol extract shown in figure 3 and 4, unraveled twelve peaks indicative of 12 functional groups. For the ethanol extract, the peaks are in the range of 3704.812, 3498.917, 3181.879, 3013.238, 2782.866, 2665.173, 2571.721, 222.731/2018, 1622.32, 1416.411, 1235.417 and 843.5038 cm<sup>-1</sup> whereas the peaks of the methanol extract are in the range of; 3664.219/3415.311, 3155.608, 3056.931, 2915.574, 2814.952, 2500.399, 2117.752, 1830.701, 1614.028, 1393.314, 1295.721 and 852.1639 cm<sup>-1</sup>. The 12 functional groups revealed in ethanol extract of the leave are: alcohols/phenols, aromatic amines, primary amides, aromatic/unsaturated hydrocarbons, ether/amine, aldehydes, phosphorus oxyacids, alkynes, primary amines and trisubstituted benzene. In the methanol extract, the functional groups revealed were: alcohols/phenols, amino acids, aromatic/saturated hydrocarbons, aliphatic compounds, hydrohalides, isonitriles,  $\beta$ -lactones, vinyl ethers, t-butyl groups, sulfones and primary amines. The following vibrational functional groups: alcohols/phenols, aromatic/unsaturated hydrocarbons, ether/amine are all common to both extracts and produce, OH stretch, =CH-H stretch and CH stretch respectively. With its hydrogen-bonding capabilities, the OH group is likely responsible for the inhibitory effect against pathogenic microbial agents observed in both methanol and ethanol extract of the leave<sup>80</sup>.

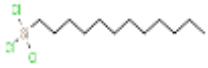

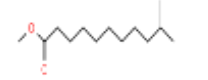
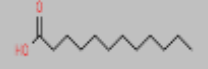







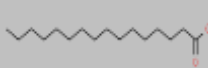


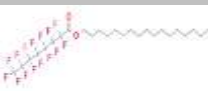






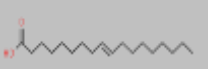

Figures 5 and 6 illustrate, respectively, the percentage abundance of the classes of bioactive chemicals found in the ethanol and methanol extracts. In the ethanol extract, fatty acids had a total abundance of 57.58%, alkene, 12.58%, cycloalkane, 11.62%, monounsaturated fatty acids (MUFA), 6.78% while alkane had a total abundance of 4.31%. The least abundant class of bioactive compounds was acrylate ester (0.22%) and alcohol (0.22%). For the methanol extract, as seen in ethanol extract, fatty acid has a total abundance of 56.15%, followed by alkene (20.01%), MUFA (10.02%), phthalate esters (2.97%) and delta-lactams (2.66%). The least abundant class of bioactive compounds were cycloalkane (0.8%) and fatty acid esters (0.18%).

Table 7 showed the bioactive compounds common to both ethanol and methanol extracts of *U. lobata*. The fatty acids were more prevalent in the methanol extract (26.65%) than in the ethanol (11.73%), including n-Hexadecanoic acid, 9-Octadecenoic acid, Dodecanoic acid, and Tetradecanoic acid. In ethanol extracts, 9-octadecenoic acid predominated (16.8%) compared to methanol extracts (3.45%). Dodecanoic acid was prevalent in ethanol extract (13.43%) compared to the methanol extract (6.89%). The varied concentrations of these bioactive compounds in the two extracts underscored the importance of choosing the right extraction solvents during research bearing in mind the bioactive compound of target.



PK	RT	Area %	Bioactive compound	Structure	MF	MW
1	6.959	0.19	Decane		C <sub>10</sub> H <sub>22</sub>	142.28
2	9.634	0.61	Dodecane		C <sub>12</sub> H <sub>26</sub>	170.33
3	14.674	0.39	Cyclopentane,1-hexyl-3-methyl-		C <sub>12</sub> H <sub>24</sub>	168.32
4	14.947	1.75	Tetradecane		C <sub>14</sub> H <sub>30</sub>	198.39
5	16.519	0.28	3-Heptafluorobuty Roxydodecane		C <sub>20</sub> H <sub>35</sub> F <sub>7</sub> O <sub>2</sub>	424.00



6	16.541	0.31	Silane, trichlorodocosyl-		<b>C<sub>12</sub>H<sub>25</sub>Cl<sub>3</sub>Si</b>	303.77
7	17.406	2.42	Phenol, 3,5-bis(1,1-dimethylethyl)		<b>C<sub>14</sub>H<sub>22</sub>O</b>	206.32
8	17.664	1.11	Undecanoic acid, 10-methyl-, methyl ester		<b>C<sub>13</sub>H<sub>26</sub>O<sub>2</sub></b>	214.34
9	19.124	13.43	Dodecanoic acid		<b>C<sub>12</sub>H<sub>24</sub>O<sub>2</sub></b>	200.32
10	19.563	0.39	9-Eicosene, (E)-		<b>C<sub>20</sub>H<sub>40</sub></b>	280.53
11	19.811	0.72	Hexadecane		<b>C<sub>16</sub>H<sub>34</sub></b>	226.44
12	21.542	0.22	Dodecyl acrylate		<b>C<sub>15</sub>H<sub>28</sub>O<sub>2</sub></b>	240.38
13	23.464	2.14	Tetradecanoic acid		<b>C<sub>14</sub>H<sub>28</sub>O<sub>2</sub></b>	228.37
14	24.003	0.38	9-Eicosene, (E)-		<b>C<sub>20</sub>H<sub>40</sub></b>	280.53
15	24.215	0.37	Octadecane		<b>C<sub>18</sub>H<sub>38</sub></b>	254.49
16	26.469	0.47	Pentadecanoic acid, 14-methyl-, methyl ester		<b>C<sub>17</sub>H<sub>34</sub>O<sub>2</sub></b>	270.45
17	27.601	11.73	n-Hexadecanoic acid		<b>C<sub>16</sub>H<sub>32</sub>O<sub>2</sub></b>	256.42
18	28.043	1.11	1-Octadecene		<b>C<sub>18</sub>H<sub>36</sub></b>	252.47
19	28.207	0.26	Methoxyacetic acid, 2-tetradecyl ester		<b>C<sub>17</sub>H<sub>34</sub>O<sub>3</sub></b>	286.45
20	28.669	0.43	Pentadecafluorooctanoic acid, octadecyl ester		<b>C<sub>26</sub>H<sub>37</sub>F<sub>15</sub>O<sub>2</sub></b>	666.5
21	28.796	2.02	9-Octadecenoic acid		<b>C<sub>18</sub>H<sub>34</sub>O<sub>2</sub></b>	282.46
22	29.042	0.40	Cyclohexane, butyl-		<b>C<sub>10</sub>H<sub>20</sub></b>	140.26
23	29.140	0.22	1-Eicosanol		<b>C<sub>20</sub>H<sub>42</sub>O</b>	298.54
24	29.253	0.52	cis-13-Octadecenoic acid, methyl ester		<b>C<sub>19</sub>H<sub>36</sub>O<sub>2</sub></b>	296.48
25	29.319	0.25	cis-Vaccenic acid		<b>C<sub>18</sub>H<sub>34</sub>O<sub>2</sub></b>	282.5
26	29.578	0.24	Methyl stearate		<b>C<sub>19</sub>H<sub>38</sub>O<sub>2</sub></b>	298.5
27	29.762	16.80	9-Octadecenoic acid		<b>C<sub>18</sub>H<sub>34</sub>O<sub>2</sub></b>	282.46
28	29.870	2.03	Cyclohexadecane, 1,2-diethyl-		<b>C<sub>20</sub>H<sub>40</sub></b>	280.53
























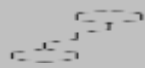




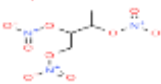
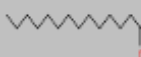



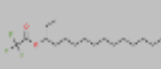

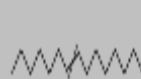

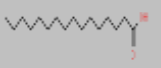





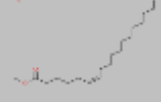



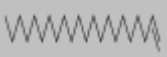







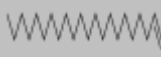



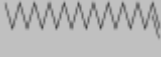
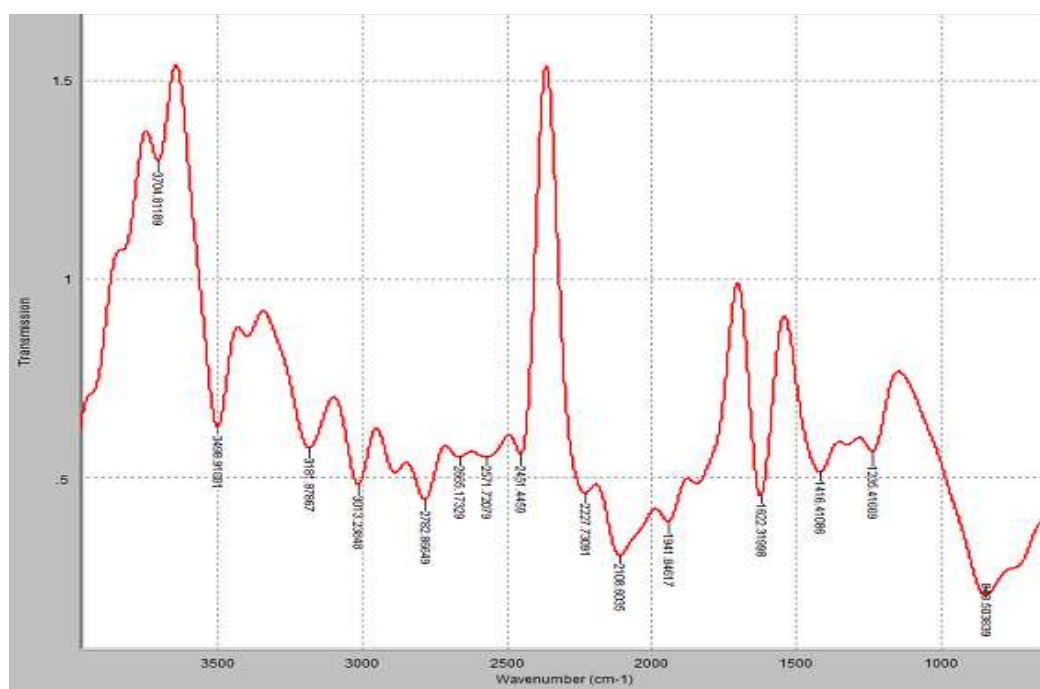
29	29.935	0.91	9-Octadecenoic acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.46
30	29.981	9.78	Octadecanoic acid		C <sub>18</sub> H <sub>36</sub> O <sub>2</sub>	284.47
31	30.094	4.74	Oleic Acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.5
32	30.159	1.79	Oleic Acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.5
33	30.243	9.57	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58
34	30.307	4.34	Cycloeicosane		C <sub>20</sub> H <sub>40</sub>	280.53
35	30.392	4.85	Cyclohexadecane, 1,2-diethyl-		C <sub>20</sub> H <sub>40</sub>	280.53
36	30.582	1.63	5-Eicosene, (E)-		C <sub>20</sub> H <sub>40</sub>	280.53
37	30.789	0.27	3-Eicosene, (E)-		C <sub>20</sub> H <sub>40</sub>	280.53
38	31.482	0.35	Heptadecylheptafluorobutyrate		C <sub>21</sub> H <sub>35</sub> F <sub>7</sub> O <sub>2</sub>	452.49
39	32.063	0.31	Bis(2-ethylhexyl) phthalate		C <sub>24</sub> H <sub>38</sub> O <sub>4</sub>	390.55
40	32.437	0.12	Nonadecyltrifluoroacetate		C <sub>21</sub> H <sub>39</sub> F <sub>3</sub> O <sub>2</sub>	380.52
41	33.502	0.16	6,11-Dimethyl-2,6,10-dodecatrien-1-ol		C <sub>14</sub> H <sub>24</sub> O	208.34

TABLE 2: BIOACTIVE COMPOUNDS PRESENT IN *U. LOBATA* METHANOL LEAVE EXTRACT BY GC-MS

Pk	RT	Area %	Bioactive compound	Structure	MF	MW
1	9.634	0.15	Dodecane		C <sub>12</sub> H <sub>26</sub>	170.33
2	14.662	0.43	5-Tetradecene, (E)-		C <sub>14</sub> H <sub>28</sub>	196.37
3	14.945	0.61	Tetradecane		C <sub>14</sub> H <sub>30</sub>	198.38
4	16.492	0.35	Cyclododecane		C <sub>12</sub> H <sub>24</sub>	168.31
5	17.404	1.86	2,4-Di-tert-butylphenol		C <sub>14</sub> H <sub>22</sub> O	206.32
6	17.663	0.42	Undecanoic acid, 10-methyl-, methyl ester		C <sub>13</sub> H <sub>26</sub> O <sub>2</sub>	214.34
7	19.221	6.89	Dodecanoic acid		C <sub>12</sub> H <sub>24</sub> O <sub>2</sub>	200.31
8	19.268	2.53	Dodecanoic acid		C <sub>12</sub> H <sub>24</sub> O <sub>2</sub>	200.31
9	19.566	1.41	9-Octadecene, (E)-		C <sub>18</sub> H <sub>36</sub>	252.47

10	19.812	0.96	Hexadecane		C <sub>16</sub> H <sub>34</sub>	226.44
11	20.815	0.22	Cyclohexane,1,1'-(1,4-butanediyl) bis-		C <sub>16</sub> H <sub>30</sub>	222.40
12	21.243	0.23	Cyclotetradecane		C <sub>14</sub> H <sub>28</sub>	196.37
13	21.546	1.59	1-Dodecene		C <sub>12</sub> H <sub>24</sub>	168.32
14	22.063	0.31	Heptadecane		C <sub>17</sub> H <sub>36</sub>	240.46
15	22.273	0.22	Tridecanoic acid, 12-methyl-, methyl ester		C <sub>15</sub> H <sub>30</sub> O <sub>2</sub>	242.39
16	22.904	0.26	1,2,4-Butanetriol, trinitrate		C <sub>4</sub> H <sub>7</sub> N <sub>3</sub> O <sub>9</sub>	241.11
17	23.518	3.01	Tetradecanoic acid		C <sub>14</sub> H <sub>28</sub> O <sub>2</sub>	228.37
18	24.006	0.53	1-Octadecene		C <sub>18</sub> H <sub>36</sub>	252.47
19	24.214	0.24	Heptadecane,2 methyl-		C <sub>18</sub> H <sub>38</sub>	254.49
20	24.880	0.23	Pentadecanoic acid		C <sub>15</sub> H <sub>30</sub> O <sub>2</sub>	242.39
21	25.557	0.18	Trifluoroacetic acid, pentadecyl ester		C <sub>17</sub> H <sub>31</sub> F <sub>3</sub> O <sub>2</sub>	324.42
22	26.479	0.25	Hexadecanoic acid, methyl ester		C <sub>17</sub> H <sub>34</sub> O <sub>2</sub>	270.45
23	26.977	1.64	9-Hexadecenoic acid		C <sub>16</sub> H <sub>30</sub> O <sub>2</sub>	254.40
24	27.600	26.65	n-Hexadecanoic acid		C <sub>16</sub> H <sub>32</sub> O <sub>2</sub>	256.42
25	27.735	9.11	n-Hexadecanoic acid		C <sub>16</sub> H <sub>32</sub> O <sub>2</sub>	256.42
26	28.044	3.54	1-Octadecene		C <sub>18</sub> H <sub>36</sub>	252.47
27	28.209	0.94	Heptacosane,1-chloro-		C <sub>27</sub> H <sub>55</sub> Cl	415.17
28	28.594	0.46	Heptadecanoic acid		C <sub>17</sub> H <sub>34</sub> O <sub>2</sub>	270.45
29	28.703	0.57	Butyl eicosyl ether		C <sub>26</sub> H <sub>56</sub> OSi	412.80
30	28.854	1.50	trans-13-Octadecenoic acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.46
31	29.255	0.14	6-Octadecenoic acid, methyl ester, Z		C <sub>19</sub> H <sub>36</sub> O <sub>2</sub>	296.48
32	29.826	3.45	9-Octadecenoic acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.46
33	30.005	0.68	Octadecanoic acid		C <sub>18</sub> H <sub>36</sub> O <sub>2</sub>	284.47

34	30.091	0.28	Hexadecanoic acid, 1,1-dimethylethyl ester		C <sub>20</sub> H <sub>40</sub> O <sub>2</sub>	312.53
35	30.245	0.56	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58
36	30.310	0.10	3-Eicosene, (E)-		C <sub>20</sub> H <sub>40</sub>	280.53
37	30.599	1.59	cis-Vaccenic acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.46
38	30.791	0.74	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58
39	31.103	1.67	cis-Vaccenic acid		C <sub>18</sub> H <sub>34</sub> O <sub>2</sub>	282.46
40	31.249	2.67	Palmitoleic acid		C <sub>16</sub> H <sub>30</sub> O <sub>2</sub>	254.40
41	31.382	2.66	2-Piperidinone,N-[4-bromo-n-butyl		C <sub>9</sub> H <sub>16</sub> BrNO	234.13
42	31.483	6.06	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58
43	31.621	2.84	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58
44	31.739	2.04	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58
45	31.821	4.09	Erucic acid		C <sub>22</sub> H <sub>42</sub> O <sub>2</sub>	338.60
46	32.063	2.97	Diisooctyl phthalate		C <sub>24</sub> H <sub>38</sub> O <sub>4</sub>	390.55
47	32.437	0.17	1-Docosene		C <sub>22</sub> H <sub>44</sub>	308.58

Figure 3: FTIR Spectrum of ethanol extract of *U. lobata* leaves



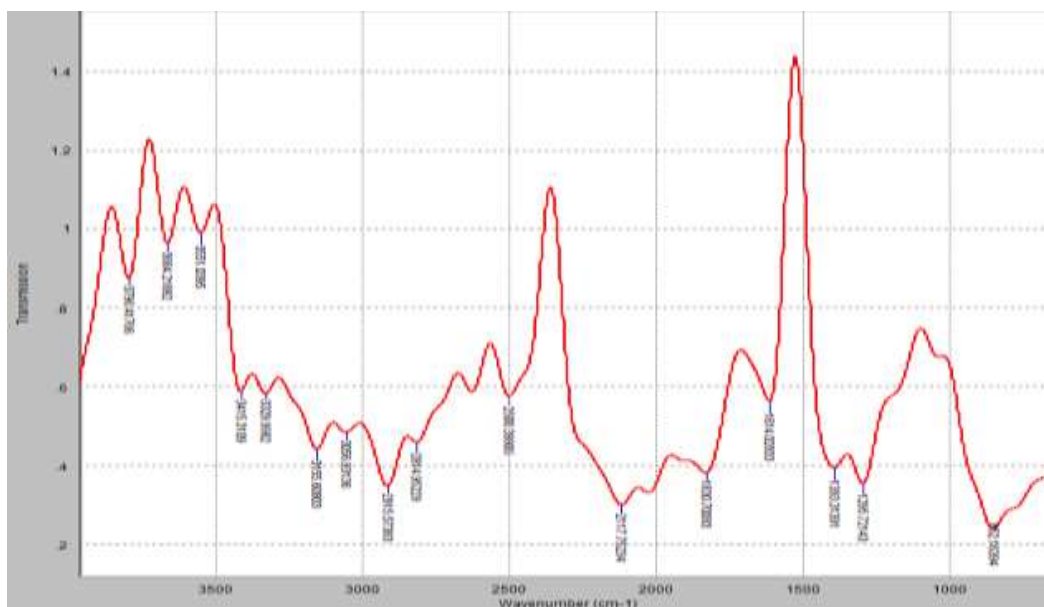


Figure 4: FTIR Spectrum of methanol extract of *U. lobata* leaves

Table 3: FTIR peak values and functional groups of ethanol extract of *U.lobata* leaves

	Wavenumber (cm <sup>-1</sup> )	Functional group/mode of vibration	Inference
1	3704.812	-OH, OH Stretching	OH in Alcohols and Phenols
2	3498.917	-NH <sub>2</sub> , NH Stretch	NH in primary amines, amides and aromatic amines,
3	3181.87	-NH <sub>2</sub> ,NH <sub>2</sub> Symmetric stretch	NH <sub>2</sub> in primary amides
4	3013.238	= CH, = CH-H Stretching	= CH in unsaturated and aromatic hydrocarbons
5	2782.866	-CH <sub>3</sub> , CH Stretch	CH <sub>3</sub> attached to N or O
6	2665.173	-CHO, CH Bending	CH in Aldehydes
7	2571.721	-OH, OH Stretching	OH in phosphorus Oxyacids
8	2227.731	C ≡ C, C ≡ C Stretch	C ≡ C in Alkynes
	2108		
9	1622.32	N-H, NH Deformation	NH in Primary amides
10	1416.411	C-N, C-N Stretch	C-N in Primary amides
11	1235.417	C-N, C-N Stretch	C-N in Primary amides
12	843.5038	1,3,5-trisubstituted benzenes, CH out-of-plane deformation	In tri-substituted benzenes

Table 4: FTIR peak values and functional groups of methanol extract of *U.lobata* leaves

S/N	Wavenumber (cm <sup>-1</sup> )	Functional group/mode of vibration	Inference
1	3664.219 3415.311	-OH , OH stretch	OH in Phenols and Alcohols
2	3155.608	NH <sub>3</sub> <sup>+</sup> , Antisymmetric stretch	NH in amino acids
3	3056.931	=CH, =CH-H stretch	CH in unsaturated and aromatic hydrocarbons
4	2915.574	-CH and -CH <sub>2</sub> -, -CH <sub>3</sub> - Antisymmetric and symmetric stretching	CH in Aliphatic compounds
5	2814.952	-CH <sub>3</sub> , CH Stretch	CH <sub>3</sub> attached to N or O
6	2500.399	NH <sub>3</sub> <sup>+</sup> , NH Stretching	NH in Amine in hydrohalides
7	2117.752	N≡C, N≡C Stretch	N≡C in isonitriles

8	1830.701	C=O, C=O Stretch	C=O in $\beta$ -lactones
9	1614.028	C = C, C = C Stretch	C = C in Vinyl ethers
10	1393.314	t-butyl groups, CH <sub>3</sub> deformations	t-butyl groups
11	1295.721	SO <sub>2</sub> , SO <sub>2</sub> Antisymmetric stretch	SO <sub>2</sub> in Sulfones
12	852.1639	R-NH <sub>2</sub> , NH <sub>2</sub> wag	NH <sub>2</sub> in Primary amines

TABLE 5: ACTIVITIES OF BIOACTIVE COMPOUNDS PRESENT IN *U. LOBATA* ETHANOL LEAVE EXTRACT

S/N	COMPOUNDS	CLASS OF COMPOUND	BIOLOGICAL ACTIVITY
1	Decane	Alkane	Possess alarm pheromonal activity <sup>21</sup> Possess tumor-promoting activity <sup>22</sup> Antifungal and Antibacterial <sup>23</sup>
2	Dodecane	Alkane	Neurotrophic action <sup>24</sup>
3	Cyclopentane, 1-hexyl-3-methyl-	Alkane	No activity reported
4	Tetradecane	Alkane	Possess tumor-promoting and cocarcinogenic activity <sup>22,25</sup> Antimicrobial, Diuretic & Antituberculosis <sup>26</sup> Antifungal <sup>27</sup> Antibacterial <sup>28</sup>
5	3-Heptafluoro-butyroxydecane	Alkane	No activity reported
6	Silane, trichlorodocosyl	Organosilicon	No activity reported
7	Phenol, 3,5-bis(1,1-dimethylethyl)	Phenol	Antimicrobial activity <sup>29</sup> Antibacterial activity <sup>30</sup>
8	Undecanoic acid, 10-methyl, methylester	FAME	Antioxidant, increase aromatic amino acid decarboxylase activity <sup>31</sup>
9	Dodecanoic acid	Fatty acid	Antibacterial and Antioxidant activities <sup>32</sup>
10	9-Eicosene	Fatty acid	Antimicrobial <sup>33</sup>
11	Hexadecane	Alkane	Antifungal, antibacterial and antioxidant activities <sup>34</sup> Cytotoxicity, antimicrobial, antioxidant, antipyretic, anthelmintic, anti-inflammatory, antidiarrhea, antidiabetic <sup>35</sup>
12	Dodecyl acrylate	Acrylate ester	Antioxidant and antimicrobial activities <sup>36</sup>
13	Tetradecanoic acid	Fatty acid	Antifungal, antioxidant, cancer preventive, nematocide, hypocholesterolemic, lubricant <sup>31</sup>
14	9-Eicosene	Fatty acid	Antimicrobial <sup>33</sup>
15	Octadecane	Alkane	Antibacterial and antifungal activity <sup>26</sup> Possess anti-inflammatory, detoxification, cough, lung disease, fever, cold, antioxidant, antiseptic and anticorrosion activities, <sup>35</sup>
16	Pentadecanoic acid, 14-methyl-methylester	FAME	Antioxidant <sup>37</sup>
17	n-Hexadecanoic acid	Fatty acid	Anti-inflammatory, antioxidant, cancer as well as hypocholesterolemic acid preventive activities <sup>38</sup> Nematocide, pesticide, lubricant, antiandrogenic activity <sup>31</sup>
18	1-Octadecene	Alkene	Antibacterial, antioxidant and anticancer activity <sup>39</sup>
19	Methoxyacetic acid, 2-tradecylester	Fatty-acid ester	Antibacterial activity <sup>40</sup>
20	Pentadecafluorooctanoic acid, octadecyl ester	Fatty-acid ester	No activity reported

21	9-Octadecenoic acid	Fatty acid	Antimicrobial activity <sup>31</sup>
22	Cyclohexane, butyl	Cycloalkane	No activity reported
23	1-Eicosanol	Alcohol	Antimalarial, antifungal and antioxidant activities <sup>31</sup>
24	Cis-13-Octadecenoic acid, methyl ester	FAME	Therapeutic uses in medicine and surgery <sup>41</sup>
25	Cis-Vaccenic acid	MUFA	Anti-hypocholesterolemic and anti-inflammatory activities <sup>41</sup>
26	Methyl Stearate	FAME	Antihelminthic(nematodes), inhibits GABA aminotransferase, antinociceptive, regulates intestinal lipid metabolism, anti-inflammatory, inhibits gastric acid activities <sup>42</sup>
27	9-Octadecanoic acid,	Fatty acid	Antimicrobial activity <sup>31</sup>
28	Cyclohexadecane, 1,2-diethyl	Cycloalkane	No activity reported
29	9-Octadecenoic acid	Fatty acid	Antimicrobial activity <sup>31</sup>
30	Octadecanoic acid	Fatty acid	Antimicrobial activity <sup>31</sup>
31	Oleic acid	MUFA	Antimicrobial activity <sup>43</sup> Antifungal, anti-inflammatory, antioxidant and antibacterial activities <sup>41</sup>
32	Oleic acid	MUFA	Antimicrobial activity <sup>43</sup> Antifungal, anti-inflammatory, antioxidant and antibacterial activities <sup>41</sup>
33	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>
34	Cycloeicosane	Cycloalkane	No activity reported
35	Cyclohexadecane, 1,2 -diethyl	Cycloalkane	No activity reported
36	5-Eicosene	Alkene	Antimicrobial activity <sup>44</sup>
37	3-Eicosene	Alkene	Antimicrobial, anti-hyperglycemic, cytotoxic activity, antioxidant and insecticide activities <sup>35</sup>
38	Heptadecylheptafluorobutyrate	Ester	No activity reported
39	Bis-(2-ethylhexyl) phthalate	Phthalate Ester	Antimicrobial and cytotoxic activities <sup>45</sup>
40	Nonadecyltrifluoroacetate	Ester	No activity reported
41	6,11-Dimethyl-2,6,10-dodecatrien-1-ol	Phenol	Antimicrobial activity <sup>46</sup>

TABLE 6: ACTIVITIES OF BIOACTIVE COMPOUNDS PRESENT IN *U. LOBATA* METHANOL LEAVE EXTRACT

Pk	Bioactive compound	Class of compound	Biological activity
1	Dodecane	Alkane	Antibacterial activity Antifungal activity <sup>47</sup> Enhances antifungal activity. Possess tumor-promoting Activity <sup>25</sup>
2	5-Tetradecene	Alkene	No activity reported
3	Tetradecane	Alkane	Possess tumor-promoting and cocarcinogenic activity <sup>25</sup> ; Antimicrobial, Diuretic & Antituberculosis <sup>22</sup> ; Antifungal <sup>26-28</sup> , Antibacterial, Antipyretic and bronchitis <sup>35</sup>
4	Cyclododecane	Cycloalkane	Serves as a hydrophobic mask, melt and as a temporary consolidant agent <sup>48</sup>
5	2,4-Di-tert-butylphenol	Phenol	Antibacterial, Antimalarial, Anticancer, Antifungal activities <sup>49-52</sup>
6	Undecanoic acid, 10-methyl-, methyl ester	FAME	Antioxidant, increase aromatic amino acid decarboxylase activity <sup>31</sup>
7	Dodecanoic acid	Fatty acid	Antibacterial and Antioxidant activities <sup>32</sup>
8	Dodecanoic acid	Fatty acid	Antibacterial and Antioxidant activities <sup>32</sup>

9	9-Octadecene	Alkene	Antifungal, antioxidant, anticarcinogenic and antimicrobial activity <sup>53-54</sup>
10	Hexadecane	Alkane	Antifungal, antibacterial and antioxidant activities <sup>34</sup> Cytotoxicity, antimicrobial, antioxidant, antipyretic, anthelmintic, anti-inflammatory, antidiarrheal, antidiabetic <sup>35</sup>
11	Cyclohexane,1,1'-(1,4-butanediyl) bis-	Cycloalkane	No activity reported
12	Cyclotetradecane	Cycloalkane	Antimicrobial <sup>55</sup>
13	1-Dodecene	Alkene	Antibacterial activity <sup>56</sup>
14	Heptadecane	Alkane	Antibacterial activity <sup>57</sup>
15	Tridecanoic acid, 12-methyl-, methyl ester	FAME	No activity reported
16	1,2,4-Butanetriol, trinitrate	Nitrate ester	An energetic plasticizer <sup>58</sup>
17	Tetradecanoic acid	Fatty acid	Antifungal, antioxidant, cancer preventive, nematocide, hypocholesterolemic, lubricant <sup>31</sup>
18	1-Octadecene	Alkene	Antibacterial activity; Antioxidant activity <sup>39,59</sup> Anticancer activity <sup>60,39</sup>
19	Heptadecane,2 methyl-	Alkane	No activity reported
20	Pentadecanoic acid	Fatty acid	Anticancer <sup>61</sup>
21	Trifluoroacetic acid, pentadecyl ester	Fatty acid ester	No activity reported
22	Hexadecanoic acid, methyl ester	FAME	Promotes aortic dilation <sup>62</sup> ,Promotes membrane autolysis <sup>63</sup> ,Inhibits phagocytosis and influences nitric oxide production in some cells <sup>64</sup>
23	9-Hexadecenoic acid	Fatty acid	Antimicrobial <sup>65</sup>
24	n-Hexadecanoic acid	Fatty acid	Cancer preventing activities, anti-inflammatory,antioxidant, hypocholesterolemic <sup>38</sup> , Nematicidal, pesticidal, lubricant and anti-androgenic <sup>31</sup>
25	n-Hexadecanoic acid	Fatty acid	Cancer preventing activities, anti-inflammatory,antioxidant, hypocholesterolemic <sup>38</sup> , Nematicidal, pesticidal, lubricant and anti-androgenic <sup>31</sup>
26	1-Octadecene	Alkene	Antibacterial, antioxidant and anticancer <sup>39</sup>
27	Heptacosane,1-chloro-	Chloroalkane	No activity reported
28	Heptadecanoic acid	Fatty acid	Antioxidant <sup>66</sup>
29	Butyl eicosyl ether	Ether	No activity reported
30	trans-13-Octadecenoic acid	Fatty acid	Anti-inflammatory activity <sup>67</sup>
31	6-Octadecenoic acid, methyl ester	FAME	Possess strong analgesic, anti-inflammatory and antipyretic activity <sup>68</sup>
32	9-Octadecenoic acid	Fatty acid	Antimicrobial <sup>31</sup>
33	Octadecanoic acid	Fatty acid	Antimicrobial <sup>31</sup>
34	Hexadecanoic acid, 1,1-dimethylethyl ester	FAME	No activity reported
35	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>
36	3-Eicosene	Alkene	Antimicrobial, anti-hyperglycemic, cytotoxic activity, antioxidant, insecticide activity <sup>35</sup>
37	cis-Vaccenic acid	MUFA	Anti-hypocholesterolemic and anti-inflammatory <sup>41</sup>
38	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>
39	cis-Vaccenic acid	MUFA	Anti-hypocholesterolemic and anti-inflammatory <sup>41</sup>

40	Palmitoleic acid	MUFA	Anti-inflammatory; Increases insulin sensitivity, inhibits hepatic steatosis. <sup>69</sup>  Ameliorates development of hypertriglyceridemia and hyperglycemia, reduces body weight increase  Down-regulates mRNA expression of proinflammatory adipocytokine genes in mice <sup>70</sup> .
41	2-Piperidinone,N-[4-bromo-n-butyl	Delta-lactams	Antimicrobial activity <sup>71</sup>
42	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>
43	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>
44	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>
45	Erucic acid	MUFA	Regulates mesenchymal stem cell differentiation into osteoblasts and adipocyte <sup>72</sup> ; Broad-spectrum antiviral activity against IAV <sup>73</sup> ; Anti-inflammatory/Pro-inflammatory amplification effect. Inhibits NF- $\kappa$ B and p38 MAPK
46	Diisooctyl phthalate	Phthalate ester	Antimicrobial and antifouling <sup>74</sup>  Antibacterial <sup>75</sup>
47	1-Docosene	Alkene	Antibacterial activity <sup>43</sup>

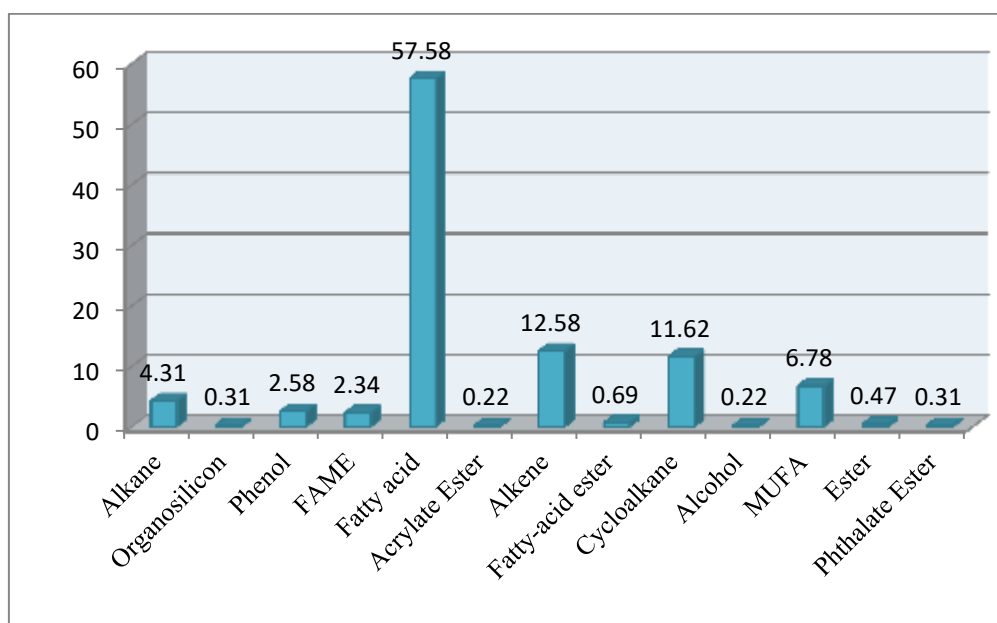


Figure 5: Percentage Abundance of Classes of phytochemicals in *U. lobata* ethanol leaf extract

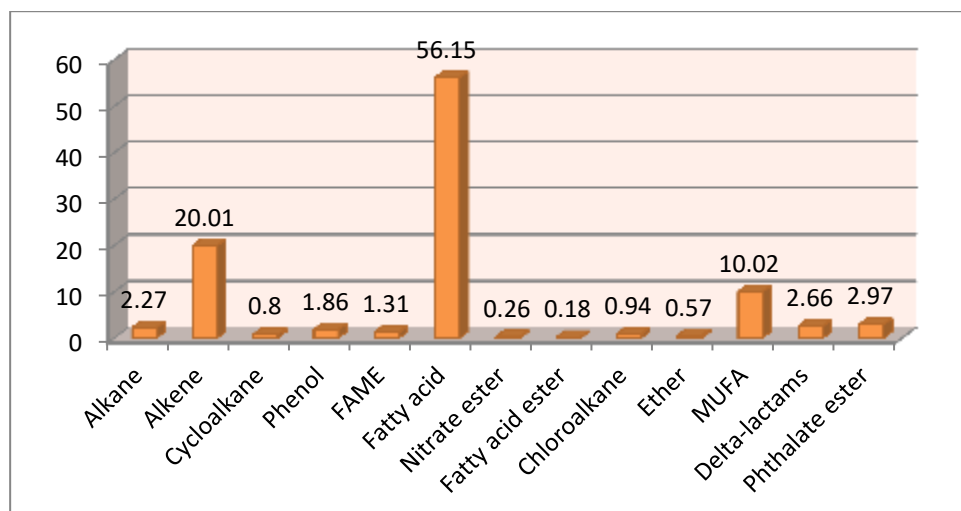


Figure 6: Percentage Abundance of Classes of phytochemicals in *U. lobata* methanol leaf extract



**Table 7: Bioactive compounds common to ethanol and methanol extracts of *U.lobata* leaves**

Bioactive compounds		Percentage Abundance in extraction solvents (%)	
		Ethanol	Methanol
1.	Dodecane	0.61	0.15
2.	Undecanoic acid, 10-methyl-, methyl ester	1.11	0.42
3.	Dodecanoic acid	13.43	6.89, 2.53
4.	Hexadecane	0.72	0.96
5.	Tetradecanoic acid	2.14	3.01
6.	n-Hexadecanoic acid	11.73	26.65, 9.11
7.	1-Octadecene	1.11	3.54
8.	9-Octadecenoic acid	16.8, 2.02, 0.91	3.45
9.	cis-Vaccenic acid	0.25	1.59, 1.67
10.	Octadecanoic acid	9.78	0.68
11.	1-Docosene	9.57	2.84, 0.56, 0.74, 6.06, 2.04, 0.17
12.	3-Eicosene, (E)-	0.27	0.1

**Figure 7: Whole plant of *U. lobata***

## CONCLUSION

GC-MS and FT-IR investigation of ethanol and methanol leave extracts of *Urena lobata* demonstrated the abundance of bioactive phytochemicals and their inherent functional groups. These bioactive phytocompounds have a wide range of biological and therapeutic actions, lending credence to the plant's ethnomedicinal use for the treatment of various disease conditions amongst ethnic groups.

## CONFLICT OF INTEREST

No conflict of interest exist

## REFERENCES

1. Saranya D and Sekar J. GC-MS and FT-IR Analyses of Ethylacetate Leaf extract of *Abutilon indicum* (L.) Sweet. *International Journal of Advanced Research in Biological Sciences*. 2016; 3(2):193-197. <http://s-o-i.org/1.15/ijarbs-2016-3-2-26>
2. Patil VT and Jadhav VD. GC-MS and FTIR analysis of methanolic leaf extract of *Rhynchosia minima* (L.) DC. *Current Botany* 2020; 11:221-225 <https://doi.org/10.25081/cb.2020.v11.6415>

3. Sahithya S and Krishnaveni C. FT-IR and GC-MS Analysis of Stem Extract of Ethanomedicinal Plant: Bridelia Montana (Roxb.) Willd. *Asian Journal of Biological and Life Sciences*. 2022; 2(2). <http://dx.doi.org/10.5530/ajbls.2022.11.61>
4. Konappa N, Udayashankar AC, Krishnamurthy S, Pradeep CK, Chowdappa S, and Jogaiah S. (2020). GC-MS analysis of phytoconstituents from Amomum nilgiriicum and molecular docking interactions of bioactive serverogen in acetate with target proteins. *Scientific reports*. 2020; 10(1), 1-23. <https://doi.org/10.1038/s41598-020-73442-0>
5. Adeloye OA, Akinpelu AD, Ogundaini OA and Obafemi, AC Studies on antimicrobial, antioxidant and phytochemical analysis of Urenalobata leave extract. *Journal of Physical and Natural Sciences* (2007); 1(2)
6. Islam MT and Uddin MA. A revision on Urenalobata L. *International Journal of Medicine*. (2017); 5(11). 126-131. <https://doi.org/10.14419/ijm.v5i11.7525>
7. Njoku CE, Alaneme KK, Omotoyinbo JA, Ekeleme AC, Ugwu EI, and Ikele US. Phytochemical, Proximate and Mineral Analyses of Urenalobata stems from Imo State Nigeria. In *IOP Conference Series: Materials Science and Engineering* 2021; 1107(1), 012183. <https://doi.org/10.1088/1757-899X/1107/1/012183>
8. Fagbohun ED, Asare RR, and Egbebi AO. Chemical composition and antimicrobial activities of Urenalobata L. (Malvaceae). *Journal of Medicinal Plants Research*. 2012; 6(12), 2256-2260.
9. Jia L, Jing LL, Zhou SA, and Kong DY. Three new flavonoid glycosides from Urenalobata. *Journal of Asian natural products research*. 2011; 13(10), 907-914. <https://doi.org/10.1080/10286020.2011.599802>
10. Shaba EY, Mathew TJ, Otori AA, Tsado AN, Mustapha S and NajeebWali I. Nutritional assessment of Urenalobata leaves. *Magnesium*. (2017); 66, 0-19.
11. Islam MT, Ibrahim M, Ahsan MQ, Chowdhury MMU, Hossain MA, and Rashid MA. Phytochemical and Pharmacological Investigations of Urarialagopodies DC. and Urenalobata L. *Dhaka University Journal of Pharmaceutical Sciences*. (2012); 11(1), 65-69. <https://doi.org/10.3329/dujps.v11i1.12490>
12. Purnomo Y, Makdasari J and Fatahillah FI. Inhibitory activity of Urenalobata leaf extract on alpha-amylase and alpha-glucosidase: in vitro and in silico approach. *Journal of Basic and Clinical Physiology and Pharmacology*. (2021); 32(4), 889-894 <https://doi.org/10.1515/jbcp-2020-0430>
13. Mathappan R, Krishnan SK, Sujeet S, and Tribedi S. Evaluation of antitumor activity of Urenalobata against Ehrlich ascites carcinoma treated mice. *Oriental Pharmacy and Experimental Medicine*. (2019); 19(1), 21-26. <https://doi.org/10.1007/s13596-018-0342-x>
14. Gao XL, Liao Y, Wang J, Liu XY, Zhong K, Huang YN, ... and Xu, ZJ. Discovery of a potent anti-yeast triterpenoidsaponin, clematoside-S from Urenalobata L. *International journal of molecular sciences*. 2015; 16(3), 4731-4743. <https://doi.org/10.3390/ijms16034731>
15. Ali MS, Faruq KO, & Rahman MAA. Antioxidant and cytotoxic activities of methanol extract of Urenalobata (L) Leaves. *The Pharma Innovation*. 2013; 2(2).
16. Mathappan R, Umachigi SP and Prasanth VV. Wound healing activity of the methanolic extract of Urenalobata Linn. *International Journal of Pharmaceutical and Chemical Sciences*. 2013; 2(2), 793-800.
17. Babu SS, Madhuri DB and Ali SL. A pharmacological review of Urenalobata plant. *Asian Journal of Pharmaceutical and Clinical Research*. 2016; 20-22.
18. Handayani N and Gofur A. The Potency of pulutan (Urenalobata L.) leaves decoction as antifertility based on its effect on uterine development of balb c mice (Mus musculus). *Jurnal Kedokteran Hewan* 2017; 11(4), 153-155. <https://doi.org/10.21157/j.ked.hewan.v11i4.6261>
19. Dhanapal R, Ratna JV, Gupta M, and Sarathchandran I. Preliminary study on antifertility activity of Enicostemma axillare leaves and Urenalobata root used in Indian traditional folk medicine. *Asian Pacific journal of tropical medicine*. 2012; 5(8), 616-622. [https://doi.org/10.1016/S1995-7645\(12\)60127-8](https://doi.org/10.1016/S1995-7645(12)60127-8)
20. Odo IF, Ezeanyika LUS, Ogugua VN, Joshua PE and Okagu IU. FTIR and GC-MS spectroscopic analysis of methanol and chloroform extracts of Brenaniabriei root bark. *AJ Res. Commun*. 2017; 5(3), 44-54.
21. Fujiwara-Tsujii N, Yamagata N, Takeda T, Mizunami M and Yamaoka R. Behavioral responses to the alarm pheromone of the ant Camponotus obscuripes (Hymenoptera: Formicidae). *Zoological science*. 2016; 23(4), 353-358. <https://doi.org/10.2108/zsj.23.353>
22. Van Duuren BL and Goldschmidt BM. Cocarcinogenic and tumor-promoting agents in tobacco carcinogenesis. *Journal of the National Cancer Institute*. 1976; 56(6), 1237-1242. <https://doi.org/10.1093/jnci/56.6.1237>
23. Asghari G, Jalali M, and Sadoughi E. Antimicrobial activity and chemical composition of essential oil from the seeds of Artemisia aucheri Boiss. *Jundishapur journal of natural pharmaceutical products*. 2012; 7(1), 11-15.
24. Kumaresan S, Senthikumar V, Stephen A and Balakumar BS. GC-MS Analysis and Pass-Assisted Predictions of Biological Activity Spectra of Extract of Phomopsis sp. Isolated from Andrographis paniculata. *World Journal of Pharmaceutical Research* 4. 2015.
25. Bird MG. *Early biochemical and morphological changes in the mouse skin from cutaneous application of petrochemical and petroleum derived materials*. University of Surrey (United Kingdom) 1985.
26. Girija S, Duraipandiyan V, Kuppusamy PS, Gajendran H, Rajagopal R. Chromatographic characterization and GC-MS evaluation of the bioactive constituents with antimicrobial potential from the pigmented ink of Loligoduvauli. *International Scholarly Research Notices*. 2014; 2014. <http://dx.doi.org/10.1155/2014/820745>
27. Guo L, Wu JZ, Han T, Cao T, Rahman K, Qin LP. Chemical composition, antifungal and antitumor properties of ether extracts of Scapaniaverrucosa Heeg. and its endophytic fungus Chaetomium fusiforme. *Molecules*. 2008 Sep 4; 13(9):2114-25. <https://doi.org/10.3390/molecules13092114>
28. Nahid R, Ali S, Farshid S. Antimicrobial activity and constituents of the hexane extracts from leaf and stem of Origanum vulgare L. ssp. Viride (Boiss.) Hayek. growing wild in Northwest Iran. *Journal of Medicinal Plants Research*. 2012 Apr 9; 6(13):2681-5.
29. Dhanya KI, Swati VI, Vanka KS, Osborne WJ. Antimicrobial activity of Ulvareticulata and its endophytes. *Journal of Ocean University of China*. 2016 Apr; 15(2):363-9. <https://doi.org/10.1007/s11802-016-2803-7>
30. Abdullah AS, Mirghani ME, Jamal P. Antibacterial activity of Malaysian mango kernel. *African Journal of Biotechnology*. 2011; 10(81):18739-48.
31. Duke J, Bogenschutz MJ. Dr. Duke's phytochemical and ethnobotanical databases. Washington, DC: USDA, Agricultural Research Service; 1994.
32. Renugadevi K, Nachiyar VC, Zaveri M. Bioactivity of Dodecanoic Acid Extracted from Geitlerinema sp. TRV57. *Indian Journal of Pharmaceutical Education And Research*. 2021 Jan 1; 55(1):224-31.
33. Ugbogu EA, Akubugwo IE, Ude VC, Gilbert J, Ekeanyanwu B. Toxicological evaluation of phytochemical characterized aqueous extract of wild dried Lentinus squarrosulus (Mont.) mushroom in rats. *Toxicological research*. 2019 Apr; 35(2):181-90. <https://doi.org/10.5487/TR.2019.35.2.181>
34. Faridha Begum I, Mohankumar R, Jeevan M, Ramani K. GC-MS analysis of bio-active molecules derived from Paracoccupantotrophus FMR19 and the antimicrobial activity against bacterial pathogens and MDROs. *Indian journal of microbiology*. 2016 Dec; 56(4):426-32. <https://doi.org/10.1007/s12088-016-0609-1>

35. Banakar P, Jayaraj M. GC-MS analysis of bioactive compounds from ethanolic leaf extract of *Waltheria indica* Linn. and their pharmacological activities. *Int. J. Pharm. Sci. Res.* 2018 May 1; 9(5):2005-10. [http://dx.doi.org/10.13040/IJPSR.0975-8232.9\(5\).2005-10](http://dx.doi.org/10.13040/IJPSR.0975-8232.9(5).2005-10)
36. Joo SS, Kim YB, Lee DI. Antimicrobial and Antioxidant Properties of Secondary Metabolites from White Rose Flower. *Antimicrobial and Antioxidant Properties of Secondary Metabolites from White Rose Flower*. *Plant Pathology Journal*. 2010; 26(1):57-62.
37. Sharma S, Saxena DC, Riar CS. Changes in the GABA and polyphenols contents of foxtail millet on germination and their relationship with in vitro antioxidant activity. *Food Chemistry*. 2018 Apr 15; 245:863-70. <https://doi.org/10.1016/j.foodchem.2017.11.093>
38. Mazumder K, Nabila A, Aktar A, Farahnaky A. Bioactive variability and in vitro and in vivo antioxidant activity of unprocessed and processed flour of nine cultivars of Australian lupin species: a comprehensive substantiation. *Antioxidants*. 2020 Mar 27; 9(4):282. <https://doi.org/10.3390/antiox9040282>
39. Belakhdar G, Benjouad A, Abdennebi EH. Determination of some bioactive chemical constituents from *Thesium humile* Vahl. *J Mater Environ Sci*. 2015; 6(10):2778-83.
40. Olusola AO, Olabode Ogunsina I, Samson Ayedogbon O, Olusola Adesayo O. In vivo Antimalarial activity of bark extracts of *Lannea acida* (Anacardiaceae) and chloroquine against *Plasmodium berghei* in mice. *International Journal of Biomedical and Clinical Sciences*. 2020; 5(3):229-35. <https://doi.org/10.1007/s12639-016-0815-0>
41. Awonyemi OI, Abegunde SM, Olabiran TE. Analysis of bioactive compounds from *Raphia taedigera* using gas chromatography-mass spectrometry. *Eur Chem Commun*. 2020; 2(8):933-44. <https://doi.org/10.22034/ecc.2020.107898>
42. Adnan M, Nazim Uddin Chy M, Mostafa Kamal AT, Azad MO, Paul A, Uddin SB, Barlow JW, Faruque MO, Park CH, Cho DH. Investigation of the biological activities and characterization of bioactive constituents of *Ophiorrhizarugosa* var. *prostrata* (D. Don) & Mondal leaves through in vivo, in vitro, and in silico approaches. *Molecules*. 2019 Apr 8; 24(7):1367. <https://doi.org/10.3390/molecules24071367>
43. Albratty M, Alhazmi HA, Meraya AM, Najmi A, Alam MS, Rehman Z, Moni SS. Spectral analysis and Antibacterial activity of the bioactive principles of *Sargassum tennerrimum* J. Agardh collected from the Red sea, Jazan, Kingdom of Saudi Arabia. *Brazilian Journal of Biology*. 2021 Oct 18; 83. <https://doi.org/10.1590/1519-6984.249536>
44. Yogeswari S, Ramalakshmi S, Neelavathy R, Muthumary JY. Identification and comparative studies of different volatile fractions from *Monochaetia kansensis* by GCMS. *Global Journal of Pharmacology*. 2012; 6(2):65-71.
45. Lotfy MM, Hassan HM, Hetta MH, El-Gendy AO, Mohammed R. Di-(2-ethylhexyl) Phthalate, a major bioactive metabolite with antimicrobial and cytotoxic activity isolated from River Nile derived fungus *Aspergillus awamori*. *Beni-Suef University Journal of Basic and Applied Sciences*. 2018 Sep 1; 7(3):263-9. <https://doi.org/10.1016/j.bjbas.2018.02.002>
46. Das SK, Dash S, Thatoi H, Patra JK. In vitro  $\alpha$ -amylase and  $\alpha$ -glucosidase Inhibition, Antioxidant, Anti-Inflammatory Activity and GC-MS Profiling of *Avicennia alba* Blume. *Combinatorial Chemistry & High Throughput Screening*. 2020 Nov 1; 23(9):945-54.
47. Stopiglia CD, Collares FM, Ogliari FA, Piva E, Fortes CB, Samuel SM, Scrofernek ML. Antimicrobial activity of [2-(methacryloyloxy) ethyl] trimethylammonium chloride against *Candida* spp. *Revista Iberoamericana de Micología*. 2012 Jan 1; 29(1):20-3. <https://doi.org/10.1016/j.riam.2011.03.003>
48. Rowe S, Rozeik C. The uses of cyclododecane in conservation. *Studies in Conservation*. 2008 Jun 1; 53(sup2):17-31. <https://doi.org/10.1179/sic.2008.53.Supplement-2.17>
49. Seenivasan A, Manikkam R, Kaari M, Sahu AK, Said M, Dastager SG. 2, 4-di-tert-butylphenol (2, 4-DTBP) purified from *Streptomyces* sp. KCA1 from *Phyllanthus niruri*: isolation, characterization, antibacterial and anticancer properties. *Journal of King Saud University-Science*. 2022 May 15:102088. <https://doi.org/10.1016/j.jksus.2022.102088>
50. Kusch P, Deininger S, Specht S, Maniako R, Haubrich S, Pommerening T, Lin PK, Hoerauf A, Kaiser A. In vitro and in vivo antimalarial activity assays of seeds from *Balanites aegyptiaca*: compounds of the extract show growth inhibition and activity against plasmodial aminopeptidase. *Journal of parasitology research*. 2011 May 25; 2011. <https://doi.org/10.1155/2011/368692>
51. Nair RV, Jayasree DV, Biju PG, Baby S. Anti-inflammatory and anticancer activities of erythrodil-3-acetate and 2, 4-di-tert-butylphenol isolated from *Humboldtia nujuga*. *Natural product research*. 2020 Aug 17; 34(16):2319-22. <https://doi.org/10.1080/14786419.2018.1531406>
52. Belghit S, Driche EH, Bijani C, Zitouni A, Sabaou N, Badji B, Mathieu F. Activity of 2, 4-Di-tert-butylphenol produced by a strain of *Streptomyces mutabilis* isolated from a Saharan soil against *Candida albicans* and other pathogenic fungi. *Journal de mycologie medicale*. 2016 Jun 1; 26(2):160-9. <https://doi.org/10.1016/j.mycmed.2016.03.001>
53. Akpuaka A, Ekwenchi MM, Dashak DA, Dildar A. Biological activities of characterized isolates of n-hexane extract of *Azadirachta indica* A. Juss (Neem) leaves. *Nature and science*. 2013; 11(5):141-7.
54. Godwin A, Akinpelu BA, Makeinde AM, Aderogba MA, Oyedapo OO. Identification of n-hexane fraction constituents of *Archidium ohioense* (Schimp. Ex Mull) extract using GC-MS technique. *British Journal of Pharmaceutical Research*. 2015 Jan 1; 6(6):366-75.
55. Chuah XQ, Okechukwu PN, Amini F, Teo SS. Eicosane, pentadecane and palmitic acid: The effects in in vitro wound healing studies. *Asian Pacific Journal of Tropical Biomedicine*. 2018 Oct 1; 8(10):490.
56. Togashi N, Shiraishi A, Nishizaka M, Matsuoka K, Endo K, Hamashima H, Inoue Y. Antibacterial activity of long-chain fatty alcohols against *Staphylococcus aureus*. *Molecules*. 2007 Feb 5; 12(2):139-48. <https://doi.org/10.3390/12020139>
57. Uma B, Parvathavarthini R. Antibacterial effect of hexane extract of sea urchin, *Temnopleurus alexandri* (Bell, 1884). *International Journal of PharmTech Research*. 2010; 2(3):1677-80.
58. Huo N, Yang S, Wei Z, Li SS, Xia JB, Li J. Photoresponsive and gas sensing field-effect transistors based on multilayer WS<sub>2</sub> nanoflakes. *Scientific reports*. 2014 Jun 9; 4(1):1-9. <https://doi.org/10.1038/srep05209>
59. Mishra PM, Sree A. Antibacterial activity and GCMS analysis of the extract of leaves of *Finlaysonia obovata* (a mangrove plant). *Asian Journal of Plant Sciences*. 2007. <https://dx.doi.org/10.3923/ajps.2007.168.172>
60. Lee YS, Kang MH, Cho SY, Jeong CS. Effects of constituents of *Amomum xanthioides* on gastritis in rats and on growth of gastric cancer cells. *Archives of pharmacological research*. 2007 Apr; 30(4):436-43. <https://doi.org/10.1007/BF02980217>
61. To NB. *Pentadecanoic acid as a novel anti-cancer agent in human breast cancer stem-like MCF-7/SC cells* (Doctoral dissertation, 제주대학교 대학원).
62. Hagr T, Adam I. Phytochemical analysis, antibacterial and antioxidant activities of essential Oil from hibiscus sabdariffa (L) Seeds, (Sudanese Karkadi). *Prog. Chem. Biochem. Res*. 2020; 3(3):194-201.
63. Ajoku GA, Okwute SK, Okogun JI. Isolation of hexadecanoic acid methyl ester and 1, 1, 2-ethanetricarboxylic acid-1-hydroxy-1, 1-dimethyl ester from the calyx of green Hibiscus Sabdariffa (Linn). *Natural Products Chemistry & Research*. 2015 Feb 6.
64. Lohdip AM, Oyewale AO, Amupitan JO. Isolation and characterization of hexadec-12-enoic acid from the stem of *Chenopodium ambrosioides* (Linn). *J Chem Soc Nigeria*. 2014; 39:32-7.



65. Ubaid JM, Hussein HM, Hameed IH. Analysis of bioactive compounds of *Tribolium castaneum* and evaluation of anti-bacterial activity. *International Journal of Pharmaceutical and Clinical Research*. 2016;8(7):655-70.
66. Ashley EA, Dhorda M, Fairhurst RM, Amaratunga C, Lim P, Suon S, Sreng S, Anderson JM, Mao S, Sam B, Sopha C. Spread of artemisinin resistance in *Plasmodium falciparum* malaria. *New England Journal of Medicine*. 2014 Jul 31; 371(5):411-23.
67. Hameed IH, Altameme HJ, Mohammed GJ. Evaluation of antifungal and antibacterial activity and analysis of bioactive phytochemical compounds of *Cinnamomum zeylanicum* (Cinnamon bark) using gas chromatography-mass spectrometry. *Oriental Journal of Chemistry*. 2016; 32(4):1769. <http://dx.doi.org/10.13005/ojc/320406>
68. Jaddoa HH, Hameed IH, Mohammed GJ. Analysis of volatile metabolites released by *Staphylococcus aureus* using gas chromatography-Mass spectrometry and determination of its antifungal activity. *Oriental Journal of Chemistry*. 2016 Jan 1; 32(4):8-24. <http://dx.doi.org/10.13005/ojc/320439>
69. Astudillo AM, Meana C, Guijas C, Pereira L, Lebrero P, Balboa MA, Balsinde J. Occurrence and biological activity of palmitoleic acid isomers in phagocytic cells. *Journal of Lipid Research*. 2018 Feb 1; 59(2):237-49. <https://doi.org/10.1194/jlr.M079145>
70. Yang ZH, Miyahara H, Hatanaka A. Chronic administration of palmitoleic acid reduces insulin resistance and hepatic lipid accumulation in KK-Ay Mice with genetic type 2 diabetes. *Lipids in Health and disease*. 2011 Dec; 10(1):1-8. <https://doi.org/10.1186/1476-511X-10-120>
71. Al-Salman HN. Antimicrobial activity of the compound 2-Piperidinone, N-[4-Bromo-n-butyl]-extracted from pomegranate peels. *Asian Journal of Pharmaceutics (AJP)*. 2019 Feb 9; 13(01). <https://doi.org/10.22377/ajp.v13i01.3008>
72. Takahashi A, Dohi H, Egashira Y, Hirai S. Erucic acid derived from rosemary regulates differentiation of mesenchymal stem cells into osteoblasts/adipocytes via suppression of peroxisome proliferator-activated receptor  $\gamma$  transcriptional activity. *Phytotherapy Research*. 2020 Jun; 34(6):1358-66. <https://doi.org/10.1002/ptr.6607>
73. Liang X, Huang Y, Pan X, Hao Y, Chen X, Jiang H, Li J, Zhou B, Yang Z. Erucic acid from *Isatis indigotica* Fort. suppresses influenza A virus replication and inflammation in vitro and in vivo through modulation of NF- $\kappa$ B and p38 MAPK pathway. *Journal of pharmaceutical analysis*. 2020 Apr 1; 10(2):130-46. <https://doi.org/10.1016/j.jpha.2019.09.005>
74. Ingole SN. Phytochemical analysis of leaf extract of *Ocimum americanum* L.(Lamiaceae) by GCMS method. *World Scientific News*. 2016(37):76-87.
75. Chen S, Liu J, Gong H, Yang D. Identification and antibacterial activity of secondary metabolites from *Taxus endophytic fungus*. *Sheng wu Gong Cheng xuebao/Chinese Journal of Biotechnology*. 2009 Mar 1; 25(3):368-74.
76. Fadillah UF, Hambali E, Muslich M. Identifikasi Senyawa Aktif Ekstrak Daun Pulutan (*Urena lobata* L) dengan GC-MS: Identification of Active Compound in Pulutan (*Urena lobata* L) Leaf extract using GC-MS. *Jurnal Sains dan Kesehatan*. 2020 Jun 30; 2(3):217-21. <https://doi.org/10.25026/jskv2i3.155>
77. Queiroga Neto V, Bora PS, Diniz ZN, Cavaleiro JM, Queiroga KF. *Dipteryx lacunifera* seed oil: characterization and thermal stability. *Ciência e Agrotecnologia*. 2009; 33:1601-7. <https://doi.org/10.1590/S1413-70542009000600020>
78. Venn-Watson SK, Butterworth CN. Broader and safer clinically-relevant activities of pentadecanoic acid compared to omega-3: Evaluation of an emerging essential fatty acid across twelve primary human cell-based disease systems. *PloS one*. 2022 May 26; 17(5):e0268778. <https://doi.org/10.1371/journal.pone.0268778>
79. Movasaghi Z, Rehman S, ur Rehman DI. Fourier transform infrared (FTIR) spectroscopy of biological tissues. *Applied Spectroscopy Reviews*. 2008 Feb 1; 43(2):134-79. <https://doi.org/10.1080/05704920701829043>
80. Ashokkumar R, Ramaswamy M. Phytochemical screening by FTIR spectroscopic analysis of leaf extracts of selected Indian medicinal plants. *International journal of Current Microbiology and applied Sciences*. 2014; 3(1):395-406.