

Available online on 15.05.2019 at <http://jddtonline.info>

Journal of Drug Delivery and Therapeutics

Open Access to Pharmaceutical and Medical Research

© 2011-18, publisher and licensee JDDT, This is an Open Access article which permits unrestricted non-commercial use, provided the original work is properly cited

Open  Access

Research Article

Extraction, Characterization and Evaluation of Okara Mucilage

Shankar M. Dhobale*, Shilpa S. Kolhe, Pratiksha P. Darekar, Tanhaji R. Dere, Shubhangi H. Date, Pooja V. Badhe

VJSM's Vishal Institute of Pharmaceutical Education and Research, Ale, Tal-Junnar Dist-Pune 412441

ABSTRACT

Mucilage is the thick, gluey substances produced by nearly all plant and some microorganisms. Okra mucilage is extracted from the plant of the *malvaceae* [*A. esculantus*]. Which is originally from Egypt, but it also in cropped in southern Asia elsewhere for nutritional purposes. Their use as potential reinforcement in polymer composites requires the understanding of their microstructure and mechanical properties. This work investigates the extraction methods, solubility behavior, TLC, loss on drying, ash value, FTIR spectra, surface tension, organoleptic properties. Extracted mucilage is soluble in warm water while insoluble in organic solvents. This can shows that it safely used in dosage form without causing any adverse effect.

Keywords: Okara Mucilage, Pharmaceutical Excipients, Controlled-Release Formulation**Article Info:** Received 27 March 2019; Review Completed 04 May 2019; Accepted 08 May 2019; Available online 15 May 2019

Cite this article as:

Dhobale SM, Kolhe SS, Darekar PP, Dere TR, Date SH, Poja V. Badhe PV, Extraction, Characterization and Evaluation of Okara Mucilage, Journal of Drug Delivery and Therapeutics. 2019; 9(3):325-328
<http://dx.doi.org/10.22270/jddt.v9i3.2665>

*Address for Correspondence:

Shankar M. Dhobale, VJSM's Vishal Institute of Pharmaceutical Education and Research, Ale, Tal-Junnar Dist-Pune 412441

INTRODUCTION

Mucilages are most commonly used adjuvant in pharmaceutical preparations. Plant mucilages are pharmaceutically important polysaccharide with wide range of applications such as thickening gelling agent, binding, disintegrating, suspending, and emulsifying, stabilizing and gelling agents. They have been also used as matrices for sustained and controlled release drugs. Naturally available mucilages are preferred to synthetic materials due to their non toxicity low cost, emollient and non irritating nature. Acacia, tragacanth, gum ghati, gum karaya are popular examples of plant mucilages. As a present paper deals with isolation, phytochemical screening and evaluation of binding properties of *Hibiscus esculentus* mucilage's. As a dose formulators essential to develop cost-effective and less tedious procedures for preparation of sustained release formulations on the industrial scale. The most commonly used method for fabricating drugs in a controlled-release formulation is by incorporating them into a matrix containing a hydrophilic rate controlling natural polymer¹.

Now a day many research are going on for the use of natural occurring biocompatible polymeric material in designing of pharmaceuticals dosage form for oral controlled release administration. Most of the natural gums are biodegradable and nontoxic, which hydrate and swell contact with aqueous media, so these have been used for the preparation of

dosage form. Polysaccharide obtains from plants has been shown to be useful for the construction of drug delivery systems². Regular research is going on in field of use of natural occurring biocompatible polymeric material in designing of dosage form for oral controlled release administration. Natural gums are biodegradable and nontoxic, which hydrate and swell on contact with aqueous media, and these have been for the preparation of dosage form³.

Natural are playing an important role as pharmaceutical excipients. These are easily available biodegradable and having economic. Biocompatibility of these natural polymers promotes their use as in pharmaceutical formulations. Present work used granulation compression technique to prepare tablets. In present study diclofenac sodium a non-steroidal anti-inflammatory drug is selected as model drug [NSAID] with analgesic and antipyretic properties⁴. Diclofenac sodium is used treatment is used in treatment of pain, dysmenorrhea, ocular inflammation, osteoarthritis, rheumatoid arthritis, ankylosing spondylitis, and actinic keratosis.



MATERIAL AND METHODS

Extraction:

Extraction procedure:-okra (a. *Escarlatus*) was obtained from local market of India. Collected okra was carefully washed and dried under shade for 24h, further dried at 30-40°C until constant weight is obtained. Size was reduced through sieve no. #22 and stored in air tight container for further use. Extractions of mucilage include 2 steps⁵.

Step 1:-method used for extraction is soxhlet extraction method. Take a 10gm powder of okra. Pack into the soxhlet apparatus. Then using the solvent extraction starts. The solvents can be a use for the extraction is water. Start the extraction. After the completion perform second step.

Step2:-isolation of mucilage: the obtained extract pour in petriplate. The mucilage was further dried to constant weight at 35-45°C in hot air oven. Hard mucilage cake was grinded and sieved through sieve #22, store in dessicator for further use^{6,7}.



Soxhlet extraction of mucilage

1. Organoleptic evaluation of isolated mucilage: isolated mucilage was characterized for organoleptic properties such as colour, odour, taste, fracture and texture

2. Determination of purity of okra mucilage: to determine purity of extracted mucilage, facts, tannins and amino acids were performed

3. Phytochemical test:

Aqueous extract was mixed molish's reagent followed by addition of sulphuric acid the violet color ring appeared at junction, showing presence of other chemicals.

4. Physicochemical properties of okra mucilage:

A. Loss on drying: the test was carried out according to the procedure described by authors elsewhere. One gram of powder was weighed accurately in a weighing bottle and was dried in a weighing bottle and was dried in a hot air oven at 105°C and the weight was checked at intervals of 10 min, until a constant weight was obtained. The percentage of weight lost by the powder was calculated using equation

$$\text{Loss on drying} = \left(\frac{\text{initial weight} - \text{final weight}}{\text{initial weight}} \right) \times 100$$

B. Ash values: as discussed by authors in previous publication ash values such as total ash, acid insoluble ash and water-soluble ash were determined using equation 1, 2, 3 respectively

$$\text{Total ash value} = \left(\frac{\text{weight of ash}}{\text{weight of polymer}} \right) \times 100$$

$$\text{Acid insoluble ash} = \left(\frac{\text{weight of acid insoluble ash}}{\text{weight of dried powder}} \right) \times 100$$

$$\text{Water soluble ash} = \left(\frac{\text{weight of water soluble ash}}{\text{weight of dried powder}} \right) \times 100$$

Solubility behavior: as already described by authors one part of dry mucilage powder was shaken with different solvents and further solubility was determined

C. pH of mucilage: the mucilage was weighed and dissolved in water separately to get a 1% w/v solution. The pH of solution was determined using digital pH meter as described by previous publication

D. swelling index: as described by authors in previous publication swelling index were calculated as per equation

$$\text{Swelling index} = \left(\frac{\text{final volume} - \text{initial volume}}{\text{final volume}} \right) \times 100$$

E. surface tension: the surface tension of the selected mucilage was determined by drop count method, using a stalagmometer. The surface tension of the polymer has been reported to influence the binding quality of the polymer. Surface tension was calculated as per equation 5.

$$\sigma_{\text{solution}} = \sigma_{\text{water}} \times \frac{M_{\text{(solution)}}}{M_{\text{(water)}}}$$

Where,

Σ = surface tension of solution

Σ = surface tension of water

$M_{\text{(solution)}}$ = weight of water

F. viscosity: as described by authors viscosity of okra

Mucilage was determined using oswald viscometer were

Calculated using following equation;

$$S = \frac{W \times t_{\text{ps}}}{t_{\text{wp}}}$$

Where,

S = viscosity of solution W = viscosity of water

T = time P = density

5. TLC characterization:-TLC is method for analyzing mixtures by separating the compounds in the mixture. It is used to identify the compound and purity by comparing with standard. It is calculated from retention factor^{8,9}.

5. FTIR spectra:-

Take about 1/8" of the solid sample on a microspatula and about 0.25-0.50 teaspoons of KBR. Mix thoroughly in a mortar while grinding with a pestle. If the sample is in the large crystals, grind the sample separately before adding KBR.

Place just enough special to cover bottom in pellet die. Place in press and press at 5000-10000 psi. Check pellet press brochure for details. Carefully remove the pressed sample from die and place in the ftir sample holder. The pressed disc should be nearly clear if properly made. If it is translucent, regrind and repress. Add 1% of mucilage in the 99% of kbr repeat the above procedure¹⁰.

RESULTS AND DISCUSSION

Extraction:

The extraction was carried out using soxhlet extraction and 4 cycles are obtained. Yield was found to be 12.14w/w.

Mucilage obtained was 12.14w/w

Organoleptic evaluation of isolated mucilage:

Sr.no.	Parameters	Obs.
1	Colour	Brownish
2	Odour	None
3	Taste	Characteristic

Phytochemical test:

Sr.no.	Test	Present/absent
1	Carbohydrates	+
2	Hexose sugar	+
3	Monosaccharide's	-
4	Protein	-
5	Fats and oils	-
6	Tannins and phenolic compounds	-
7	Alkaloids	-
8	Amino acids	-
9	Mucilage	+
10	Gums	-

Physicochemical properties of okra mucilage:

Sr.no.	Parameter	Obs.
1	Loss on drying	8.97%
2	Ash value:	
A	Total ash value	6.93%
B	Water soluble	5%
C	Acid insoluble	0.87%
3	Ph of mucilage	7.5
4	Swelling index	28
5	Surface tension	0.0305 joule/m sq
6	Viscosity	62.32 CP

TLC:



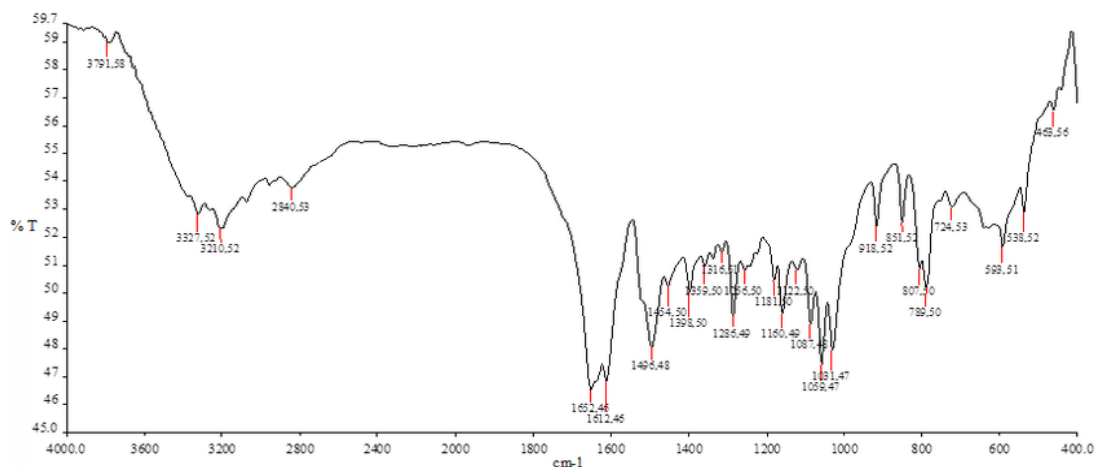
TLC for okra mucilage

R_f value:-

Spot 1:-0.302

Spot 2:-0.794

FTIR:-



FTIR spectra for okra mucilage

Interpretation of FTIR spectra:-

Sr. No.	Wave no. (cm ⁻¹)	Functional group
1	1662	C=O
2	1258	C-O
3	1080	C-C
4	2840	C-H
5	3327	O-H

After extraction and further precipitation by ethyl alcohol the yield of mucilage was 12.14w/w obtained. The isolated sample was subjected to identification; this showed presence of carbohydrates in sample powder. Confirmation of mucilage was done when it gave negative test of tannins, Alkaloids and protein. This can be considered as proof for purity of the isolated mucilage as depicted in table 1.

The results for loss of drying showed value of 8.97%. This indicated that mucilage is hygroscopic in nature and need to be stored in air-tight containers. In solubility behavior of okra mucilage was found to be soluble in warm water, slightly soluble in cold water and insoluble in benzene, glycerine, paraffin. Surface tension of 0.25%w/v. Solution of mucilage was found to be 0.0305 joule/m sq. Other phyto-constituents were absent in the isolated powder, pH of 1% solution was found to be 7.5. Result obtained of okra mucilage and observed that mucilage is brownish colour, odourless, testless, rough and irregular in shape. Ash values were calculated to characterize mucilage; total ash acid insoluble ash and water soluble ash were found 6.93%, 0.87% and 5% respectively. The TLC characterization was also studied and R_f value was found to be 0.302, 0.794. FTIR spectra also obtained. The all data obtained tabulated.

CONCLUSION

Results of evaluated parameters showed that okra derived mucilage can be used as pharmaceutical excipient to formulate solid oral dosage form. It has acceptable pH and organoleptic properties, so can be easily used to formulate various dosage form.

REFERENCES

1. Krishna, L.N.V., P.K. Kulkarni, M. Dixit, D. Lavanya and P.K. Raavi, Brief introduction of natural Gums, mucilages and their applications in novel drug delivery systems. International journal of drug Formulation and research, 2011; 2(6):54-71.
2. Bogoeva-gaceva G, Avella M, Malinconico M, Buzarovska A, Grozdanov A, Gentile G, et al. Natural fiber eco-composites. Polym compos 2007; 28(1):98-107.
3. Reddy N, Yang Y. Natural cellulose fibers from switchgrass with tensile Properties similar to cotton and linen. Biotechnol bioeng 2007; 97(5):1021-7.
4. Paiva MC, Ammar I, Campos AR, Cheikh RB, Cunha AM. Mechanical, Morphological and interfacial characterization. Compos sci technol 2007; 67(6):1132-8.
5. Srivastava P., Malviya R., Kulkarni G. T.: formulation and evaluation of paracetamol Tablets to assess binding property of orange peel pectin. International journal of pharmaceutical Sciences review and research, 2010; 3(1):30-34.
6. Malviya R., Srivastava P, Bansal M., Sharma P. K.: preparation and evaluation of disintegrating Properties of *cucurbita maxima* pulp powder. International journal of pharmaceutical Sciences, 2010; 2:1395-399.
7. Dharmendra S., J.K. Sujata SM, Shweta S. Natural excipients. International Journal of pharmaceutical and biological archives, 2012; 3(5):1028-1034.
8. Hanan, M.A., al-sayad, Nagwa M.H. Rasmy, Ibrahim R.S. Rizk and Amaan E.I. Yousef. Functional properties of some fat replacers and their uses in preparation of reduced-fat Mayonnaise. World general of dairy and food science, 2012; 7(1):109-119.
9. Ogaji I.J., E.I. Nep and J.D. Audu-peter. Advances in natural polymers as pharmaceutical Excipients. Pharmaceutical analytica acta 2011; 3(1):10-15
10. Morkhade D.M., Fulzele S.V., Satturwar P.M. ; Joshi S.B.. Novel matrix forming materials For sustained drug delivery. Indian J. Pharm. Sci. 2006; 68(1):53-58.