

RESEARCH ARTICLE

PREPARATION AND EVALUATION OF MUPIROCIN LOADED POLYMER COMPOSITE FILMS

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ABSTRACT

Polymer composite films were prepared by using chitosan and sodium alginate alone and in combination with aloe vera, with & without glutaraldehyde were prepared by solvent casting method. Mupirocin was incorporated into selected polymeric films. All the polymeric composite films were characterized by IR study suggested that there was no chemical reaction has taken place, only ionic complex was formed. All the films were evaluated for thickness, folding endurance, and tensile strength. The thickness of all the films was uniform as the concentration of polymer was kept constant. The folding endurance suggested good flexibility of the films as propylene glycol was used as a plasticizer. The water vapour penetration suggested that films prepared without cross linker absorbs more moisture as compared to films containing cross linking agent. All the films subjected to tensile strength and in-vitro bio adhesion. The films showed good tensile strength, suggested good mechanical property for handling of dressing film. All the films showed the good bioadhesion, which must require to adhere perfectly over the wound. The presence of cross linking agent decreases the bioadhesion. All the polymer composite films were evaluated for in vitro swelling study. The films showed good swelling in water more than 6 hrs retaining the shape of the films. The addition of cross linking agent decreased the swelling. Selected polymer composite films were evaluated for in vivo wound healing activity. All the polymeric films showed more than 80% reduction in wound contraction. The mupirocin loaded polymeric composite containing aloe vera, shown more than 98% reduction in wound area after 12th day. Hence, from the overall study it can be concluded that polymer composite films of chitosan-alginate containing mupirocin along with aloe vera showed good wound healing and could be used in effective management of all type of wounds.

Keywords: Chitosan; sodium alginate; aloe vera; mupirocin; polymer composite films; wound healing.

INTRODUCTION

According to the Wound Healing Society, a wound is the result of disruption of normal anatomic structure and function. Based on the nature of repair process, wound can be classified as acute or chronic wounds¹. The management of wounds can be done by using different wound dressings which have been developed over the years from the crude applications of plant herbs, animal fat and honey to tissue engineered scaffolds². Recognition of the importance of cleanliness and good aseptic practice in medicine and surgery has lead to improvements in the management of quality of wound management materials³. In the last few decades lot of work have been done towards effective management of wound. In the present scenario of advanced technology, modern wound dressing have been introduced which include hydrocolloid dressing, alginate/chitosan dressing and hydrogel dressing not only as bandage but also in the form of sponges, fiber, films and solution⁴. Chitosan is currently receiving a great deal of attention for medical and pharmaceutical applications due to its beneficial intrinsic properties. Chitosan is a deacetylated chitin derivative containing amino sugar. Chitosan possess many properties that are advantageous for wound dressing, namely bio-compatibility, biodegradability, haemostatic activity, anti-infection and wound acceleration properties⁵. Alginate is another polymer which is bio-degradable and obtained from natural origin having wound healing property and good bio-adhesion which is necessary for more retention over the skin⁶. Prajapati et al worked on polyelectrolyte complex of chitosan alginate for local drug delivery and explain that combination of appropriate drug and chitosan can help to recover topical infections. Polyelectrolyte complex film of chitosan and sodium alginate film can be

used for sustained drug delivery of potent antimicrobial and antifungal drugs by transdermal drug delivery⁷. Aloe vera also known as true or medical aloe is species of succulent plant, which is used as herbal medicine. Aloe vera is used topically and systemically for the treatment of different diseases. Aloe vera could improve wound healing after topical application by the different mechanisms, such as keeping the wound moist, increase epithelial cell migration and more rapid maturation of collagen and reduction in inflammation. Aloe vera has skin hydration effects, anti-microbial and anti-fungal activity⁸. The purpose of applying antibiotics and other antibacterial is mainly to prevent or combat infections especially for diabetic foot ulcers, surgical and accidents wound where the incident of infections can be high due to reduced resistance resulting from extreme trauma⁹. Mupirocin is a topical antimicrobial indicated in the treatment of impetigo and secondary skin infections. Mupirocin is an antibiotic produced from *Pseudomonas fluorescens* and structurally unrelated to any other topical or systemic antibiotic. Commercially it is available in the form of creams and ointments for topical application¹⁰.

Hence in the present work an attempt will be made to prepare and evaluate polymeric composite films of chitosan-alginate and incorporated with mupirocin and aloe vera extracts for the effective management of different types of wounds.

MATERIALS AND METHOD

Chitosan was obtained as gift sample from India Sea Foods, Cochin, Kerala and mupirocin was obtained as gift sample from Glenmark Pharmaceuticals Ltd, Nasik, Pune.

1. Preparation of polymer composite films

The films were prepared by solvent casting method¹¹ as shown in Table 1. Polymeric films of chitosan alone and along with aloe vera was prepared by dispersing specified amount of chitosan dispersed in 3% v/v lactic acid in water, agitated for 1 h and add aloe vera into the chitosan dispersion followed by addition of glutaraldehyde with gentle stirring, then add the propylene glycol. The mixed solution was left to stand until air bubbles had disappeared, then poured onto a petri dish and allowed to air-dry at 40°C for 24 h. To prepare chitosan-alginate composite

films, sodium alginate solution was prepared by dissolving the alginate alone and along with aloe vera in the deionised water followed by chitosan solution which was prepared by dispersion of the chitosan in to 3% v/v lactic acid. Acetone was added to alginate solution. Finally the dispersed solution of the chitosan was added to sodium alginate solution drop wise under rapid agitation, then glutaraldehyde (0.03% v/v) was added followed by propylene glycol. The casting and drying of the films was done similar as explained above.⁸ Similarly mupirocin incorporated films were prepared by adding mupirocin in alginate solution and followed the same procedure as explained above.

Table 1: Formulation of polymer composite films

Ingredients (% w/v)	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10	F11	F12
Chitosan	3	3	2	2	1.5	1.5	1	1	1.5	1.5	1	1
Sodium alginate	--	--	--	--	1.5	1.5	1	1	1.5	1.5	1	1
Aloe vera	--	--	1	1	--	--	1	1	--	--	1	1
Mupirocin	--	--	--	--	--	--	--	--	2	2	2	2
Glutaraldehyde	--	0.03	--	0.03	--	0.03	--	0.03	--	0.03	--	0.03
Propylene Glycol	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5

2. Characterization of polymer composite films

2.1 Fourier-transformation infrared spectroscopy (FTIR)

The drug-polymer and polymer-polymer interaction were studied using FTIR spectrometer (Perkin-Elmer (spectrum-100) Japan) by taking 2% w/w of the sample with respect to potassium bromide disc, ground in to a fine powder and then compressed into a discs in a hydraulic press. Each disc was scanned 16 times at 2 mm/sec at a resolution of 4 cm⁻¹ using adopization. The characteristics peaks were recorded.

3. Evaluation of polymer composite films

3.1 Thickness⁷

The thickness of film influence the time required to absorb the polymer into the body. To determine the uniformity in thickness of film and change in thickness film after drug loading, it was measured for each film using screw gauge at three different sites of the film and the mean was calculated.

3.2 Folding endurance

It was determined to find the flexibility of film which is needed to handle the film easily and for comfortable, secured application of film on the wound. It was determined by repeatedly folding one film at same place till it breaks or folded up to 300 times manually. The number of times of film could be folded at the same place without breaking give the value of folding endurance¹².

3.3 Swelling index

Weighed pieces (1cm²) of film were immersed in distilled water, then soaked films were removed from the medium at predetermined time, blotted to remove excess liquid and weighed immediately¹². The swelling index was calculated

$$\text{as } \% S = \frac{w_2 - w_1}{w_1} \times 100.$$

Where **w1** and **w2** are the weight of the film before and after immersion in the medium.

3.4 Water vapour penetration

To measure the water vapour penetration, the films were cut and placed on top of open 2.5- cm bottles containing 5g of silica gel and held in place with a screw lid (test area: 4.9 cm²). The bottles were conditioned in desiccators containing silica gel for 12 h. The bottles were then placed in desiccators containing a saturated solution of sodium chloride at 75 % RH. The equilibrium vapour penetration was determined by weighing the bottles at 0, 12, 24 and 48 hours respectively¹².

3.5 Tensile strength¹⁴

The mechanical properties of films were evaluated using a texture analyzer (Instron Universal Model) equipped with a 500 gm load cell¹³. Film strip in 10 mm X 10 mm of dimension and free from air bubbles or physical imperfections, was held between two clamps positioned at a distance of 1 cm. During measurement, the film was pulled by top clamp at a rate of 10mm/minutes. The force and elongation were measured when the films broke. The tensile strength was calculated as,

$$\text{Tensile strength (kg/mm}^2\text{)} = \frac{\text{Breaking force (kg)}}{\text{Cross sectional area of sample (mm}^2\text{)}}$$

3.6 *Invitro* bioadhesion study

The bioadhesive property of the film was performed using an in-house pulley system instrument¹³. The proximal portion of a chicken pouch was used to represent the mucous-like texture of a fresh wound. The freshly slaughtered chicken pouch washed with physiological saline at 4°C and attached to a platform (test area: 4.9 cm²). A prewetted film was placed a top the chicken pouch and held under 100 g weight for 2 min, with the other side of the weight connected to a pulley system. The water was added to a container attached to the pulley system until the

film was detached. The weight of water needed to detach the film from the intestine was recorded.

3.7 Wound healing activity by excision model

Male Wister albino rats (150-250 g) in a total 8 groups of each having three animals were used after obtaining approval from institutional animal ethics committee (No.346/CPCSEA) by excision wound model^{13,14,15}. Animals were housed under standard conditions of laboratory. Excision wound was inflicted under light ether anesthesia by excising a circular piece of (20 mm²) of full thickness skin from the dorsal interscapular region. Selected films were adhering over the wound and marketed povidone-iodine was used as a standard. Wound contraction was monitored by measuring wound area planimetrically, every alternate day till the wound was completely healed. Wound contraction was calculated as percent reduction in wound.

RESULTS AND DISCUSSION

In the present study natural based polymer composite films were prepared for the effective management of wound. To

enhance the therapeutic efficacy of the film antimicrobial agents like mupirocin and wound healing accelerators like aloe vera was incorporated in the film. Chitosan and sodium alginate was used as main polymers alone and along with aloe vera was followed by glutaraldehyde as cross linking agent and propylene glycol as a plasticizer. All the films were prepared by solvent casting method. The characterization of prepared polymer composite films was studied by FTIR (Fourier's transform infrared spectroscopy) and the obtained spectra are represented in Figure 1. The IR study confirms that mupirocin remains intact in the polymer composite and also the IR spectrum of chitosan with sodium alginate results in formation of ionic complexes as both the polymers are polyionic. Chitosan contains cationic amino group which interact ionically with anionic carboxylic group of alginate and further glutaraldehyde crosslinked within chitosan to form strong ionic complex.

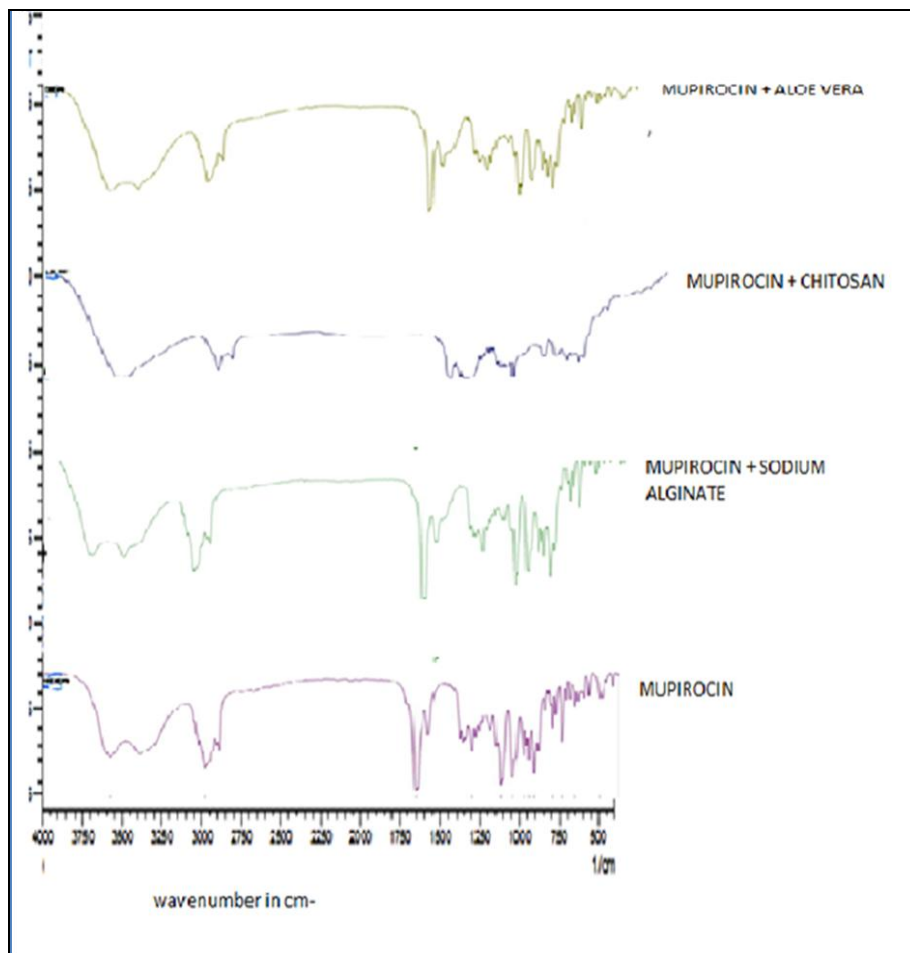


Figure 1: FTIR spectra of mupirocin loaded polymer composite films

All the polymer composite films were evaluated for different parameters and results are tabulated in Table 2. The thickness was varied from 0.6 to 1.0 mm as the total polymer concentration was kept constant (3% w/w). Chitosan-alginate composite films along with aloe vera without glutaraldehyde showed the thickness 0.6 ± 0.0577 mm. The thickness of the films was increased when the cross linker was added. The water vapour penetration of films had showed the percent increase in weight as shown in Table 2,

from $26.22 \pm 0.66\%$ to $77.94 \pm 0.89\%$. The water vapour permeability of the films was decrease with the increase in the concentration of the chitosan in the films. Thickness and the water vapour permeability inversely related, with thinnest films showed the highest water vapour permeability and vice-versa. The water vapour permeability of the mupirocin incorporated films showed from $28.44 \pm 0.68\%$ to $65.23 \pm 0.99\%$.

Table 2: Evaluation parameters of polymer composite films

FC	Thickness (mm)	Folding endurance	Water Vapor penetration (%)	Tensile strength (Kg/mm ²)	Maximum swelling index (%)	In-vitro bioadhesion gm/cm ²
F1	0.8 ± 0.05	235 ± 5.21	61.61 ± 1.22	0.122 ± 0.01	2542.85 ± 12.22	21.28 ± 0.58
F2	1 ± 0.06	258 ± 2.86	26.22 ± 0.66	0.296 ± 0.02	1114.28 ± 8.22	11.59 ± 0.64
F3	0.7 ± 0.05	177 ± 2.52	77.94 ± 0.89	0.128 ± 0.01	1811.11 ± 9.05	19.24 ± 0.44
F4	0.8 ± 0.07	210 ± 2.05	32.80 ± 0.45	0.266 ± 0.01	922.22 ± 7.94	12.47 ± 0.33
F5	0.7 ± 0.07	194 ± 4.51	77.66 ± 0.42	0.258 ± 0.02	2900 ± 13.12	26.98 ± 0.39
F6	0.9 ± 0.05	219 ± 3.25	57.33 ± 0.88	0.294 ± 0.04	1144.44 ± 8.99	25.95 ± 0.54
F7	0.6 ± 0.05	208 ± 2.25	68.42 ± 0.58	0.244 ± 0.08	2340 ± 12.58	26.12 ± 0.45
F8	0.9 ± 0.06	222 ± 2.51	48.44 ± 0.64	0.276 ± 0.06	1266.66 ± 13.52	24.25 ± 0.77
F9	0.8 ± 0.05	195 ± 3.52	65.23 ± 0.99	0.245 ± 0.02	911.70 ± 7.42	22.12 ± 0.59
F10	1 ± 0.03	214 ± 8.14	30.0 ± 1.01	0.296 ± 0.02	724.42 ± 7.92	21.41 ± 0.12
F11	0.7 ± 0.05	211 ± 3.61	47.35 ± 0.68	0.242 ± 0.06	965.29 ± 7.35	22.66 ± 0.31
F12	0.9 ± 0.05	210 ± 5.21	28.44 ± 0.65	0.278 ± 0.05	850 ± 6.75	20.25 ± 0.78

(FC= Formulation Cod Note: Values in parenthesis are standard deviation (\pm SD); n=3)

The tensile strength of polymer composite films expressed in Kg/mm² as depicted in Table 2. Tensile strength represents the mechanical property of the films for the safe handling of dressing. Tensile strength depends upon the polymer, presence of plasticizer and addition of glutaraldehyde. The tensile strength of polymer composite films was 0.122 ± 0.0115 to 0.296 ± 0.0130 Kg/mm². The results suggested that films containing glutaraldehyde showed decrease in tensile strength due to more flexibility of films as compared to films did not contained glutaraldehyde. The addition of mupirocin in the chitosan-alginate composite films did not affect the tensile strength.

All the prepared polymer composite films were evaluated for bioadhesion and results are depicted in Table 2. The

bioadhesion is an important property required to adhere to the wound for effective healing of the wound. The bioadhesion is affected by nature and molecular weight of polymer used, presence of cross linker, contact time and the degree of swelling of polymer. Chitosan and alginates are highly hydrophilic cationic and anionic polymers and hydrated to form slightly adhesive mucilage. The bioadhesion of polymer composite films was in the range of 11.59 ± 0.69 to 26.98 ± 0.39 . The bioadhesion was decreased by the addition of glutaraldehyde. The addition of mupirocin in the chitosan-alginate composite films did not affect the bio-adhesion. All the prepared composite films were evaluated for swelling index, and average swelling index is depicted in Table 2 and also the data up to 9 h is represented in figure 2 & 3.

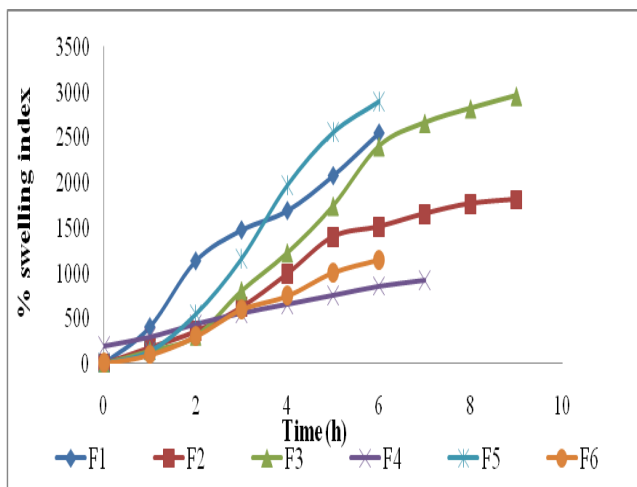


Figure 2: Swelling study of polymer composite films with and without glutaraldehyde along with aloe vera

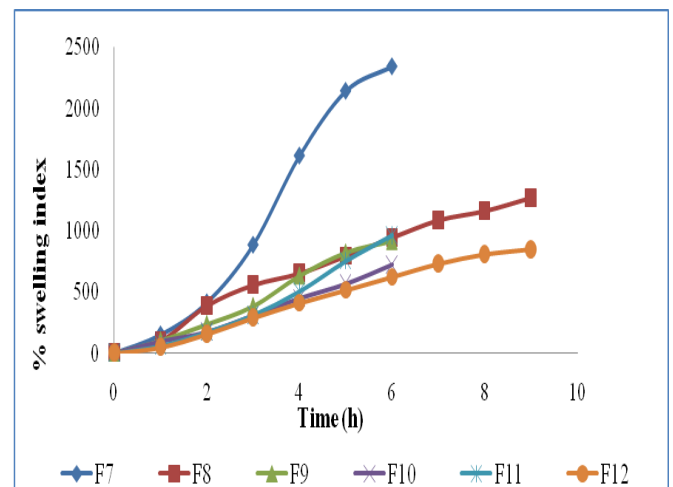


Figure 3: Swelling study of mupirocin incorporated polymer composite films with and without glutaraldehyde along with aloe vera

The swelling behavior is related with the mucoadhesion and depends on the nature and viscosity of polymer, media used and the presence of cross linker. The degree of swelling increased as the time passes and at certain time some of the formulations loose the integrity which did contained glutaraldehyde. In the chitosan composite films, chitosan alone showed rapid swelling up to 6 h and could not hold the shape, whereas addition of aloe vera prolonged the swelling up to 9 h. The percent swelling of polymer composite films was from 911.70 ± 7.42 to 2900 ± 13.12 (F1-F8). Mupirocin incorporated films showed the swelling from 724.42 ± 7.92 to 965.29 ± 7.35 (F9-F12). Addition of glutaraldehyde

decreased the swelling due to formation of more rigid network. Wound healing is a process by which damaged tissue is restored as closely as possible to its normal state. Wound contraction is the process of shrinkage of area of the wound. It mainly depends on the repairing ability of tissue, which may be reduced due to infection. Chitosan-alginate composite films containing aloe vera and mupirocin with and without glutaraldehyde were subjected to in vivo wound healing activity by method of excision model in albino rats. The results are expressed in percent contraction in wound area as depicted in Table 3 & 4 and also represented in bar graph in figure 4 and 5.

Table 3: Wound healing data of polymer composite films formulated with and without glutaraldehyde along with aloe vera

Post-wounding days	Wound area (mm)					
	Control	Standard	F5	F6	F7	F8
0	20.03 ± 0.81 (0%)	20.35 ± 0.76 (0%)	20.06 ± 0.42 (0%)	19.92 ± 0.93 (0%)	19.95 ± 0.56 (0%)	19.98 ± 0.78 (0%)
2	18.06 ± 0.77 (9.7%)	18.02 ± 0.52 (9.9%)	17.52 ± 0.63 (12.4%)	17.12 ± 0.84 (19.4%)	16.65 ± 0.92 (16.75%)	16.12 ± 0.06 (19.4%)
4	16.09 ± 0.32 (19.6%)	15.69 ± 0.53 (21.55%)	15.5 ± 0.29 (22.2%)	18.24 ± 0.25 (24.4%)	13.95 ± 0.69 (30.25%)	13.25 ± 0.55 (33.75%)
6	15.03 ± 0.88 (24.55%)	13.46 ± 1.20 (32.7%)	12.98 ± 0.95 (35.1%)	12.22 ± 1.29 (39%)	6.8 ± 0.25 (67.5%)	6.12 ± 0.99 (69.4%)
8	14.07 ± 1.04 (29.65%)	12.5 ± 0.5 (37.5%)	6.98 ± 0.46 (65.1%)	6.25 ± 0.25 (68.25%)	3.25 ± 0.32 (83.75%)	3.06 ± 0.84 (84.7%)
10	13.04 ± 0.62 (34.85%)	9.35 ± 0.94 (53.25%)	3.21 ± 1.25 (83.95%)	3.12 ± 0.15 (84.14%)	1.75 ± 0.57 (91.25%)	1.53 ± 0.28 (92.35%)
12	10.09 ± 0.40 (49.55%)	4.04 ± 0.38 (79.55%)	1.5 ± 0.05 (92.5%)	1.4 ± 0.84 (93%)	0.87 ± 0.46 (95.65%)	0.76 ± 0.24 (96.5%)

(Values are mean ± SD of three animals in each group. Numbers in parenthesis indicates the % wound contraction)

Table 4: Wound healing data of mupirocin loaded chitosan-alginate composite films formulated with and without glutaraldehyde along with aloe vera

Post-wounding days	Wound area (mm)					
	Control	Standard	F9	F10	F11	F12
0	20.03 ± 0.81 (0%)	20.35 ± 0.76 (0%)	20.02 ± 0.79 (0%)	19.98 ± 0.32 (0%)	19.98 ± 0.65 (0%)	19.25 ± 0.65 (0%)
2	18.06 ± 0.77 (9.7%)	18.02 ± 0.52 (9.9%)	17.14 ± 0.93 (14.13%)	17.01 ± 0.49 (14.95%)	15.92 ± 0.77 (20.4%)	15.95 ± 0.24 (20.25%)
4	16.09 ± 0.32 (19.6%)	15.69 ± 0.53 (21.55%)	14.92 ± 1.26 (25.4%)	14.91 ± 0.86 (25.45%)	13.13 ± 1.11 (34.35%)	12.92 ± 0.14 (35.25%)
6	15.03 ± 0.88 (24.55%)	13.46 ± 1.20 (32.7%)	12.25 ± 0.22 (38.75%)	12.01 ± 0.69 (39.45%)	6.14 ± 0.28 (69.3%)	6 ± 0.29 (70%)
8	14.07 ± 1.04 (29.65%)	12.5 ± 0.5 (37.5%)	6.25 ± 0.38 (68.75%)	5.84 ± 0.26 (70.75%)	2.92 ± 0.29 (85.4%)	2.98 ± 0.94 (85%)
10	13.04 ± 0.62 (34.85%)	9.35 ± 0.94 (53.25%)	3.14 ± 1.43 (84.3%)	2.95 ± 0.82 (89.25%)	1.4 ± 0.22 (93%)	1.32 ± 0.46 (93.4%)
12	10.09 ± 0.40 (49.55%)	4.04 ± 0.38 (79.55%)	1.1 ± 0.21 (94.5%)	1.25 ± 0.28 (93.75%)	0.22 ± 0.06 (98.9%)	0.12 ± 0.04 (99.4%)

(Values are mean ± SD of three animals in each group. Numbers in parenthesis indicates the % wound contraction)

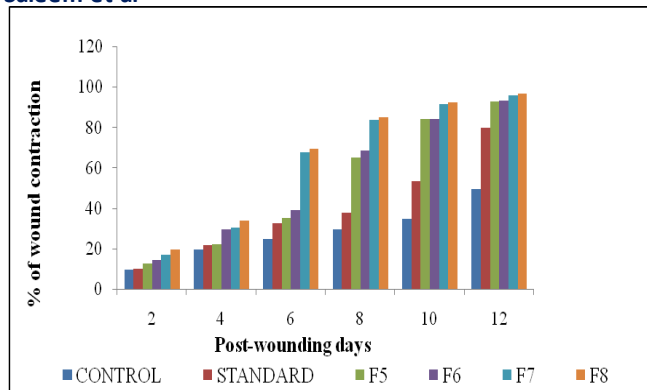


Figure 4: Comparative % wound healing of control and standard with polymer composite films formulated with and without glutaraldehyde containing aloe vera

All the plane chitosan composite films without glutaraldehyde showed increase in % wound contraction as compared to control and standard at 12th day. Control showed 49.55%, standard 79.55% where as F5 showed 92.5% and F7 95.65%. The chitosan-alginate composite films prepared with glutaraldehyde showed percentage of wound contraction as F6 93% and F8 96.5%. The results of addition of glutaraldehyde in the chitosan-alginate did not show much deviation as compared to films without glutaraldehyde. Aloe vera is ancient herbal medicine used as

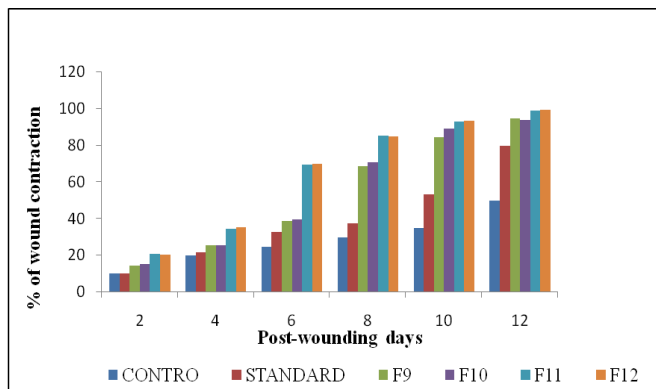
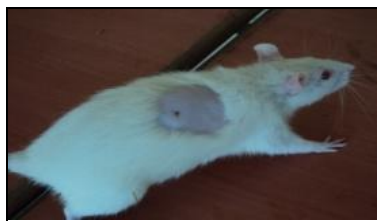


Figure 5: Comparative % wound healing of control and standard with mupirocin incorporated polymer composite films formulated with and without glutaraldehyde containing aloe vera

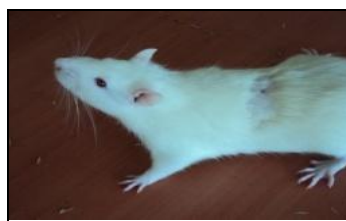


Figure 6: Completely healed wound after 12th day for mupirocin loaded polymer composite films F11 & F12

CONCLUSION

The chitosan polymer composite films containing glutaraldehyde as crosslinker have uniform thickness with good flexibility, good tensile strength, absorb more moisture with excellent bioadhesion and good swelling. All the polymeric films showed more than 80% reduction in wound contraction. The mupirocin loaded polymeric composite containing aloe vera, showed more than 98% of reduction in wound area after 12th day. Hence it can be concluded that mupirocin incorporated chitosan-alginate films along with aloe vera containing glutaraldehyde may be excellent new dressing for wound occlusion/tissue repairing and could be

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