RESEARCH ARTICLE

TRANSUNGUAL PERMEATION OF THE VORICONAZOLE NAIL LACQUER AGAINST *TRICHOPHYTON RUBRUM**Tandel Amruta¹, Agrawal Surendra², Wankhede Shaijesh³¹Department of Pharmacology, Narsee Monjee Institute of Management Studies, School of Pharmacy & Technology Management, Shirpur - 425405, Maharashtra, India.²Department of Pharmaceutics, Narsee Monjee Institute of Management Studies, School of Pharmacy & Technology Management, Shirpur - 425405, Maharashtra, India.³Department of Anatomy, Physiology and Pharmacology, Oriental college of Pharmacy, Navi Mumbai, Maharashtra, India*Corresponding author's Email: amruta_tandel@hotmail.com Ph: 91-9967835838, 91-9967522305,

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ABSTRACT

The oral therapies encounter side effects and topical therapies for nail diseases are limited by poor permeability of nail plate. An optimal penetration enhancer would improve drug delivery through nail plate facilitating new possibilities for treating neighboring target sites if the systemic circulation is reached. The purpose of study was to determine amount of voriconazole permeating through the nail plate from the nail lacquer formulation containing permeation enhancer and to demonstrate its susceptibility to the onychomycotic dermatophyte. The permeability studies were performed on avulsed human cadaver nail plates using modified Franz diffusion apparatus containing phosphate buffer saline in the acceptor chamber. The amount of Voriconazole permeated was quantitatively determined by the UV spectrophotometer. The amount of Voriconazole remaining in the nail after permeation had been retrieved by milling test. The addition of thioglycolic acid, 5% improved the permeability of the drug by 0.7 as the enhancement factor. The formulation followed zero order kinetics and Korsmeyer-Peppas mechanism of drug release on analyzing on DDSolver. The voriconazole flux of 5592.1 µg/cm/min and 596.75 µg/cm/min had been observed at 24 and 48 hours aliquots respectively for the nail lacquer containing an enhancer. Significantly higher permeation rates were achieved in the presence of an enhancer. The inhibitory zones for the voriconazole transungual nail lacquer were evaluated by testing against the dermatophyte, *Trichophyton rubrum* on Sabouraud's Dextrose Agar. The effective zones of inhibitions suggested Voriconazole nail lacquer exhibiting moderate antifungal activity for the onychomycosis treatment.

Key words: Onychomycosis, *Trichophyton rubrum*, Thioglycolic acid, Zero order kinetics, Korsmeyer-Peppas model

INTRODUCTION

Over the years, the importance of nail permeability to topical therapeutics has been realized, primarily in relation to the treatment of onychomycosis; a fungal infection of fingernails and toenails which affects approximately 19% of the world population and is responsible for approximately 50% of all nail disorders¹. Topical therapy is highly desirable because of its non-invasiveness, ability to target drugs to the site of action, minimizing systemic adverse effects and improving patient compliance. Recent advances in transungual delivery technology have led to the introduction of antifungal nail lacquers. Potent and latest second generation triazole antifungal agent Voriconazole with its lipophilic nature fulfils the requirement of an effective drug. Voriconazole [(2R,3S)-2-(2,4-difluorophenyl)-3-(5-fluoro-4-pyrimidinyl)-1-(1H-1,2,4-triazol-1-yl)-2-butanol] is derived from the structure of fluconazole by replacement of one triazole moiety by a fluoropyrimidine group and alpha methylation is a new broad spectrum antifungal agent². It is used in the treatment of superficial and systemic fungal infections caused by *Aspergillus*, *Fusarium* species, *Cladophialophora bantiana*, *Rhizopus arrhizus*, *Scedosporium apiospermum*, *Wangiella dermatitidis*, dimorphic fungi and *Trichophyton rubrum*, *Trichophyton mentagrophytes* & *Candida* species³.

The human nail plate is a much more complex structure than it looks at the first sight. It protects the nail bed, the part directly under the nail plate filled with blood vessels; and the nail matrix, the part at the proximal ventral surface of the

nail responsible for cell's proliferation and nail growth. Although thin, the nail plate has 80-90 layers of dead cells and mainly consists of keratins; 4/5 is hard hair-type keratin and 1/5 is soft skin-type keratin¹; and is mainly associated with stem cell function⁴.

The present work investigated the permeability of the antifungal drug, Voriconazole through the human nail plate from the nail lacquer formulation with and without a penetration enhancer, thioglycolic acid. The relation between the Voriconazole penetrated in to the nail, which forms a reservoir in the nail was investigated after the permeation experiment followed by the drug release kinetics and mechanisms. Finally the formulations were discussed in respect to their enhancement factors by estimating the zone of inhibition against the dermatophyte, *Trichophyton rubrum*.

MATERIALS AND METHODS

Materials

Voriconazole was obtained as a gift sample from Cipla Pvt. Ltd., Mumbai, India; which was chosen as the model antifungal agent. Cellulose nitrate RS, thioglycolic acid, potassium dihydrogen phosphate, and disodium hydrogen phosphate were purchased from Qualigens Fischer Scientific. Aqueous solutions were prepared using deionised water supplied by Millipore water purification system, France. The pH was analyzed using the pH meter (Lab India)

and the permeation aliquots were quantitated using UV spectrophotometer lambda-25 (Perkin Elmer).

Collection, evaluation and characterization of nail samples

Collection of nail samples

Human cadaver nail samples were collected from human corpses at the Mahatma Gandhi Missions Medical Centre and Research Institute, Aurangabad, Maharashtra, India and Rajiv Gandhi Medical College & Chhatrapati Shivaji Maharaj Hospital Mumbai, Maharashtra. Two week old corpses, which have been used in anatomy courses, were filled with 3% solution of formaldehyde. Nails were wet, mostly soft, and strongly bound to the surrounding tissue. By removing the skin, nail edges were liberated and then the nail bed was pressed down along the whole nail plate by easily placing forceps between the ventral nail plate and the nail bed, moving in the direction of nail matrix. Using claws, the nail plate was uprooted. This technique ensured fast collection of the whole nail plates, without breaking them. Only healthy nail plates were used in this study. It is suggested that the permeability through healthy and fungal nail plates is not significantly different. Thus, the fungal nail permeability can be estimated from healthy nail permeability data ¹.

Evaluation of nail samples

Since nail samples were from two week old corpses, possible interaction of formaldehyde might have occurred. The nail plates were washed at room temperature using clean mild liquid detergent containing sodium lauryl sulphate (SLS) and distilled water. These were further dried to constant weight at 45°C. For each nail sample, the dry weight and thickness were measured at three points with a vernier caliper [Mitutoyo] and averaged for each nail ².

Characterization of nail samples

Nail samples were left over night for equilibration at open air and room temperature. On the next day, weight of the whole nail and thickness (Vernier caliper, Mitutoyo) were noted. Samples were placed in PBS solution for 60 min, in order to achieve maximal hydration. The wet nails were mounted in Franz diffusion cells containing PBS in the acceptor chamber.

Preparation and characterization of the formulations

The formulations were made by the standard protocols for the nail lacquer formulation. The solvent system of isopropyl alcohol, butyl acetate, ethyl acetate and n-butyl alcohol was chosen with Cellulose nitrate RS as the film forming agent. Five different formulation batches containing differing concentration of Cellulose nitrate RS [5%, 10%, 15%, 20% and 25%] were made and evaluated for pH (pH meter, Lab India), viscosity, hardness, drying rate, water resistance and stability analysis (Stability Chamber, Thermolab). The selected formulation batches were assessed for increase in the penetration by adding thioglycolic acid (5%) in the formulation, and one extra batch devoid of it ⁵⁻⁷.

Permeation studies

Permeation studies were performed using Franz diffusion cells with a diffusion area of 0.785 cm². The acceptor chamber was filled with 15 ml of Phosphate buffer saline [pH 7.4] solution and constantly stirred with a 3 mm

magnetic stir bar at 400 rpm. The water jacket retained a temperature of 32 ± 1°C. The formulation (400 µl) was applied on the nail surface. Intermittent samples of 4 ml were drawn from the receiver compartment till 72 h and the acceptor chamber was refilled each time with equal amount of Phosphate buffer saline solution, which was kept in a dark place at room temperature. An occlusive effect was attained throughout the experiment and the whole set up was protected from daily light by cardboard. The amount of Voriconazole in the collected samples was determined by UV spectrophotometer at 254nm. The flux is defined as the amount of drug permeated through the nail per time and unit area; which was extrapolated to account for the permeability coefficient (cm/s). The enhancement factor was determined representing as the ratio of permeability coefficient of the formulation containing an enhancer to the permeability coefficient of the formulation devoid of an enhancer ^{2,5,8}.

Milling test

After the permeation experiment, the nails were milled in order to detect the amount of Voriconazole remained in the film and nail plate. The milling test was performed by pulverizing the mounted nail plate using a filer [Soligen, Germany]. After an equilibration time at room temperature, the pulverized nail was suspended in the phosphate buffer saline (pH 7.5) and was shaken for 5 min ^{1,7,8}. Immediately after centrifugation for 5 min at 10,000 rpm, supernatants were diluted with acetonitrile and measured by UV spectrophotometer at 254nm.

Antifungal susceptibility test

Preparation of Sabouraud dextrose agar (SDA)

Briefly, 23.5 g of the powdered SDA base with Emmon's modification [Himedia Lab Pvt. Ltd.] was suspended in 500 ml of distilled water. The mixture was then heated to boiling point whilst stirring to dissolve the powdered agar completely. The agar solution was then sterilised in an autoclave for 15 minutes at 121°C [15 lbs]. The pH was maintained at 7 ± 0.2.

Inoculum suspension for susceptibility testing

The sterilized SDA was poured in to sterile tubes and slants were made for subculturing. After the growth period of 7 – 8 days, the *T.rubrum* species were isolated and each isolate was suspended in 5ml of sterile distilled water. For the Kirby Bauer's disk diffusion method, an inoculum concentration of 1×10⁶ - 5×10⁶ colony forming units/ml [cfu/ml] of the *Trichophyton rubrum* dermatophyte were spread on the sterile plates ⁹.

Kirby Bauer's Disk diffusion test

The assays were performed with the experimental method of Kirby Bauer in three replications. After inoculation, a circular paper disc of 5 mm was cut and sterilized. The discs were then applied with the selected optimized nail lacquer (400 µl) and dried, and were placed on the surface of the SDA and pressed lightly to ensure good contact ⁹. The plates were incubated at 27°C and the zones of inhibition were measured in millimeters using vernier caliper [Mitutoyo]. The interpretative range of standard zone was adopted from Ontengco *et al.* (1992).

The Voriconazole on receipt was characterized with respect to its melting point [Micro controller based melting point apparatus, Chemiline] which accounted in the range of 127°C - 130°C. The IR scan [Hydraulic pellet press KP, Kimaya Engg.] and wavelength analysis [UV

spectrophotometer lambda-25, Perkin Elmer] revealed the strong presence of -OH bond around the frequency range of 3200 cm^{-1} while that around 1351 cm^{-1} depicted the presence of C-N stretch which can be seen in the chemical structure of Voriconazole. The peaks around the frequency range of 3000 cm^{-1} showed the presence of aromatic rings and the wavelength of 254 nm provided the Voriconazole peak.

Figure 1 The effect of hydration on the human nail plates and nail clippings

The nail plates as well as the nail clippings [fig. 1] were found to equally accept water in the hydration studies^{1, 10}. The average weight increase of ten nail plates after 24 hours was determined to be $45.84 \pm 1.24\%$. The difference in weight between wet, i.e., for 60 min immersed nails in PBS, and dry nails was expressed as percentage of weight increase. After the immersion time of 1 h, detected weight increase was found to be 18.34% with a standard deviation of $\pm 0.50\%$. The weight of the dry nails before and 24 hours

after the experiments were also compared. Slight increase of weight was detected in dry samples measured after the permeation studies, with an average of 1.25%. All the formulations showed pH values in the range of 5 – 7.5 and revealed Newtonian flow behavior [Brookefield DV-II + Pro viscometer] except F1 and F2 formulation batches⁸ as shown in the fig. 2. The formulations were found to be stable till thirty days at 40°C, 75% RH [Stability chamber Th-400G, Thermolab].

Figure 2 The effect of shear stress on the viscosity of the nail lacquer formulations; F1, F2, F3, F4 and F5

The *Trichophyton rubrum* seeded SDA plates showed the zones of inhibitions for the Voriconazole nail lacquer when compared with the control nail lacquer formulation containing the nail lacquer base only. The lacquer paper discs of 5 mm applied with 400 μl (8mg) of the formulation inhibited the growth of the *Trichophyton rubrum* colonies while the growth was seen around the control disc. The

observations showed the zone of inhibition of about 14 mm and 16 mm for the Voriconazole nail lacquer without an enhancer and one with an enhancer respectively. No zones of inhibition were seen for the control disc which contained the nail lacquer base only.

The nail permeation decrease after 48 hours can be attributed to the reapplication of the lacquer containing the Voriconazole and thus the fungal dermatophyte inhibitory activity can be achieved. The reapplication of the

formulation would maintain the resistivity to these species as the 400 μ l of the disc containing 8 mg of the Voriconazole gave moderate zones according to Segismundo *et al.*, 2010.

DISCUSSION

Based on the preliminary parameters [Table 1], the formulation 1 and formulation 2 were rejected for the permeation studies as they lacked the Newtonian viscosity.

Table I Preliminary evaluation parameters for the five nail lacquer formulation batches containing varying concentration of Cellulose nitrate RS

Formulations Parameters	F1	F2	F3	F4	F5
Color, clarity	Light pale yellow, clear				
Non-volatile content	95.91 % - 97.91 %				
Drying rate	87 s -111 s				
Smoothness	Lumps at intervals	Smooth, lumps at intervals	Smooth, evenly dispersed		
Water resistance	86.27 %	88.00 %	95.65 %	97.91 %	97.91 %
Hardness	Fair	Fair	Very good	Excellent	Excellent
Viscosity	Considerable decrease in viscosity		Slight decrease on shear application		
Stability	Found to be stable at 40°C and 75 % RH till 30 days

Figure 3 The permeation profiles of the voriconazole nail lacquer formulation 3 containing 15% of Cellulose nitrate RS

Figure 4 The permeation profiles of the voriconazole nail lacquer

formulation 4 containing 20% of Cellulose nitrate RS

Figure 5 The permeation profiles of the voriconazole nail lacquer

formulation 5 containing 25% of Cellulose nitrate RS

The amount of Voriconazole permeated through the nail plate [fig. 3, fig. 4 and fig. 5] during *ex vivo* permeation study of the applied formulations shows the extent of nail penetration of Voriconazole by the penetration enhancer in the selected three formulations. Addition of thioglycolic acid improved the permeability of the Voriconazole to approximately about 0.7 as an enhancement factor in all the three formulations. Formulation 3 was found to release the drug with higher rate than the remaining two formulations [fig. 6]; and was optimized from the study.

The formulation 3 and formulation 4 showed less release of drug as compared to formulation 1 in the 72 hour permeation study. No correlation between permeability rate and nail weight was found before experiment, so it can be suggested that the main influence on the permeability rate was derived from the formulation. The effect of thioglycolic acid was attributed to its small molecular weight and damage caused on the keratin network and decrease in lipid content in the dorsal nail layer; this act which loosened the nail structure, allowing Voriconazole to penetrate easier. Thioglycolic acid showed the best enhancing potential and proved a good solvent for lipophilic drugs^{5,8}. The higher levels of Voriconazole reservoirs were seen in the nails with the formulation devoid of an enhancer, thioglycolic acid. This was the same order in which the permeability rate decreased as shown in fig. 7. The logical conclusion was that if more Voriconazole permeated the nail, more Voriconazole was inside the nail after the experiment and thus one more proof for the decrease in the rate of permeation after a certain lag time. Correlation between enhancement factor and remaining Voriconazole in the nail is shown in the fig. 8.

Figure 6 Comparison of the permeation profiles of the voriconazole nail lacquer formulations F3, F4 and F5 containing thioglycolic acid 5% as penetration enhancer

Figure 7 Profiles of the remaining amount of the nail lacquer present in the nail after the permeation studies of the three formulations, each with and without an enhancer

Figure 8 Enhancement factors calculated with respect to permeation study, where the formulation 3 shows the highest enhancement factor of 0.73

The model selection criterias [MSC], correlation coefficient [R], Mean Square Error [MSE] and Akaike's Information Criteria [AIC] considered for the analysis of the release kinetics for the optimized formulation 3 containing 15% of

Cellulose nitrate RS showed the formulation 3 to follow Korsmeyer-Peppas model and zero order kinetics for both, the lacquer without an enhancer and with an enhancer [Table 2].

Table II Drug release mechanistic models and kinetics for the optimized formulation batch 3

Drug release models	Control				L				Le			
	R _{obs}	MSE	MSC	AIC	R _{obs}	MSE	MSC	AIC	R _{obs}	MSE	MSC	AIC
Z	0.961	951.262	2.366	144.452	0.951	38413.046	1.969	199.928	0.950	929330.671	1.764	247.719
F	0.752	9796.083	0.034	179.43	0.490	513666.786	-0.624	238.825	0.343	10433806.069	-0.653	283.994
H	0.947	3533.736	1.0540	164.137	0.917	121892.526	0.814	217.249	0.902	2754724.480	0.678	264.018
K	0.960	1017.35	2.239	146.34	0.947	35794.488	1.980	199.757	0.949	691003.324	2.001	244.162

L: Lacquer without enhancer

Le: Lacquer with enhancer

AIC: Akaike's Information Criteria

MSE: Mean Square Error

R_{obs}: Correlation coefficient

Z: Zero order release kinetics

F: First order release kinetics

H: Higuchi release kinetics

K: Korsmeyer-peppas release kinetics

MSC: Model Selection Criterion

The Voriconazole flux of the optimized lacquer with an enhancer was found to be 596.75 µg/cm/min at an interval of 48th hour, and provided the permeability coefficient of 58.56 cm/s. Similarly, the flux of lacquer without an enhancer was found to be 540.34 µg/cm/min at the 48th hour interval which gave 53.02 cm/s as the permeability coefficient. Thus, the permeability was found to reduce after 48th hour [fig. 9].

Avulsed human cadaver nails are a suitable model for the permeation studies. No formaldehyde was used in the nail formulation which keeps the formulation in pace with the Campaign for Safe Cosmetics [Safe Cosmetics Action

Network] and free of the toxic trio. Thioglycolic acid was seen to be an effective penetration enhancer for drug delivery through the nail plate. An effective enhancer is one, which can facilitate drug permeation through the keratinized nail membrane, could find application not only in the treatment of nail diseases, but also in the treatment of neighboring target sites, for example in the therapy of rheumatoid arthritis of hands and feet, if systemic circulation is reached. Thus, thioglycolic acid can even act as a periungual drug delivering agent when the target is the blood vessels in the nail bed.

There is no efficient and approved topical formulation containing Voriconazole as an active drug in the market yet for the treatment of onychomycosis. However, a need for topical delivery of Voriconazole exists due to severe adverse effects of the oral therapy. Further, more research is needed to check the nail plate as a membrane in a deeper way. These

insights will be helpful to develop therapies for other diseases through this alternative pathway; for inflammatory and infectitious diseases. Combining all the knowledge, expertise, and technical modalities the vision is that the human nail plate becomes one of the conventional routes for drug delivery.

Figure 9 Permeability coefficient of the optimized nail lacquer formulation 3

A higher Voriconazole concentration and therefore higher concentration gradient has a positive influence on passive diffusion. The type of the pharmaceutical formulation and dosage form can contribute to therapeutic efficacy. The swelling behavior of nails is suggested to be less affected by the formulations with even an increased exposure over 24 hours. The evaluated enhancer, thioglycolic acid; facilitated the Voriconazole's permeation and penetration in to the nail plate. The stability tests showed that the formulations were stable. The optimized formulation was found to follow zero order kinetics and Korsmeyer-Peppas mechanism of drug release with lower mean square values which suggests that the concentration gradient would remain same even on increasing the Voriconazole bedding on the constant surface area. The model selection criterion [MSC], Mean square error [MSE], correlation coefficient [R_{observed}] and Akaike's Information Criterion [AIC] provided permeation profiles for the evaluation of goodness of fit of the model [DDSolver: An Add-In Program for modeling and comparison of drug permeation profiles].

The Voriconazole nail lacquer was also found to be effective in inhibiting the growth of the nail fungi, *Trichophyton rubrum* and gave moderate zones of inhibition conferring for the reapplication of the formulation at intervals for sustaining the certain state of inhibition to cure the fungal infection. Therefore, formulations or treatments, which improve nail hydration, have potential to improve topical therapy for onychomycosis.

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