

Available online on 15.11.2019 at <http://jddtonline.info>

# Journal of Drug Delivery and Therapeutics

Open Access to Pharmaceutical and Medical Research

© 2011-18, publisher and licensee JDDT, This is an Open Access article which permits unrestricted non-commercial use, provided the original work is properly cited



Open Access

Research Article

## Evaluation of isoniazid-oxidative reactions in mice model

Kamel Mokhnache\*, Ahlem Karbab, Soraya Madoui, Hanane Khither, El-Khamsa Soltani, Noureddine Charef

Laboratory of Applied Biochemistry, University Ferhat Abbas Setif 1, 19000, Algeria

### ABSTRACT

In this study the anti-tubercular drug; isoniazid (**INH**) was investigated for their adverse effect; the oxidative stress. This effect was evaluated by using mice model, at the dose of 151 mg/kg. We found that oxidative stress induced by **INH** is associated with lipid peroxidation expressed by the increase in the level of MDA from  $76.9 \pm 1.74$  to  $79.61 \pm 2.67$  nmol/g tissue. The oxidative stress of **INH** is accompanied by a decrease in reduced GSH level (from  $79.9 \pm 12$   $\mu$ mol / mg to  $68.48 \pm 4.28$   $\mu$ mol / mg compared to of the control group). After treatment with **INH** at 151 mg/kg, a decrease in CAT activities occurred compared to control ( $2.53 \pm 0.39$  U/mg Pr vs  $5.07 \pm 0.73$  U/mg Pr).

**Keywords:** isoniazid, oxidative stress, MDA, GSH, CAT

**Article Info:** Received 05 Sep 2019; Review Completed 13 Oct 2019; Accepted 17 Oct 2019; Available online 15 Nov 2019



#### Cite this article as:

Mokhnache K, Karbab A, Madoui S, Khither H, Soltani E, Charef N, Evaluation of isoniazid-oxidative reactions in mice model, Journal of Drug Delivery and Therapeutics. 2019; 9(6):36-37 <http://dx.doi.org/10.22270/jddt.v9i6.3654>

#### \*Address for Correspondence:

Kamel Mokhnache, Laboratory of Applied Biochemistry, University Ferhat Abbas Setif 1, 19000, Algeria

### INTRODUCTION

Isoniazid (**INH**), chemically known as isonicotinic acid hydrazide, is a low-molecular weight and water-soluble compound that can be rapidly absorbed from the gastrointestinal tract. **INH** has been a first-line drug for the treatment of tuberculosis for more than 50 years, with maximum doses for adult of 300 mg. It inhibits bacterial cell wall synthesis, thus killing *Mycobacterium tuberculosis* organisms. Due to its low efficacy towards resistant tuberculosis, it is generally used in combination with other anti-tubercular drugs such as pyrazinamide, ethambutol, and/or rifampicin [1]. In literature, **INH** is one of oxidative drugs. Three principal mechanisms for the oxidative reactions of **INH**; Immune cell products ( $H_2O_2$  and peroxidase) activate **INH** to pro-oxidant hydrazyl, inducing oxidative stress [2]. The damage of lysosomal membrane causes ROS formation via Fenton reaction, which induces oxidative stress and cytotoxicity. The oxidation of **INH** leads to the formation of nitric oxide radical, causing oxidative stress [3,4]. In this study, the oxidative effect of **INH** was evaluated in vivo using mice model, which estimated by lipid peroxidation expressed the level of MDA, the activities of enzymatic antioxidants such as CAT, and non-enzymatic antioxidants such as GSH.

### EXPERIMENTAL SECTION

#### In vivo oxidative stress parameters estimation

In this experiment, **INH** was used as an oxidizing agent that induces oxidative stress in mice. After an 18 h of fasting, **INH**

was injected with intraperitoneal route at 151 mg/kg [5]. The experimental mice were treated as followed:

Group I: control (untreated),

Group II: treated with, **INH** (151 mg/kg)

After 4 h, animals were scarified.

#### Preparation of tissue homogenates

Liver samples were stored at  $-4$  °C until homogenation, then an amount of 500 mg of liver of the different groups was added to 5 mL of the ice KCl buffer (0.15 M). The mixture was homogenized at 1200 rpm using electric homogenizer.

#### Determination of CAT activity

The CAT activity was determined in supernatant, obtained after a 15 min centrifugation of liver homogenates at 4000–5000 rpm. The CAT activities were measured as follows: 30 mM hydrogen peroxide was used as a substrate. To each sample, 1.9 mL phosphate buffer (pH 7.2) and 1 mL 30 mM  $H_2O_2$  were added. The decrease in  $H_2O_2$  concentration at 25 °C was spectrophotometrically determined at 240 nm. The extinction coefficient was read exactly 1 min after  $H_2O_2$  addition. CAT activities presented as units permg protein (Units/mg Protein).

#### Estimation of GSH

The GSH levels were estimated in the homogenates of liver using (5,5'-dithiobis-(2-nitrobenzoic acid), (DTNB), in which 5 mL of sodium phosphate buffer (0.1 M, pH = 8) was added to 25  $\mu$ L of supernatant homogenate, then 1.5 mL of the

mixture was added to 10  $\mu\text{L}$  of DTNB (0.01 M). Determination of GSH is based on the reaction of DTNB with GSH to yield a yellow colored chromophore with a maximum absorbance at 412 nm. The amount of GSH present in the tissue was calculated using its extinction coefficient ( $1.36 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$ ) at 412 nm and expressed as  $\mu\text{moles/g}$  tissue [5].

#### Estimation of MDA

The hepatic malondialdehyde (MDA) content was an indicator to determine hepatic lipid peroxidation levels. In this experiment, 125  $\mu\text{L}$  of trichloric acid (TCA, 20%) and 250  $\mu\text{L}$  of thiobarbituric acid (TBA, 0.67%) were added to 125  $\mu\text{L}$  of tissue homogenate. The mixture was incubated at 100  $^\circ\text{C}$  during 20 min. After incubation, the sample was cooled. The MDA-TBA complex was extracted with 1 mL of butanol. The organic phase was separated by centrifugation at 3000 rpm for 15 min. Absorbance was measured at 530 nm. The concentration of MDA was calculated using its extinction coefficient ( $1.56 \times 10^5 \text{ M}^{-1} \text{ cm}^{-1}$ ) at 530 nm and expressed as nanomoles MDA/g tissue [6].

## RESULTS AND DISCUSSION

Due to the good predicted results and the high free radical-scavenging effect and to the similar structure to indole-derivatives, which are considered potential antioxidant agents [7], the oxidative stress induced by the drug **INH**.

Results of this study indicated that intraperitoneal injections of isoniazid at 151 mg/kg led to an increase in malondialdehyde (MDA) formation, decrease in the activities of enzymatic antioxidants such as CAT, and a decrease in non-enzymatic antioxidants such as GSH when compared with the control.

We found that oxidative stress induced by **INH** is associated with lipid peroxidation expressed by the increase in the level of MDA from  $76.9 \pm 1.74$  to  $79.61 \pm 2.67$  nmol/g tissue. This number was decreased to  $68.02 \pm 2.27$  and  $60.46 \pm 4.63$  nmol/g tissue. The reduced GSH is one of non-enzymatic antioxidant present in the liver which scavenges reactive toxic metabolites of anti-tubercular drugs; liver damage was observed when GSH stocks were significantly decreased [8]. The oxidative stress of **INH** is accompanied by a decrease in reduced GSH level (from  $79.9 \pm 12$   $\mu\text{mol} / \text{mg}$  to  $68.48 \pm 4.28$   $\mu\text{mol} / \text{mg}$  compared to of the control group). Results of the enzymatic antioxidant, CAT activities of treated mice are presented in **Table 1**. After treatment with **INH** at 151 mg/kg, a decrease in CAT activities occurred compared to control ( $2.53 \pm 0.39$  U/mg Pr vs  $5.07 \pm 0.73$  U/mg Pr). The decreased CAT activity could be due to the enhanced generation of ROS during isoniazid metabolism [9] that is deactivated by CAT as well as to the activation of the peroxide oxidation of lipids, proteins, nucleic acids, and other macromolecules.

**Table 1.** Effect of **INH** on oxidative stress parameters.

Groups	CAT (U/mg protein)	MDA (nmol/g tissue)	GSH ( $\mu\text{mol}/\text{mg}$ protein)
Control	$5.07 \pm 0.19$	$76.9 \pm 1.74$	$79.90 \pm 12$
<b>INH 151 (mg/kg)</b>	$2.53 \pm 0.63^{***}$	$79.61 \pm 2.67\text{ns}$	$68.48 \pm 4.28^{***}$

Values expressed as mean  $\pm$  SEM, n= 5 animals /group;\*\*\*:  $p < 0.001$ , \*\*:  $p < 0.01$ , **ns**: no significant differences.

## CONCLUSION

Isoniazid increased the consumption of endogenous antioxidants which could be responsible for the increasing in oxidative stress during their metabolism.

## ABBREVIATIONS

**INH**: isoniazid

**ns**: no significant differences

**MDA**: Malondialdehyde

**GSH**: Gluthation

**CAT**: Catalase

**Pr**: Protein

## REFERENCES

- [1] Koul A, Arnoult E, Lounis N, Guillemont J, Andries K, The challenge of new drug discovery for tuberculosis. *Nature*. 469 (2011) 483–490.
- [2] Tafazoli S, Mashregi M, O'Brien PJ, Role of hydrazine in isoniazid-induced hepatotoxicity in a hepatocyte inflammation model. *Toxicology and applied pharmacology*. 229 (2008) 94–101.
- [3] Rickman KA, Swancutt KL, Mezyk SP, Kiddle JJ, Isoniazid: Radical-induced oxidation and reduction chemistry. *Bioorganic & medicinal chemistry letters*. 23 (2013) 3096–3100.

- [4] Timmins GS, Master S, Rusnak F, Deretic V, Nitric Oxide Generated from Isoniazid Activation by KatG: Source of Nitric Oxide and Activity against Mycobacterium tuberculosis. *Antimicrobial agents and chemotherapy*. 48 (2004) 3006–3009.
- [5] Georgieva N, Gadjeva V, Dimitrova D, Study on the influence of isoniazid alone or combined with new synthesized isoniazid structural analogues upon catalase activity, *Bulgarian journal of veterinary medicine*. 7 (2004) 9–16.
- [6] Aouachria S, Boumerfeg S, Benslama A, Benbacha F, Guemmez T, Khennouf S, Arrar L, Baghiani A. Acute, sub-acute toxicity and antioxidant activities (in vitro and in vivo) of *Reichardia picroide* crude extract. *Journal of Ethnopharmacology*. 208 (2017) 105–116
- [7] Suzen S, Cihaner SS, Coban T, Synthesis and Comparison of Antioxidant Properties of Indole-Based Melatonin Analogue Indole Amino Acid Derivatives, *Chemical biology & drug design*. 79 (2012) 76–83.
- [8] Suresh R, Naik N, Vandana S, Panda M. Hepatoprotective effect of Ginkgo select Phytosome in rifampicin induced liver injury in rats: Evidence of antioxidant activity. *Fitoterapia*. 79 (2008) 439–445.
- [9] Mates JM, Perez-Gomez C, Nunenez de Castro I, Antioxidants enzymes and human diseases, *Clinical Biochemistry*. 32 (1999) 595–603.