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Research Article

Phytochemical and in-vitro antioxidant activity of *Careya arborea* Roxb. leaves successive extracts

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ABSTRACT

The successive extracts of *Careya arborea* Roxb leaves, a traditional medicinal plant used in India. It possessed a significant amount of total phenolic and flavonoid contents with potent antioxidant activities in scavenging DPPH, ABTS radicals, and good total antioxidant capacity. On the other hand, the ethyl acetate and ethanol extracts showed the highest inhibition of DPPH, ABTS radicals and phosphomolybdate assay. This study verified that the ethyl acetate and ethanol extracts have strong antioxidant activities which might be due to high level of phenolics and flavonoids. Thus results confirm its traditional claims can be used as a source of potential antioxidant.

Keywords: *Careya arborea* Roxb, successive, leaves, antioxidant.

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INTRODUCTION

During normal physiological functions, reactive oxygen species (ROS) such as hydrogen peroxide, singlet oxygen, superoxide anion radical, hydroxyl radical are generated from the auto-oxidation of lipids, as well as reactive nitrogen species (RNS) by UV irradiation. These ROS and RNS damages several cellular components such as lipids, proteins, nucleic acid, and DNAs through oxidation or nitration processes results into various degenerative diseases, including cancer, ageing, arteriosclerosis, and rheumatism¹. It has been observed that natural antioxidants are safer than synthetic antioxidants. Therefore, there is an increasing interest amongst scientific communities in identifying natural source of antioxidants derived from plant origin.

Careya arborea Roxb. commonly known as wild guava, is a medium-sized deciduous tree; exhibiting dark grey colour and exfoliating in thin strips. It is widely available in India, Ceylon, Malay and Peninsula. The plant has a variety of traditional uses. The leaves are used for orally in fever while applied locally to relive swelling.² The juice of the leaves is applied in ulcers and skin diseases in India.³ Leaves found to contain triterpenoids and steroids such as taraxerol, n-hexacosanol, α -spinasterol, taraxerol, taraxeryl acetate, 2 α hydroxy ursolic acid, Triterpene ester-careaborin and β -

sitosterol. It also reported to contain tannins and flavonoids and tannins as sitosterol, ellagic acid and quercetin.⁴⁻⁶ The bio-activity guided fractionation of methanol extract of leaves reported to present triterpenoids saponines with good antileishmanial activity. Previous works have shown that *C. arborea* bark and leaves possess antimicrobial activity.⁷ Despite traditional claims, the leaves of *C. arborea* have not been evaluated to antioxidant potential. Thus, the present study was undertaken to evaluate the traditional claim of the leaves of *C. arborea*

MATERIAL AND METHODS

Leaves of *C. arborea* were collected at Mahur-Kinwat region of Nanded district, Maharashtra. The plant was identified by Prof. Vishal R. Marathe, Science College, Nanded, and herbarium voucher specimen was deposited.

The air-dried leaves of *C. arborea* were powdered and exhaustively extracted with different polarity solvent, successively. The petroleum ether (60-80^o) (PE-CA), chloroform (CH-CA), ethyl acetate (EA-CA), and ethanol (EO-CA) extracts were filtered, evaporated under reduced pressure to obtain a viscous dried extracts. The extracts were preliminary investigated for presence of secondary metabolites using standard qualitative chemical tests.

Total Phenolic content

Total phenolic content was estimated using the Folin-Ciocalteu method.⁸ 100 µl of test extracts were mixed thoroughly with 2 ml of 15% Na₂CO₃. After 2 min to this mixture, 200 µl of Folin-Ciocalteu reagent was added and incubated at room temperature for 30 min. The absorbance was measured at 760 nm against a blank. The standard calibration curve was prepared using gallic acid in place of test extract. Total phenolic content was expressed milligram of gallic acid equivalents (GAE) per gram of dried extract. It was calculated by using regression equation.

Total Flavonoids content

The different concentrations (20-100 µg/ml) of standard quercetin solutions (0.5 ml) were separately mixed with 1.5 ml of ethanol, 0.1 ml of 10% aluminium nitrate, 0.1 ml of 1 M sodium acetate and 2.8 ml of water. The resultant mixture was kept at ambient temperature for 40 min. The absorbance of reaction mixture was measured at 415 nm; calibration curve was plotted for concentration against absorbance. Same procedure was followed for the extracts. In the blank solution, the volume of 10% aluminium nitrate was substituted with the same volume of distilled water.⁹ The total flavonoid content in the extract expressed as milligram per gram of quercetin equivalents (QE) with formulae as mentioned for total phenolic content.

Antioxidant activity

DPPH radical-scavenging activity

The DPPH radical-scavenging activity was performed as per Ebrahimzadeh with minor modifications.¹⁰ Different concentrations of test extracts mixed with an equal volume, methanolic DPPH (100mM) solution and added separately in wells of the micro-titre plate. After 20 min incubation at room temperature, the absorbance was measured at 517 nm using microplate spectrophotometer (BIO-Tek, USA. Model-96 well micro plate). Same procedure was followed for control by using methanol in place of extract. The percentage inhibition was estimated based on the percentage of DPPH radical scavenged using the following formula:

$$\% \text{ Inhibition} = \left[\frac{(\text{Control absorbance} - \text{Sample absorbance})}{(\text{Control absorbance})} \right] \times 100$$

ABTS radical cation scavenging activity

ABTS radical cation scavenging activity was performed using the method reported by Fellegrin with slight modifications.¹¹ In brief, ABTS solution (7 mM) was reacted with potassium persulfate (2.45 mM) solution and kept overnight in the dark to yield a dark colored solution containing ABTS^{•+} radical cation. Prior to use in the assay, the ABTS radical cation was diluted with 50% methanol for an initial absorbance of about 0.700 at 734 nm. After the addition of 1.0 ml of diluted ABTS^{•+} to 10 µl of sample, the absorbance was measured after 5 min of initial mixing. The percentage inhibition was calculated according to the formula used for DPPH activity.

The antioxidant potential of extracts was expressed as IC₅₀, the concentration necessary for a 50% reduction of DPPH and ABTS^{•+} radicals.

Total antioxidant capacity by phosphomolybdenum method

The total antioxidant capacity of CAL extracts was determined as ascorbic acid equivalent.¹² The standard curve for total antioxidant capacity was plotted using ascorbic acid standard solution (20-100 µg/ml). An aliquot of 100 µl of extract solutions were combined with 1 ml of reagent (0.6 M sulfuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate). All tubes were capped and incubated in a boiling water bath at 95°C for 90 min. Tubes were allowed to cool at room temperature. Absorbance of the test and standard solutions was measured at 695 nm against blank containing 0.1 ml of distilled water and 1 ml of reagent. An antioxidant capacity was expressed as millimolar equivalents of ascorbic acid.

RESULTS AND DISCUSSION

Plant phenolics or polyphenols are very important secondary metabolites recognised for their potent their antioxidant activity. They act by different mechanism such as chelating redox-active metal ions, inactivating lipid free radical chains and preventing hydroperoxide conversions into reactive oxyradicals. The total phenolic content of successive CAL extracts was expressed as gallic acid equivalents (GAE), varied between 3.64 ± 0.91 mg and 33.03 ± 1.39 mg/g dry weight of fraction. The ethanolic extract exhibited the highest total phenolics content (33.03 ± 1.39 mg gallic acid equivalent/g of extract), followed by ethyl acetate extract.

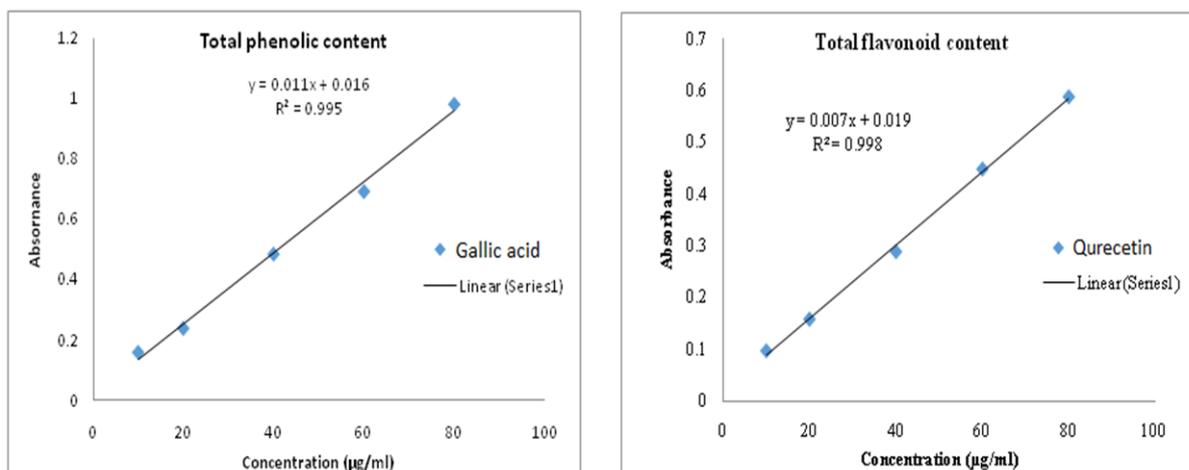


Figure 1: Calibration curve of standard gallic acid and quercetin.

The content of total flavonoids expressed as quercetin equivalents, varied from 3.33 ± 0.82 to 33.33 ± 2.18 mg as quercetin equivalent/g of extract. Phenolic acids and flavonoids have been reported to be the main

phytochemicals responsible for the antioxidant capacity of plant drug. It is well documented that polyphenolics could be a good source of antioxidants and having ability to protect against lipid peroxidation.^{13,14}

Table 1: Total phenolic and flavonoid content of *C. arborea* leaves extracts

Extracts	Total phenolic (mg/g gallic acid)	Total flavonoid (mg/g quercetin)
PE- CAL	3.64 ± 0.91	3.33 ± 0.82
CH-CAL	16.16 ± 1.57	6.67 ± 0.82
EA- CAL	26.36 ± 2.40	33.33 ± 2.18
EO- CAL	33.03 ± 1.39	25.71 ± 1.43

(n=3)

DPPH is a stable free radical at room temperature and accepts an electron or hydrogen radical to become a stable diamagnetic molecule.¹⁵ It has been widely accepted as a tool for estimating free radical scavenging activities of antioxidants.¹⁶ The reduction capability of DPPH was determined by the decrease in its absorbance at 517 nm, which is induced by antioxidants. Positive DPPH test suggests that the samples were free radical scavengers. The scavenging effect of CML extracts and ascorbic acid on DPPH radical was compared. On the DPPH radical, CAL had

significant scavenging effects with increasing concentration in the range of 40–200 $\mu\text{g/ml}$; when compared with that of ascorbic acid. The DPPH activity of CAL extracts were found to increase in dose dependent manner. Ethanol extracts had the highest DPPH radical scavenging activity, shown by the lowest value of IC_{50} values (78.10 $\mu\text{g/ml}$). A higher DPPH radical scavenging activity is associated with a lower IC_{50} value. It was evident that the extracts did show the hydrogen donating ability to act as antioxidants.

Table 2: Antioxidant effect (IC_{50}) on free DPPH radicals of *C. arborea* leaves extracts

Conc. of Sample ($\mu\text{g/ml}$)	PE-CAL	CH-CAL	EA-CAL	EO-CAL	As. Acid	Conc. of Std ($\mu\text{g/ml}$)
40	18.60	31.16	28.84	38.14	20.47	10
80	29.30	40.00	45.58	54.88	26.51	20
120	40.93	53.95	55.35	60.93	36.74	40
160	52.09	56.74	58.60	64.65	51.63	60
200	58.60	60.00	64.65	66.98	62.33	80
IC_{50} ($\mu\text{g/ml}$)	159.26	128.82	117.54	78.10	73.45	

ABTS radical scavenging assay is based on the ability of the antioxidant compound to scavenge the protonated radical cation $\text{ABTS}^{+\cdot}$. This scavenging produces a decrease in the absorbance at 734 nm.¹⁷ The extracts of CAL showed significant dose dependant ABTS radical scavenging activity.

Although the IC_{50} values of the extracts were higher than that of ascorbic acid. Among all ethanol extract of CML extracts showed potent ABTS radicals scavenging ability (68.73 $\mu\text{g/ml}$). The result was shown in Table 3.

Table 3: Antioxidant effect (IC_{50}) on ABTS radicals of *C. arborea* leaves extracts

Conc. of Sample ($\mu\text{g/ml}$)	PE- CAL	CH-CAL	EA- CAL	EO- CAL	As. Acid	Conc. of Std ($\mu\text{g/ml}$)
40	20.80	31.60	39.87	43.20	37.60	10
80	30.13	43.07	46.13	52.80	46.93	20
120	41.07	53.60	54.00	60.40	57.20	40
160	49.47	60.27	56.53	65.20	67.33	60
200	52.80	64.40	58.93	67.07	69.47	80
IC_{50} ($\mu\text{g/ml}$)	173.80	152.69	111.32	68.73	46.52	

The DPPH and ABTS radicals are soluble in water and organic solvent, thus enabling the determination of antioxidant capacity of both hydrophilic and lipophilic compounds.¹⁸ The DPPH and ABTS radical scavenging activity of the EA-CML and EO-CML extract revealed highest antioxidant activity, respectively. The possible reason might be the different contents and sorts of bioactive compounds including phenolics and other compounds responsible for antioxidant capacity. These results were consistent with the findings of many research groups, who reported such correlations between total phenolic content and free radical scavenging activity.^{19,20}

Total antioxidant capacity based on the reduction of Mo(VI) to green colour Mo(V) complex by the antioxidant compounds, which results in the formation of a green coloured Mo(V) complex having a maximum absorbance at 695 nm. A high absorbance value indicated that the sample possesses high antioxidant activity.^{21, 22} The results were expressed as mg/g of ascorbic acid equivalent. The EA-CAL extract showed highest total antioxidant capacity when compared with other FMB extracts (193.11 ± 1.02 mg/g ascorbic acid equivalent) (Table 4).

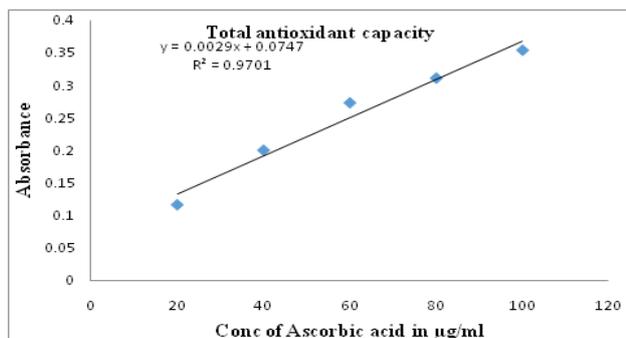


Figure 2: Standard calibration curve of ascorbic acid

Table 4: Antioxidant effect (IC₅₀) on total antioxidant capacity of *C. arborea* leaves extracts

Extracts	Total antioxidant capacity
PE-CAL	1.11 ± 0.77
CH-CAL	4.00 ± 0.67
EA-CAL	14.44 ± 0.77
EO-CAL	10.44 ± 1.02

(n=3), mean ±SD

The results of present antioxidant study of CA leaves extracts established their correlation with polyphenolic content of selected plants, since many studies have proposed that polyphenols are responsible for their antioxidant properties.^{23, 24} The polyphenolic compounds contain hydroxyl groups in their structure and have electron donating ability which allow them to act as reducing agents, hydrogen donors, and singlet oxygen quenchers and responsible for antioxidant property.²⁵ The total polyphenolic contents in EA-CAL and EO-CAL extracts were significantly higher than remaining. Therefore, it can be presumed that the major polyphenolic compounds present in these extracts are responsible for free radical scavenging ability.

CONCLUSION

The results of present work revealed that ethyl acetate and ethanol extract of *C. arborea* leaves have good antioxidant activity. The activity of these extracts is attributed to the phenolic and flavonoid contents. Consequently, our results suggested that the extract can be utilized as an effective and safe and accessible source of natural antioxidants with consequent health benefits.

CONFLICT OF INTEREST

Authors declare, there is no conflict of interest

REFERENCES

- Jeong C-H, Choi GN, Kim JH, Kwak JH, Kim DO, Kim YJ, Heo HJ. Antioxidant activities from the aerial parts of *Platycodon grandiflorum*. *Food chemistry*, 2010; 118:278-282.
- Maheshwari J, Kalakoti B, Lal B. *Ethnomedicine of Bhil tribe of Jhabua District, MP*. *Ancient Science of life*, 1986; 5:255.
- Das P, Misra M. Some ethnomedicinal plants of Koraput district Orissa. *Ancient science of life*, 1988;8:60.
- Mahato S, Banerjee S, Chakravarti R. Taraxerol from *Careya arborea*. *Bulletin of the Calcutta School of Tropical Medicine*, 1967; 15:8-12.
- Gupt R, Chakraborty N, Dutta T. Crystalline constituents from *Careya arborea* Roxb. *Indian Journal Pharmacy* 1975;376:161-162.

- Talapatra B, Basak A, Talapatra S. Terpenoids and related compounds. Part XX. Careaborin, a new triterpene ester from the leaves of *Careya arborea*. *Journal of Indian Chemical Society*, 1981; 58:814-815.
- Kumar RS, Sivakumar T, Sundaram RS, Sivakumar P, Nethaji R, Gupta M. Antimicrobial and antioxidant activities of *Careya arborea* Roxb. Stem bark. *Iranian Journal of Pharmacology & Therapeutics*, 2006; 5:35-41.
- Singleton V, Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American journal of Enology and Viticulture*, 1965; 16:144-158.
- Kalaskar MG, Surana SJ. Free radical scavenging and hepatoprotective potential of *Ficus microcarpa* L. fil. bark extracts. *Journal of natural medicines*, 2011; 65:633-640.
- Ebrahimzadeh MA, Nabavi SM, Nabavi SF, Bahramian F, Bekhradnia AR. Antioxidant and free radical scavenging activity of *H. officinalis* L. var. *angustifolius*, *V. odorata*, *B. hyrcana* and *C. speciosum*. *Pak J Pharm Sci*, 2010; 23:29-34.
- Fellegriani N, Ke R, Yang M, Rice-Evans C. [34] Screening of dietary carotenoids and carotenoid-rich fruit extracts for antioxidant activities applying 2, 2'-azinobis (3-ethylenebenzothiazoline-6-sulfonic acid radical cation decolorization assay. *Methods in enzymology*, 1999; 299:379-389.
- Abdel-Hameed E-SS. Total phenolic contents and free radical scavenging activity of certain Egyptian *Ficus* species leaf samples. *Food Chemistry*, 2009; 114:1271-1277.
- Sharififar F, Dehghn-Nudeh G, Mirtajaldini M. Major flavonoids with antioxidant activity from *Teucrium polium* L. *Food Chemistry*, 2009; 112:885-888.
- Bahramikia S, Ardestani A, Yazdanparast R. Protective effects of four Iranian medicinal plants against free radical-mediated protein oxidation. *Food Chemistry*, 2009; 115:37-42.
- Soare JR, Dinis TC, Cunha AP, Almeida L. Antioxidant activities of some extracts of *Thymus zygis*. *Free radical research*, 1997;26:469-478.
- Sanchez-Moreno C. Methods used to evaluate the free radical scavenging activity in foods and biologic systems-review. *Food Science and Technology International*, 2002; 8:121-137.
- Adedapo AA, Jimoh FO, Koduru S, Afolayan AJ, Masika PJ. Antibacterial and antioxidant properties of the methanol extracts of the leaves and stems of *Calpurnia aurea*. *BMC Complementary and alternative medicine*, 2008; 8:1.
- Magalhães LM, Segundo MA, Reis S, Lima JL. Methodological aspects about in vitro evaluation of antioxidant properties. *Analytica chimica acta*, 2008; 613:1-19.
- Duh P-D, Yen G-C. Antioxidative activity of three herbal water extracts. *Food Chemistry*, 1997; 60:639-645.
- Povichit N, Phrutivorapongkul A, Suttajit M, Chaiyasut C, Leelapornpisid P. Phenolic content and in vitro inhibitory effects on oxidation and protein glycation of some Thai medicinal plants. *Pak J Pharm Sci*, 2010; 23:403-408.
- Prieto P, Pineda M, Aguilar M. Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E. *Analytical biochemistry*, 1999; 269:337-341.
- Pan Y, Wang K, Huang S, Wang H, Mu X, He C, Ji X, Zhang J, Huang F. Antioxidant activity of microwave-assisted extract of longan (*Dimocarpus Longan* Lour.) peel. *Food Chemistry*, 2008;106:1264-1270.
- Tunalier Z, Kosar M, Ozturk N, Baser K, Duman H, Kirimer N. Antioxidant properties and phenolic composition of *Sideritis* species. *Chemistry of natural compounds*, 2004; 40:206-210.
- González-Burgos E, Carretero M, Gómez-Serranillos M. *Sideritis* spp.: uses, chemical composition and pharmacological activities—a review. *Journal of ethnopharmacology*, 2011; 135:209-225.
- Javanmardi J, Stushnoff C, Locke E, Vivanco J. Antioxidant activity and total phenolic content of Iranian *Ocimum* accessions. *Food chemistry*, 2003; 83:547-550.