MICROBIAL TRIGGERED COLON TARGETED COMPRESSION COATED TABLETS OF TENOXICAM: FORMULATION AND EVALUATION

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ABSTRACT
The present investigation deals with to formulation and evaluation of colon specific drug delivery system of Tenoxicam by compression coated tablet technique. Tenoxicam, Avicel pH101, HPMC K4M, Guar gum were used as components. Tenoxicam compression coated tablets were prepared and evaluated for differential scanning colorimetry (DSC), FTIR spectroscopy andin-vitro dissolution study without rat caecal content and with rat caecal content. The results suggest that Colon specific drug delivery system of Tenoxicam compression coated tablets were formulated successfully by using natural polymer such as Guar gum in combination with different concentration of HPMC. Tablets were evaluated for drug content, hardness, thickness, friability, weight variation, in-vitro dissolution and all the results were found within the specification. As the concentration of HPMC in compression coat increased, drug release was decreased. Based on in-vitro drug release study, it was concluded that compression coated tablets with combination of 90% Guar gum and 10% HPMC could produce a successful drug targeting to the colon with minimal amount released in the stomach and small intestine for tenoxicam.

Keywords: Tenoxicam, Guar gum, HPMC K4M, compression coated tablet.

INTRODUCTION
Solid oral dosage forms has been one of the most suitable and widely accepted by the patient for the delivery of therapeutic active drug. The conventional tablet dosage form provides minimal amount of drug in the colon with undesirable adverse effect due to variation in the transit time. Hence, to target the drug directly to the site of action in the colon, there is need to develop colon targeted drug delivery systems that will enhance the therapeutic drug level, increases the bioavailability of active medicament and reduce the dose of drug.

Targeting of drugs to colon is effective and safe for the treatment of various colonic diseases like irritable bowel disease (IBD), intestinal amoebiasis, constipation, Crohn’s disease, ulcerative colitis, carcinomas and infections etc. At present these disease are often poorly & inefficiently managed either by oral route, in which oral drugs are largely absorbed before they rich the colon or by rectal route of administration which is less acceptable. The colon specific drug delivery system (CDDS) should be capable of protecting the drug up to rich the colon i.e., drug release & absorption should not occur in the stomach as well as the small intestine & neither the bioactive agent should be degraded in either of dissolution site but only released & absorbed once the system reaches the colon. Absorption and degradation of the active ingredient in the upper part of GIT is the major problem with the delivery of drugs by the oral route and must be overcome for successful colonic drug delivery.

Time dependent, pH dependent, microbiologically controlled, multiparticulate and luminal pressure controlled system are the latest approaches of drug delivery to the colon. However, a disadvantage of pH dependent system is that a substantial amount of drug may be released in the small intestine before the delivery system arrives in the colon and limitation of time dependent release system is that they are not able to sense any variation in the upper GIT transit time.

Hence, Microbial triggered delivery system is most convenient and highly site specific approach for targeting drug to colon than all targeting system. Drug releases when the polysaccharides are degraded by the bacteria of colonic microflora. Different polysaccharides have been reported for colon targeting as carriers. Most commonly used polysaccharides are guar gum, chitosan, pectin and xanthan gum etc. In compression coated tablets drug present in inner core, surrounded by outer layer which contain polymer and other excipients. Outer coat degraded only in the colonic region by bacterial enzymes and subsequent drug releases in colon.

Tenoxicam chemically 4 –hydroxy –2 – methyl – n-(pyridinyl-2-yl) -2h-thieno [2, 3-e]-1,2-thiazine-3-
carboxamidine1, 1-dioxide, a Non steroidal anti-inflammatory drug. It is used to relieve inflammation, swelling, stiffness, i.e. it is used to relieve pain and inflammation of inflammatory bowel disease, ulcerative colitis, chorn’s disease and rheumatic arthritis. The dose of Tenoxicam is 10-20 mg daily by mouth. Tenoxicam is completely absorbed after oral administration. Tenoxicam has long elimination half life (60-75hr). For colon targeted drug delivery drug has good absorption and solubility profile in colon.

Tenoxicam works by blocking the action of a substance in the body called cyclo-oxygenase (COX). COX is involved in the production of various chemicals in the body, some of which are known as prostaglandins. Prostaglandins are produced by the body in response to injury and certain diseases and conditions and cause pain, swelling and inflammation. Tenoxicam blocks the production of these prostaglandins and is therefore effective at reducing inflammation and pain.

The aim of present investigation was to use the inexpensive, naturally and abundantly available polysaccharide such as guar gum for colon targeted drug delivery. Guar gum is natural non-ionic polysaccharides derived from seeds of Cyamopsis tetragonolobus (family: leguminaceae). It consists of linear chains of (1→4)-β-D-mannopyranosyl units with α-D-galactopyranosyl units attached by (1→6) linkage. Cross linked guar-gum has been used as a drug carrier in matrix tablets. It was concluded that guar-gum is suitable for preparation of colon specific formulations and is suitable as a carrier for drug that are not soluble in water. In this study, Hydroxyl Propyl Methyl Cellulose (HPMC) in combination with Guar gum was used to develop colonic delivery of Tenoxicam. HPMC was used to modify the drug release and improve the mechanical properties such as hardness of compression coated tablets.

The present investigation deals with development of microbial triggered systems for delivering Tenoxicam into the colon and provided “proof of concept” data for natural polysaccharides preparations designed for colonic drug delivery.

**MATERIAL AND METHODS**

Tenoxicam was a gift sample from Ramdev Chemicals, Mumbai, India, Avicel pH101, HPMC K4M and Guar gum etc. procured from Loba Chemicals, Mumbai, India. All other ingredients were of analytical grade.

**Infrared Spectroscopy of drug:**

The drug-excipient interaction study was carried out by using Infrared Spectroscopy. FTIR study was carried out to check compatibility of drugs with excipient. IR spectrum of drugs, physical mixture and optimized formulation were determined on Fourier Transform Infrared spectrophotometer (FTIR, Alpha-E, and Bruker). The base line correction was done then spectrum was run.

**DSC of Pure Drug:**

The drug-polymers compatibility was confirmed by differential scanning calorimetric (DSC), which was carried out by heating drug, and the formulated drug formulations separately from 0°C to 300°C at the heating rate of 10°C/min in a nitrogen environment of flow rate 20.0 ml/min. Aluminium pans and lids were used and temperature calibrations were performed periodically using melting transition of indium as standard. Sample size of 7.5mg used for analysis. The instrument used was SDT Q600 V20.9 Build 101 differential scanning calorimeter with Universal V4.5 software.

**Preparation of Tenoxicam core Tablets:**

The core tablets of Tenoxicam for compression coating were prepared by direct compression technique. Each core tablet (50 mg) consist of 10 mg tenoxicam, 39.50 mg Avicel PH101, and 0.5 mg magnesium stearate. The powders were thoroughly mixed and passed through mesh (60μm). The uniformity of mixing was assessed by conducting content uniformity tests on the samples of powder mix. The mixture was compressed into tablets using KBr press with an applied force of 500 kg/cm² using 4 mm round, Flat punch. The thickness and crushing strength of core tablets were measured.

**Compression coating of Tenoxicam core tablets:**

The coating materials of 150mg Guar Gum in combination with 10%, 20%, and 30% HPMC respectively were used to prepare coats respectively as shown in table 1. Half the amount of compression coating material was placed in the die cavity followed by carefully centering the core tablet and addition of the remaining coat weight. The coating material was then compressed around the core tablets using KBr press at an applied force of 1500 kg/cm² using 8 mm round, flat punch. The prepared tablets were tested for the uniformity of weight, drug content, mechanical properties (hardness and friability) and drug release characteristics. The strength of compression coat tablet was determined.

**Table 1: Composition of Guar Gum and HPMC used to cover Tenoxicam core tablets**

<table>
<thead>
<tr>
<th>Formulation code</th>
<th>Coat weight (mg)</th>
<th>Content (mg)</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Guar Gum</td>
<td>HPMC</td>
</tr>
<tr>
<td>F₁</td>
<td>150</td>
<td>135</td>
<td>15</td>
</tr>
<tr>
<td>F₂</td>
<td>150</td>
<td>120</td>
<td>30</td>
</tr>
<tr>
<td>F₃</td>
<td>150</td>
<td>105</td>
<td>45</td>
</tr>
</tbody>
</table>
Characterization of Tenoxicam compression coated tablets

The prepared tablets were evaluated for weight variation, hardness, friability and drug content.

Swelling study:
Initial diameter, height and weight of individual matrices were measured and were placed in a dissolution medium (Phosphate buffer pH 6.8) at 37±0.5°C. Swollen tablets after specific time intervals (3, 6, 9, 12 and 24) were withdrawn from the medium, extra buffer present on the matrix surface was gently wiped with the soft tissue and individual diameter, height and weight were measured at predetermined time intervals. Percent of the radial (diameter) and axial (height) swelling of tablet and percent water uptake was calculated according to the following formula

\[
\text{Percent swelling (Sw)} = \left(\frac{S_t - S_0}{S_0}\right) \times 100
\]

Where, S1 and S0 are the diameter or height of swollen tablets respectively. The percentage swelling of the original tablet was calculated and plotted vs. Time.

Preparation of rat caecal content:

Committee for the Purpose of Control and Supervision of Experiments of Animals (CPCSEA). The Institutional Animal Ethical Committee (IAEC) of Tatyasaheb College of Pharmacy, Waranagar, Maharashtra, India has approved the experimental protocols for this work (IAEC/TKCP/2014/14).

The caecal contents were obtained from Wistar rats weighing 150-200 g, after pre-treatment with oral administration of 2mL of 1% w/v of polymer dispersion (i.e. Guar gum) in water for 3 days. Thirty minutes before starting drug release studies, each rat was killed by spinal traction, after which abdomens were opened, dissected, and immediately transferred to pH 6.8 phosphate buffer previously bubbled with CO2. The caecal bags were then opened; their contents were individually weighed, homogenized, and then suspended in pH 6.8 phosphate buffer to give the desired concentration of 2% (w/v) of caecal content. As the caecum is naturally anaerobic, all these operations were carried out under CO2.

In vitro drug release studies:
The ability of the prepared tablets to retard drug release in the physiological environment of the stomach and small intestine was assessed by conducting drug release studies in stomach and small intestine pH, respectively. Dissolution test was conducted in USP I apparatus at 100 rpm and a temperature of 37±0.5°C. The tablets were tested for drug release for 2 hours in acidic buffer of pH 1.2 (900 ml), pH 7.4 phosphate buffer for 3 hours and phosphate buffer pH 6.8 for 19 hours with or without rat caecum content. 5ml of each sample was withdrawn at 1, 2, 3, 4, 5, 6, 8, 10, 12, 16, 20, and 24 hrs time and replaced with an equal volume of fresh media. The content of tenoxicam in the withdrawn samples was analyzed spectrophotometrically.

Release kinetics:
Data obtained from the in-vitro release studies of compression coated tablet of Tenoxicam formulations, were fitted to various kinetic equations such as zero order, first order, Higuchi model, Korsmeyer-Peppas model.

Stability Studies:
The optimized formulation was subjected for one month stability study at room temperature. The selected formulations were packed in aluminium foil in tightly closed Container. Formulations were stored at room temperature for one month. After one month interval, placed sample were analyzed for physical appearance, Drug content, in-vitro dissolution study, Hardness, thickness.

RESULTS AND DISCUSSION

Drug–excipients compatibility study:
The overlay of IR spectrum of drug and physical mixture is shown in Figure 1 and table 2, which shows that peaks observed in spectrum of pure drug at wave number 1144 cm\(^{-1}\), 1199 cm\(^{-1}\), 1388 cm\(^{-1}\), 1420 cm\(^{-1}\), 1493 cm\(^{-1}\) were also observed in spectrum of physical mixture. No significant changes in peak pattern in the IR spectra of pure Drug and physical mixture indicates that there is no interaction between pure drug and polymer.

<table>
<thead>
<tr>
<th>Name of Compound</th>
<th>C-O stretching</th>
<th>-O-H bending</th>
<th>CH(_3) deformation</th>
<th>Heterocyclic ring</th>
<th>-C-H deformation</th>
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<tbody>
<tr>
<td>Tenoxicam</td>
<td>1144 cm(^{-1})</td>
<td>1199 cm(^{-1})</td>
<td>1388 cm(^{-1})</td>
<td>1420 cm(^{-1})</td>
<td>1493 cm(^{-1})</td>
</tr>
<tr>
<td>Tenoxicam and HPMC</td>
<td>1142 cm(^{-1})</td>
<td>1199 cm(^{-1})</td>
<td>1385 cm(^{-1})</td>
<td>1420 cm(^{-1})</td>
<td>1492 cm(^{-1})</td>
</tr>
<tr>
<td>Tenoxicam and Guar Gum</td>
<td>1144 cm(^{-1})</td>
<td>1199 cm(^{-1})</td>
<td>1387 cm(^{-1})</td>
<td>1420 cm(^{-1})</td>
<td>1493 cm(^{-1})</td>
</tr>
<tr>
<td>Physical mixture of all ingredients</td>
<td>1442 cm(^{-1})</td>
<td>1190 cm(^{-1})</td>
<td>1387 cm(^{-1})</td>
<td>1418 cm(^{-1})</td>
<td>1491 cm(^{-1})</td>
</tr>
</tbody>
</table>
Figure 1: Overlay of FTIR spectrum of (A) Tenoxicam (B) Physical mixture of Tenoxicam and HPMC K4M (C) Physical mixture of Tenoxicam and Guar Gum (D) Physical mixture of all ingredients.

Figure 2: Overlay of DSC Thermogram of (A) Tenoxicam (B) Optimized formulation F1

Figure 2 shows the DSC thermographs of pure drug Tenoxicam and formulation F1. Thermographs obtained by DSC studies, revealed that the DSC thermogram of the drugs depicts a sharp exothermic peak at 212.5°C reflects the pure crystalline state of the drug and that of the drug in the formulation F1 depicts a exothermic peak at 216.34°C with marked decrease in sharpness. The sharpness of peak in formulation F1 decreased due to low concentration of Tenoxicam drug in formulation i.e. due to dilution of drug. As there is no much difference in the melting point of the drug in the thermograms of drug and that of in the formulation F1. It may be concluded that, the drug was in the same pure state even in the formulation without interacting with the polymers.

Physical properties of tablets:

The hardness of prepared tablets was found to be in the range of 5.30±0.11 to 6.02±0.38 kg/cm², a value which indicates good strength. Thickness of all tablets was found to be in the range of 2.13 ± 0.05 to 2.23±0.5 mm. For weight variation test, the Pharmacopoeial limit for percent deviation for tablets weighing up to 250 mg is not more than 7.5%. The average percent deviation of all tablets was found to be within the limit; hence all formulation passes the weight variation test. The values of friability for all these formulations were less than the limit of 1% which is given in the IP. The friability was found in these formulations shows a good strength of tablets to withstand abrasion during transportation and general handling. Drug content of core tablet was found to be 101.17±0.20 %. As per the IP specification drug content for tablets must be in the range of 85 to 115%. Thus, formulation passes the uniformity of drug content test as shown in table 3.
Table 3: Physical properties of compression coated tablets of Tenoxicam

<table>
<thead>
<tr>
<th>Batch code</th>
<th>Diameter (mm ± SD) a</th>
<th>Thickness (mm ± SD) b</th>
<th>Hardness (kg/cm² ± SD) c</th>
<th>Friability (% ± SD) d</th>
<th>Weight variation (%) e</th>
</tr>
</thead>
<tbody>
<tr>
<td>F₁</td>
<td>8.2±0.23</td>
<td>2.16±0.1</td>
<td>5.3±0.11</td>
<td>0.41±0.12</td>
<td>2</td>
</tr>
<tr>
<td>F₂</td>
<td>8.2±0.17</td>
<td>2.23±0.5</td>
<td>5.5±0.11</td>
<td>0.33±0.5</td>
<td>1</td>
</tr>
<tr>
<td>F₃</td>
<td>8.3±0.32</td>
<td>2.13±0.05</td>
<td>6.02±0.38</td>
<td>0.26±0.15</td>
<td>0.5</td>
</tr>
</tbody>
</table>

a (n=3±SD), b (n=3±SD), c (n=3±SD), d (n=20±SD), e (n=20±SD)

Swelling studies:
Measurement of swelling rates of different matrices were carried out to gain insight into the observed phenomena of drug release with the rates of polymer hydration and to evaluate the extent of water penetration into the tablets. The graphs of water uptake, axial and radial expansion for F₁ are enumerated in Figure 3. It is evident that water uptake was continuously rising throughout the study. This supports the observation that the diffusion was predominant. Figure 3 shows the results of water uptake, axial and radial expansion for formulation F₁. Though water uptake was continuously increasing with time, the radial and axial expansion was almost constant after 12 hr. This suggests the role of erosion to maintain constant diffusional path length because of proper synchronization between erosion and diffusion 29.

In-vitro drug release studies:
The mean drug release from the tablets F₁, F₂, F₃ after the first 5 hr was 8.21±0.15%, 8.63±0.3% and 8.07±0.3% respectively. At the end of 24 hr, the mean % drug release was 40.58±0.12%, 36.13±0.3% and 27.72±0.15%. As the concentration of HPMC increased, drug release was decreased. The increase in drug release could be explained due to HPMC creates a porous structure of the coat and consequently increases Guar Gum leaching and drug released. However further increase in HPMC % to 20% and 30% of the compression coat caused a reduction in gum leaching, with a consequent decrease in drug release as shown in Figure 4. Higher concentration of HPMC would reduce the free water volume and increase the viscosity of the coat causing a reduction in the polymer leaching and subsequent reduction in drug release [25]. Based on these results, F₁ batch optimized for further in-vitro dissolution study using rat caecal content. Since, the % drug release at the end of 5hr which was expected time for the arrival of dosage form in the colon was found to be 8.21±0.15% and 40.58±0.12% drug release was achieved after 24 hr which is higher as compared to F₂ and F₃ batch.
As shown in Figure 5, the mean drug release from the tablet F₁ after the first 5 hr was 7.54±0.5%. At the end of 24 hr, the mean % drug release was 94.54±0.4% in rat caecal medium. The maximum drug release after 24 hr in rat caecal medium was significantly higher in comparison with the drug release in control medium. This can be explained as the release of Tenoxicam in the physiological environment of colon is due to microbial degradation of Guar Gum.

**Drug Release Kinetics**

Table 4 shows optimized batch F₁ followed Koresmeyer-Peppas model indicate significant contribution of diffusion and also showed exponent n value between 0.5 and 1 which indicate non-fickian diffusion and furthermore together with the good fitting of Zero order model indicating erosion controlled release i.e. release was found to be both erosion and diffusion controlled.
Stability Studies:

Short-term stability studies were carried out on the optimized formulation F1. The tablets of optimized batch were stable for 1 month at room temperature. There was no change in the physical appearance and drug content of the tablet during storage period. Mean (± S.D) drug content at the end of 1 month was found to be 99.21±0.5% indicating no significant change in the drug content. The % CDR during the drug release study conducted in the 0.1N HCL, PBS pH 7.4 and PBS pH 6.8 after storage of 1 month was found to be 39.15±0.15% indicating no significant change in the in-vitro dissolution of tablet. The results indicated that, there were no any changes observed in tablet characteristics after stability study.

CONCLUSION

Colon specific drug delivery system of Tenoxicam compression coated tablet was formulated successfully by using natural polymer such as Guar gum in combination with different concentration of HPMC. Based on in-vitro drug release study, it was concluded that compression coated tablets with combination of 90% Guar gum and 10% HPMC could produce a successful drug targeting to the colon with minimal amount released in the stomach and small intestine.

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